

Belgium

TRENDS AND SOURCES OF ZOONOSES AND ZOOTIC AGENTS IN FOODSTUFFS, ANIMALS AND FEEDINGSTUFFS

including information on foodborne outbreaks,
antimicrobial resistance in zoonotic and indicator bacteria
and some pathogenic microbiological agents

IN 2014

PREFACE

This report is submitted to the European Commission in accordance with Article 9 of Council Directive 2003/99/EC*. The information has also been forwarded to the European Food Safety Authority (EFSA).

The report contains information on trends and sources of zoonoses and zoonotic agents in Belgium during the year 2014.

The information covers the occurrence of these diseases and agents in animals, foodstuffs and in some cases also in feedingstuffs. In addition the report includes data on antimicrobial resistance in some zoonotic agents and indicator bacteria as well as information on epidemiological investigations of foodborne outbreaks. Complementary data on susceptible animal populations in the country is also given. The information given covers both zoonoses that are important for the public health in the whole European Union as well as zoonoses, which are relevant on the basis of the national epidemiological situation.

The report describes the monitoring systems in place and the prevention and control strategies applied in the country. For some zoonoses this monitoring is based on legal requirements laid down by the European Union legislation, while for the other zoonoses national approaches are applied.

The report presents the results of the examinations carried out in the reporting year. A national evaluation of the epidemiological situation, with special reference to trends and sources of zoonotic infections, is given. Whenever possible, the relevance of findings in foodstuffs and animals to zoonoses cases in humans is evaluated.

The information covered by this report is used in the annual European Union Summary Reports on zoonoses and antimicrobial resistance that are published each year by EFSA.

* Directive 2003/ 99/ EC of the European Parliament and of the Council of 12 December 2003 on the monitoring of zoonoses and zoonotic agents, amending Decision 90/ 424/ EEC and repealing Council Directive 92/ 117/ EEC, OJ L 325, 17.11.2003, p. 31

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Gallus gallus (fowl) - broilers - Farm (not specified) - Control and eradication programmes - Industry sampling - AMR MON	259
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Pigs - breeding animals - Farm (not specified) - Control and eradication programmes - Industry sampling - OTHER AMR MON	260
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Gallus gallus (fowl) - broilers - Farm (not specified) - Control and eradication programmes - Industry sampling - AMR MON pnl2	283
Gallus gallus (fowl) - broilers - Farm (not specified) - Control and eradication programmes - Industry sampling - AMR MON	283
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Meat, mixed meat - minced meat - Slaughterhouse - Monitoring - Official sampling - AMR MON	288
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1 ANIMAL POPULATIONS

The relevance of the findings on zoonoses and zoonotic agents has to be related to the size and nature of the animal population in the country

1.1.1 Information on susceptible animal population

Sources of information

SANITEL and BELTRACE database of the Federal Agency for the Safety of the Food Chain.

Dates the figures relate to and the content of the figures

Number of animals = number of animals at a certain time point of the year. Number of slaughtered animals = total number of slaughtered animals during the year.

Definitions used for different types of animals, herds, flocks and holdings as well as the types covered by the information

Holding: any establishment, construction or, in the case of an open-air farm, any place in which animals are held, kept or handled. The location of the holding is based on the address and the coordinates of the geographical entity. A geographical entity is a unit of one building or a complex of buildings included grounds and territories where an animal species is or could be held. Herd: an animal or group of animals kept on a holding as an epidemiological unit; if more than one herd is kept on a holding, each of these herds shall form a distinct unit and shall have the same health status.

National evaluation of the numbers of susceptible population and trends in these figures

Over the last years, there's a decrease in total number of holdings of bovines, porcine, sheep, goats and farmed deer. The total number of bovine animals remains unchanged what means that the mean total number of animals per holding is increasing. The total number of holdings of porcine, sheep, goats and farmed deer is also decreasing.

Geographical distribution and size distribution of the herds, flocks and holdings

Belgium can be geographically divided into two regions: the Flemish region situated in the north of the country and the Walloon region situated in the south. There's a very dense animal population of bovines, swine and poultry in the Flemish region. The Walloon region is important for his cattle breeding holdings of the Belgian Blue White race. The number of porcine and poultry holdings in the Walloon region is rather limited.

2 DISEASE STATUS

2.1 TUBERCULOSIS, MYCOBACTERIAL DISEASES

2.1.1 General evaluation of the national situation

2.1.1.1 Mycobacterium - general evaluation

History of the disease and/or infection in the country

Zoonotic tuberculosis (*Mycobacterium bovis*). Tuberculosis in humans caused by *M. bovis* is clinically indistinguishable from tuberculosis caused by *M. tuberculosis*. In the past, the most important way of transmission of *M. bovis* for humans was the consumption of raw milk or raw milk products from infected cattle. Industrial heating production methods or pasteurization of raw milk did stop this way of transmission to humans. Nowadays tuberculosis in humans caused by *M. bovis* is rare. In regions where *M. bovis* infections in cattle are largely eliminated, only few residual cases occur among elderly persons as a result of the reactivation of dormant *M. bovis* within old lesions. Also among migrants from high-prevalence countries, infections with *M. bovis* are diagnosed. Agricultural workers may acquire infection by *M. bovis* by inhaling cough aerosols from infected cattle and may subsequently develop typical pulmonary or genito-urinary tuberculosis. Cervical lymphadenopathy, intestinal lesions, chronic skin tuberculosis (lupus vulgaris) and other non-pulmonary forms are also particularly common as clinical symptoms.

National evaluation of the recent situation, the trends and sources of infection

Recent actions taken to control the zoonoses

The surveillance program of tuberculosis is based on Directive 64/432/EEC, which is implemented and adapted in National legislation since 1963 and last modified by the Royal Decree of 17 October 2002. The control implies skin testing of animals at the occasion of trade and intensive testing of infected and contact farms in consequence of a confirmation of a bovine TB suspicious case (tracing-on and tracing-back of all contact animals). Systematic ante- and post-mortem examination are performed at all slaughterhouses. The Federal Agency for the Safety of the Food chain is informed about any doubtful or positive result of the skin test of bovines and may decide to re-examine (additional tests e.g. comparative tuberculin test, interferon-gamma test) the animals or to kill them for additional analysis (test & slaughter strategy). In case a "TB suspicious" lesion is detected, a tissue sample is sent to the National Reference Laboratory for analysis. Consequently, if *Mycobacterium bovis* suspicion is confirmed by analysis, all animals in the herd of origin are skin tested and an epidemiological investigation is realized. The total herd is considered as the 'epidemiological unit'. Isolation of *M. bovis* and biochemical testing is exclusively performed in the National Reference Laboratory where also IFN-gamma, PCR and molecular typing by means of RFLP, spoligotyping or more recently MIRU-VNTR are done to support the epidemiological investigations and to eventually prove the link between different cases.

Suggestions to the European Union for the actions to be taken

In case a holding is infected and if by epidemiological investigation and tracing-back, animals were found to have been exported to another country, the Chief Veterinary Officer of the country of destination has to be informed about the outbreak in the country of origin. This alert can help to rapidly detect an infection in the concerned holding of destination abroad. Monitoring of the type of strains circulating in each country could contribute to the understanding of the temporal-spatial spread of some specific strains between different countries and could possibly bear some epidemiological links between different outbreaks.

2.1.2 Mycobacterium in animals

2.1.2.1 *M. bovis* in animal - Deer - farmed

Monitoring system

Sampling strategy

Sampling in case of suspicious TB lesions during post-mortem examinations of "wild" and "farmed" deer at slaughterhouse/ at game handling establishment.

Frequency of the sampling

Depends on the number of hunted/slaughtered animals and the detection of suspicious lesions at post-mortem examination.

Type of specimen taken

Suspicious lesions of lungs, lymph nodes, ... at slaughterhouse or game handling establishment.

Methods of sampling (description of sampling techniques)

TB suspicious tissues: lymph nodes, lungs, ...

Case definition

An animal is positive if *Mycobacterium bovis* is isolated by culture or confirmed by laboratory analysis.

Diagnostic/analytical methods used

- Ziehl-Neelsen coloration- Culture for isolation- Interferon-gamma - PCR on lesions / organs- PCR on culture

Control program/mechanisms

The control program/strategies in place

Monitoring is done by:- systematic post-mortem examination at the slaughterhouses/game handling establishment- post-mortem examination at autopsy of hunted or killed "wild" deer by accident in the University Center of Lige, Veterinary Medicine Faculty. In case of suspected TB lesions, tissue samples are sent to the National Reference Laboratory for additional analysis to confirm the suspicion.

Recent actions taken to control the zoonoses

Surveillance program in wildlife.

National evaluation of the recent situation, the trends and sources of infection

No *Mycobacterium bovis* was detected in "wild/hunted" or "farmed" deer for the reporting year.

2.1.2.2 *M. bovis* in animal - Cattle (bovine animals)

Status as officially free of bovine tuberculosis during the reporting year

The entire country free

Belgium is officially free of bovine tuberculosis since the 25th of June 2003 (Commission Decision 2003/467/EC)

Free regions

All regions are officially free of bovine tuberculosis.

Monitoring system

Sampling strategy

Surveillance system. The control of tuberculosis is based on Council Directive 64/432/EEC, which is implemented and adapted in National legislation since 1963 and was last modified by the Royal Decree of 17 October 2002. The surveillance program implies: - skin testing of animals at purchase by the veterinarian responsible for the epidemiological sanitary situation of the holding (contract between farmer and veterinarian); - in case of a suspected/infected bovine(s) on a holding skin testing of all animals of the holding - skin testing of all 'contact' animals and herds (tracing-on and tracing-back); - systematic ante- and post-mortem examination of all slaughtered bovines, transmission to the National Reference Laboratory of all "TB suspicious" lesions for further analysis. Isolation of *M. bovis* and typing is performed at the National Reference Laboratory CODA-CERVA. Also IFN-gamma, PCR and molecular typing by means of RFLP, spoligotyping and more recently MIRU-VNTR are realised at the NRL.

Frequency of the sampling

Frequency of testing depends on: - the introduction of new animals into a herd (mandatory examination at purchase) - the results of tuberculin testing- the detection of suspected bovines- the detection of infected bovines- the epidemiological investigation related to suspected or infected animals or herds (tracing-on and tracing-back)- the follow-up testing of infected and/or eradicated herds during 5 years after partial or total stamping-out.

Type of specimen taken

Organs/tissues: lesions, lymph nodes, lungs Blood

Methods of sampling (description of sampling techniques)

Tuberculin skin testing: single (bovine tuberculin) or comparative (bovine/avian tuberculin) testing. Blood sampling: interferon-gamma tests Laboratory examination of all suspicious lesions by culture: isolation and identification Organs: lymph nodes, lungs, ...

Case definition

- A 'bovine' is defined as infected with bovine tuberculosis if the animal is positive by skin testing or if *Mycobacterium bovis* is isolated by culture or confirmed by laboratory analysis (PCR). - A 'holding' is defined as infected if *Mycobacterium bovis* was isolated from an animal of the holding.

Diagnostic/analytical methods used

- Simple skin test with bovine tuberculin- Comparative skin test with bovine and avian tuberculin- Ziehl-Neelsen coloration- Culture for isolation- Interferon-gamma assay- PCR on lesions / organs- PCR on culture - RFLP typing- Spoligotyping- MIRU-VNTR

Vaccination policy

Vaccination is prohibited by Royal Decree of 17 October 2002.

Control program/mechanisms

The control program/strategies in place

National surveillance program by the Competent Authority (FASFC) on mandatory legal base.

Recent actions taken to control the zoonoses

Draw special attention and focus on the post-mortem examination of slaughtered animals; Transmission for further analysis of any lesion that could be 'suspected' of tuberculosis to the National Reference Laboratory; Culture of *M. bovis*, biochemical testing, PCR are performed on these 'suspicious' lesions; Molecular typing by means of RFLP, Spoligotyping and more recently MIRU-VNTR are realised on all isolates to support the epidemiological investigations and to eventually prove the link between different cases or outbreaks.

Suggestions to the European Union for the actions to be taken

In case of export of bovines, inform the Chief Veterinary Officer of the Member state of destination if tuberculosis has been detected in a holding of the Member State of origin after the date of export. This information can result in an early detection or can avoid a possible further contamination in the Member State of destination.

Measures in case of the positive findings or single cases

If *M. bovis* is suspected, all animals in the herd of origin are skin tested, the herd is considered as the epidemiological unit. A complete epidemiological investigation is performed. After tracing-back and tracing-on all animals of 'contact' holdings are examined by skin testing. If any doubtful or positive result of the skin test is detected, the FASFC may decide to re-examine the animals (additional tests e.g. comparative skin testing with avian and bovine tuberculin and/or Interferon-gamma testing) or to a direct slaughter of the reactors (test slaughter) for additional analysis. In case a suspicious lesion is detected at post-mortem examination, a sample is sent to the National reference laboratory for analysis. If in consequence *Mycobacterium bovis* is isolated, all skin test positive animals during successive testing are mandatory slaughtered. If many bovines are reacting positive to skin testing, the FASFC can decide that all animals of the holding must be mandatory slaughtered (total stamping-out). In most breakdowns a sanitation plan is established taking into account the epidemiological situation. After stamping-out, new restocked animals are tested during 5 consecutive years by annual skin testing to prove the TB free status of the holding.

Notification system in place

Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of all notifiable animal diseases).

Results of the investigation

In 2001, a total of 23 infected holdings were notified. In total 792 animals reacted after tuberculinisation. In 2002, a total of 13 infected holdings were notified. A total of 799 animals reacted after tuberculinisation. Stamping-out was performed in 6 herds. In 2003, a total of 7 infected holdings were notified. Stamping out was done in 5 herds. A total of 409 animals reacted after tuberculinisation. This number corresponds to the intensive testing of infected and contact farms. In total 3.799 herds and 337.260 animals were included in epidemiological investigations. The Federal Agency for the Safety of the Food Chain, the Competent Authority, instructed the slaughter of 1014 animals. In 2004, a total of 8 infected holdings were detected. In total 229 bovines were slaughtered in consequence of the stamping-out of 3 infected herds. In 2005, a total of 5 infected holdings were detected. All these herds were eradicated by stamping-out in execution of a TB sanitation plan. In total 752 animals were slaughtered. The carcasses of only 2 animals did have to be destroyed due to general dispersed TB lesions. In 2006, a total of 8 infected holdings were detected. Seven of these were eradicated by stamping out. In total 1102 animals were slaughtered. A follow-up of the other infected holding is performed after test-slaughter of a few positive reactors, since then all results of tuberculin tests on all the animals of the herd at regular intervals are negative. In 2007, a total of 5 infected holdings were detected. Three of these were eradicated by stamping-out. In total 487 animals were slaughtered. In the other two infected holdings, partial slaughter and intense follow-up by tuberculin testing was performed. In 2008, a total of 12 infected holdings were detected. In total 812 animals were slaughtered. Finally 66 animals were detected positive in bacteriological examination. In 2009, 2 infected holdings were detected. One holding was eradicated by stamping-out. On the other holding, partial slaughter and intense follow-up by tuberculin testing was performed. In 2010 no infected holding was detected. In 2011, 1 infected holding was discovered. All animals were slaughtered. In December 2012, 1 infected holding was detected. All animals of the holding were slaughtered. In consequence 148 'contact' herds were followed-up by tuberculin testing in 2012 and 2013. In 2013, 9 infected holdings were detected. 5 holdings were eradicated by total stamping-out. 4 holdings had a partial slaughter of all tuberculin reacting animals and will be further followed up by tuberculin testing. In total 1012 contact herds had to be followed-up by tuberculin testing due to these 9 breakdown herds. In 2014, a follow-up tuberculinisation of the outbreak herd TUB_2013_09 were partial stamping-out was applied in 2013 resulted again in some reactors. Total stamping-out was decided and at slaughter one bovine with suspicious lesions was positive by bacteriological examination.

National evaluation of the recent situation, the trends and sources of infection

Number of infected herds since 2000: 2000 : 242001 : 232002 : 132003 : 72004 : 82005 : 52006 : 82007 : 52008 : 122009 : 22010 : 02011 : 12012 : 12013 : 92014 : 0

Additional information

2.2 BRUCELLOSIS

2.2.1 Brucella in animals

2.2.1.1 B. suis in animal

Monitoring system

Sampling strategy

Serological screening for Brucella is done for breeding pigs that are gathered (at a fair for example), at artificial insemination centers and in animals intended for trade. The methods used are Rose Bengal test (RBT), Slow Agglutination test (SAT) according to Wright, Complement Fixation test (CFT) and ELISA. Bacteriological examination for Brucella and Yersinia is done in case of positive serology. Regularly, false positive serological reactions are reported. These are due to a Yersinia enterocolitica O9 infection and are confirmed by Yersinia enterocolitica O9 isolation in the absence of Brucella spp. isolation. B. suis biovar 2 may be isolated from wild boars (Sus scrofa). The infection seems to be endemic in wild boar in Belgium. B. suis biovar 2, circulating among wild boars, shows only limited pathogenicity for humans, if pathogenic at all. The domestic pig population is free of brucellosis (last Brucella isolation in domestic pigs in Belgium was in 1969).

Methods of sampling (description of sampling techniques)

Blood sampling Tonsils Spleen

Case definition

An animal is positive if Brucella suis is isolated by culture or typed by additional laboratory methods.

Diagnostic/analytical methods used

Rose Bengal test RBT Complement fixation test CFT Indirect ELISA Bacteriological examination

Control program/mechanisms

The control program/strategies in place

National evaluation of the recent situation, the trends and sources of infection

2.2.1.2 B. abortus in animal - Cattle (bovine animals)

Status as officially free of bovine brucellosis during the reporting year

The entire country free

Belgium is officially free from bovine brucellosis since the 25th of June 2003 (Commission Decision 2003/467/EC)

Free regions

Belgium remained officially free of bovine brucellosis during this reporting year.

Additional information

End 2010 a brucellosis breakdown herd was detected after analyzing an abortion. The infected herd was totally depopulated. Extensive epidemiological investigations and important follow-up by serology of contact herds in 2010 and 2011 could not give any indication on the origin of the infection neither could be detected any additional other infected herd. In March 2012, again a breakdown of brucellosis was detected after analysis of an abortion. No epidemiological link could be found with the breakdown of 2010. Tracing-back and an epidemiological inquiry lead to the detection of 4 other secondary breakdowns linked to the primary case. All these 5 brucellosis breakdown herds were infected with an identical Brucella abortus biovar 3. Another infected herd of brucellosis was detected by analysis of tankmilk and an infection with Brucella suis biovar 2 was confirmed. This breakdown could be considered as an isolated case. This biotype is endemic in Belgian wildboar population. Also bovines are susceptible to this suis biovar. Finally there was a stamping-out of all the animals of the infected herds. In 2013 a breakdown herd was detected as contact herd of the primary breakdown herd of 2012. Both herds had the same veterinarian. The breakdown herd of 2013 was already examined twice by serology in 2012 with negative results. A third follow-up screening by serology indicated some positive results. This positive serology could be confirmed by culture after test-slaughter of the reactors. Finally 6 bovines were infected. There was a stamping-out of all the animals of this infected herd. 67 contact herds of this breakdown herd of 2013 had to be screened by serology and will be followed-up later on. No other infected herd could further be detected. In 2014, bovine brucellosis was not detected.

Monitoring system

Sampling strategy

Since Belgium is officially free of bovine brucellosis, the eradication program has been changed in a surveillance program. Beef cattle older than 2 years were monitored once every three years by means of serological tests. The herds for serological sampling and examination were selected by their geographical location. Dairy cattle were checked at least 4 times a year via tank milk (milk ring test). Furthermore, all animals were tested at trade (purchase) on the herd of arrival. Each abortion or premature birth in animals at risk must be subject to compulsory notification to the Federal Agency for the Safety of the Food Chain, and be tested for brucellosis. Aborting females should be kept in isolation until the results of the analysis and the investigation exclude a *Brucella* infection. Pooled tank milk was examined by means of a milk ring test. For animals older than 2 years of dairy herds, serology (i.e. micro-agglutination as screening test; in case of a positive result, an indirect ELISA test is performed) is used if no sufficient milk ring tests were performed (at least 4 tests a year). Bacteriological examination is done when serological and/or epidemiological suspicion is present. An animal is legally suspected of brucellosis in case of a positive ELISA. If, according to the epidemiology and the results of the blood test, an animal or herd is found to be at risk, a bacteriological investigation always takes place. Hence, a brucellosis animal is defined as an animal in which *Brucella abortus* has been isolated, and a cattle holding is considered as an outbreak herd if one of the animals is positive for brucellosis by bacteriological examination. In 2009, a study was realized to evaluate the current national surveillance program of bovine brucellosis. If a Member State has maintained the officially free status of brucellosis for at least 5 consecutive years, the existing surveillance program can be re-evaluated and some modifications on the sampling design are allowed on condition of further proof of freedom of disease (Council Directive 64/432/EEC). The scientific veterinary experts used risk-based models to evaluate different scenarios within the current surveillance program and the study was also based on a statistical confidence level approach. This methodology has underlined a few important features of the current brucellosis surveillance program. The study showed that in order to obtain a 99% confidence level to prove freedom of disease consistently an important decrease in total number of tested animals can be proposed (500.000 to 30.000 tests a year). The study also clearly indicated that the best approach is to test bovines imported from officially free or non-officially free Member States of *Brucella* spp., to test animals at purchase in consequence of national trade as well as to analyze aborting animals in order to early detect infection. Regarding the passive surveillance (abortions), the study indicated there is a need to increase the number of analyzed abortions. Also the mandatory analysis for brucellosis at purchase of new animals changed into a voluntary approach. A new surveillance program has been applied from the end of 2009 on. In 2012 surveillance was focused on following risk categories: - import of non officially free MSs or Third Countries at the moment of trade and follow-up testing during 3 consecutive years during the winterscreening (targeted selection)- at random selection of 450 bovine herds for serological investigation of 40 animals per herd divided in 4 different age categories: 10 animals of 6-12 months of age, 10 animals of 12-24 months of age and 20 animals older than 24 months. - number of analysis of bovines of national trade at purchase- at random selection of 750 bovine herds of all herds that did not declare any abortion during the passed year. On these herds a maximum of 20 animals are randomly selected for serological analysis of brucellosis. - due to the brucellosis ou

Frequency of the sampling

- import of non officially free MSs or Third Countries at the moment of trade: all imported animals over 12 months of age- import of non officially free MSs or Third Countries follow-up testing during winterscreening for 3 consecutive years of all imported animals over 24 months of age- at random selection of 450 bovine herds: at random selection of maximum 40 female animals in different age categories- bovines of national trade at purchase: at random selection, limited number of analysis - at random selection of 750 bovine herds where no abortion was declared/analyzed during the last year, at random selection of 20 female animals (goal is to stimulate the notification of abortions)- abortion protocol: examination of abortions for brucellosis and some other diseases which can induce an abortion in bovine animals (IBR, BVD, Neoplasiose, ...).

Type of specimen taken

BloodTankmilk

Methods of sampling (description of sampling techniques)

Blood sampling

Case definition

An animal is defined as infected if *Brucella* spp. has been isolated by culture and identified. A herd is defined as infected if one of its animals is positive by bacteriological examination for Brucellosis.

Diagnostic/analytical methods used

- Micro agglutination test - ELISA on blood or tank milk - Complement Fixation Test- Rose Bengale Test- PCR- Stamp/Ziehl Neelsen coloration- Culture

Vaccination policy

Vaccination is prohibited in Belgium since 1992.

Control program/mechanisms

The control program/strategies in place

National mandatory surveillance program organized by the FASFC.

Recent actions taken to control the zoonoses

Measures in case of the positive findings or single cases

In case of a positive result in the micro-agglutination test the same blood sample is tested with an ELISA. If this indirect ELISA is positive, this result has to be confirmed by a blocking ELISA at the NRL. If this confirmatory test is positive, the animal is considered as infected and is compulsory slaughtered (test slaughter) for additional analysis to detect a possible Brucella infection by culture.

Notification system in place

Animal Health Law of 24 March 1987 Chapter III, Royal Degree of 3 February 2014 (list of notifiable diseases)

National evaluation of the recent situation, the trends and sources of infection

An intensified bovine brucellosis control program started in Belgium in 1988. In case of active brucellosis, i.e. excretion of Brucella, the plan consisted in the culling of all animals of the infected herd (total depopulation). Culled bovines were compensated for based on the replacement value of the animals. In March 2000, the last case of bovine brucellosis was identified before obtaining the officially brucellosis free status in 2003. In case of positive serological reactors the Federal Agency for the Safety of the Food Chain instruct follow-up testing or 'test slaughter' for additional analyses. These analyses could not confirm brucellosis. To reduce the number of FPSR (False positive serological reactors) to be slaughtered, the micro-agglutination test has been used as for routine testing whereas the indirect Elisa is accepted as a complementary test by serial or parallel testing. The blocking ELISA of the NRL is considered as the confirmation test. This approach avoids the undeserved test slaughter of false positive reacting animals. In March 2012 a breakdown of bovine brucellosis was detected at a herd in the province of Namur. Bovine brucellosis was detected by analysis of an abortion and serology. Serological examination of the cow and bacteriological examination of the fetus indicated a Brucella infection that was confirmed and typed as Brucella abortus biovar 3. Extensive epidemiological investigation designated 291 contact herds for follow-up by serology. Serological analysis of all contact herds detected another 4 breakdowns of Brucella abortus biovar 3. After test & slaughter of 118 animals of the breakdown herds, bacteriological examination was positive for 11 animals. To follow-up this Brucellosis incidence, 3 rounds of blood sampling took place in 2012. Respectively 538, 455 and 176 holdings and 40.780, 30.407 and 438 animals were sampled where 39, 5 and 0 blood samples were positive by a confirmatory ELISA. In consequence, 123 bacteriological examinations took place after test & slaughter of the animals, only 1 culture was positive and finally typed as Brucella suis biovar 2. In addition to the serological follow-up of these contact herds by blood sampling, all Belgian dairy herds were tested three times by an ELISA of tank milk. During these 3 rounds of surveillance by tankmilk, respectively 8656, 8634 and 8497 herds were sampled and 23, 28 and 20 tank milk samples gave a non conform result. The dairy herds were blood sampled and finally only one bovine had to be mandatory slaughtered for examination by culture. Brucella suis biovar 3 was isolated from this animal. In January 2013 a new breakdown herd of bovine brucellosis was detected. This infected herd could be linked to the first breakdown herd of 2012. Both herds had the same veterinary surgeon. This breakdown herd was already examined twice by serology in 2012 but was only found positive by a third screening by serology in January 2013. 67 contact herds were followed up and 12.901 additional blood samples were analyzed. Finally no other infected herd or animal could be found. Also the screening of tankmilk of all dairy herds took place twice in 2013. In 2014 no infected animals were detected.

Additional information

2.2.1.3 B. melitensis in animal - Goats

Status as officially free of caprine brucellosis during the reporting year

The entire country free

Belgium is officially free of B. melitensis since 29 March 2001 (Commission Decision 2001/292/EC).

Free regions

Belgium is officially free of caprine brucellosis during the reporting year.

Monitoring system

Sampling strategy

Serum samples taken in the framework of a national monitoring program for Maedi-Visna/CAE and at export were examined for *Brucella melitensis* specific antibodies by means of an ELISA. Sheep and goats were tested for brucellosis by indirect ELISA (iELISA) at the NRL CODA-CERVA. All positive samples in the ELISA were supplementary tested by the Rose Bengal Test (RBT) and Complement Fixation Test (CFT) as confirmatory tests. Animals that were positive in the two confirmatory tests or that could not be analyzed and/or interpreted in RBT and/or CFT were sampled a second time.

Type of specimen taken

Blood

Methods of sampling (description of sampling techniques)

Blood sampling

Case definition

A goat is defined as infected with brucellosis if positive in all three tests: iELISA, Rose Bengal test and Complement Fixation test and isolation of *Brucella melitensis* by culture after test slaughter.

Diagnostic/analytical methods used

Complement Fixation Test CFT
Rose Bengal Test RBT
Indirect ELISA
Culture for isolation

Notification system in place

Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of notifiable animal diseases)

Results of the investigation

At the NRL, 6.561 caprine/ovine serum samples were tested. The results confirmed those of previous years, i.e. the absence of any epidemiological or bacteriological evidence of caprine/ovine brucellosis in Belgium.

2.2.1.4 B. melitensis in animal - Sheep

Status as officially free of ovine brucellosis during the reporting year

The entire country free

Belgium is officially free from *B. melitensis* since 29 March 2001 (Commission Decision 2001/292/EC).

Free regions

Belgium is officially free of ovine brucellosis during the reporting year.

Monitoring system

Sampling strategy

Serum samples taken in the framework of a national monitoring program for Visna-Maedi/CAE and at export were examined for *Brucella melitensis* specific antibodies by means of an iELISA. Positive samples were subsequently tested in Rose Bengal and in complement fixation test. Sheep and goats sera were tested for brucellosis by indirect ELISA (iELISA) at the NRL. All positive samples in the ELISA were then tested by the Rose Bengal Test (RBT) and Complement Fixation Test (CFT) as confirmatory tests. Animals that were positive in the two confirmatory tests or that could not be analyzed and/or interpreted in RBT and/or CFT were sampled a second time.

Type of specimen taken

Blood

Case definition

A sheep is defined as infected with brucellosis if positive in all three tests: the Elisa, the Rose Bengal test and the Complement Fixation test and isolation of *Brucella melitensis* by culture.

Diagnostic/analytical methods used

- Indirect ELISA- Rose Bengal Test RBT- Complement Fixation Test CFT- Culture for isolation- Brucellin skin test (BST)

Notification system in place

Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of notifiable animal diseases).

Results of the investigation

At the National Reference Laboratory, 6.561 caprine/ovine serum samples were tested. The results confirmed those of previous years, i.e. the absence of any epidemiological or bacteriological evidence of caprine/ovine brucellosis in Belgium.

3 INFORMATION ON SPECIFIC ZONOSSES AND ZONOTIC AGENTS

Zoonoses are diseases or infections, which are naturally transmissible directly or indirectly between animals and humans. Foodstuffs serve often as vehicles of zoonotic infections. Zoonotic agents cover viruses, bacteria, fungi, parasites or other biological entities that are likely to cause zoonoses.

3.1 SALMONELLOSIS

3.1.1 Salmonella in foodstuffs

3.1.1.1 Salmonella spp. in food - Meat from bovine animals

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

At meat processing plant

A monitoring program was organized at meat processing plants by the FASFC.

At retail

A monitoring program was organized at at retail by the FASFC.

Frequency of the sampling

At meat processing plant

Sampling distributed evenly throughout the year

At retail

Sampling distributed evenly throughout the year

Type of specimen taken

At meat processing plant

Minced meat, sausages and other

At retail

Meat, minced meat, pate, sausages, meat salads and other

Methods of sampling (description of sampling techniques)

At meat processing plant

The samples were more than 200 g of meat. The detection of Salmonella has been assessed in 10g or 25g of sample.

At retail

The presence of Salmonella has been assessed in 10g or 25g of sample.

Definition of positive finding

At slaughterhouse and cutting plant

A sample is considered positive in case of detection of Salmonella in the sample.

At meat processing plant

A sample is considered positive in case of detection of Salmonella in the sample.

At retail

A sample is considered positive in case of detection of Salmonella in the sample.

Diagnostic/analytical methods used

National evaluation of the recent situation, the trends and sources of infection

3.1.1.2 Salmonella spp. in food - Meat from broilers (Gallus gallus)

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

A monitoring program in Belgian slaughterhouses and cutting plants was organized by the FASFC. The matrices were carcasses, cuts and meat preparation of broilers. The carcass samples of broiler consisted of 10g of neck skin. The following contamination levels were analyzed: 25g cutting meat and 10g of minced meat of chicken and 1g of chicken carcasses. Sampling was done by a specially trained staff. For most matrices, independent samples were taken per matrix in order to detect a minimal contamination rate of 1% with 95% confidence.

At retail

An annual control program is designed following the strategy as explained in the MANCP.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year

At meat processing plant

Sampling distributed evenly throughout the year

At retail

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughterhouse and cutting plant

Neck skin and cutting meat

At meat processing plant

Minced meat, sausages, meat and other

At retail

Minced meat, sausages, meat and other

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

The matrices were carcasses, cuts and meat preparation of broilers. The carcass samples of broiler consisted of 10g of neck skin. The following contamination levels were analyzed: 25g cutting meat and 10g of minced meat of chicken and 1g of chicken carcasses.

At meat processing plant

The samples were about 200 g of meat. The detection of Salmonella has been assessed in 10g or 25g of sample.

At retail

The presence of Salmonella has been assessed in 25g of sample.

Definition of positive finding

At slaughterhouse and cutting plant

A sample is considered positive in case of detection of Salmonella in the sample.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: ISO 6579:2002

At retail

Control program/mechanisms

The control program/strategies in place

A microbiological control of carcasses and meat of poultry is made with the aim of following the level of contamination by Salmonella.

Measures in case of the positive findings or single cases

In case of positive findings, no measure is taken face to products which entered normally the food chain except for *S. enteritidis/typhimurium*. But corrective measures must be taken at the level of the slaughterhouse or of the cutting plant by the FBO.

National evaluation of the recent situation, the trends and sources of infection

The rate of Salmonella contamination of poultry meat observed in 2013 is comparable with the previous years.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

3.1.1.3 Salmonella spp. in food - Meat from pig

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

A monitoring program was organized by the FASFC in slaughterhouses and cutting plants. Sampling was done by a specially trained staff. For most matrices, independent samples were taken per matrix in order to evaluate the contamination with 95% confidence.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year

At meat processing plant

Sampling distributed evenly throughout the year

At retail

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughterhouse and cutting plant

Swabs of carcass

At meat processing plant

Minced meat, ham, sausages and other

At retail

Meat, minced meat, ham, pate, sausages, meat salads and other

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

The matrices were carcasses, cuts and minced meat of pork. Sampling of pork carcasses was done by means of swabs. The following contamination levels were analyzed: 10 g or 25g (cutting, minced meat of pork) and 600 cm² (pork carcasses).

At meat processing plant

The samples were more than 200 g of meat. The detection of Salmonella has been assessed in 10g or 25g of sample.

At retail

The presence of Salmonella has been assessed in 10g or 25g of sample.

Definition of positive finding

At slaughterhouse and cutting plant

A sample is considered positive in case of detection of Salmonella in the sample.

At meat processing plant

A sample is considered positive in case of detection of Salmonella in the sample.

At retail

A sample is considered positive in case of detection of Salmonella in the sample.

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: ISO 6579:2002

National evaluation of the recent situation, the trends and sources of infection

The rates of salmonella contamination of carcasses and cutting meat of pig estimated in 2013 were statistically similar to 2012.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

The main serotype found on Salmonella risk farms (fattening pigs), on carcasses and in pig meat is Salmonella Typhimurium.

3.1.1.4 Salmonella spp. in food

Monitoring system

Sampling strategy

A monitoring program was organized by the Federal Agency for the Safety of the Food Chain. More than 200 Belgian slaughterhouses, more than 100 meat cutting plants and more than 100 retail trades representative of the Belgian production, were selected for this study. The samples assayed were carcasses, cuts and minced meat from pork, carcasses, cuts and meat preparation from chicken, layer carcasses, beef minced meat and other foodstuffs. Sampling was done by a specially trained staff of the Federal Agency for the Safety of the Food Chain. For most of the matrices, approximately 100 - 300 independent samples were taken per matrix in order to detect a minimal contamination rate of 1% with 95% confidence. Salmonella isolates were serotyped and serotypes Typhimurium, Enteritidis, Virchow and Hadar were lysotyped. The antibiotic resistance profiles were determined for all isolates, and included ceftriaxone, ampicillin, kanamycin, sulfamethoxazole, tetracycline, nalidixic acid, ciprofloxacin, chloramphenicol and trimethoprim.

Frequency of the sampling

Meat samples have been taken every week from the first to the 52nd week. Samples are taken according to the national control program or in the frame of RASFF, complaints or suspicion.

Type of specimen taken

Meat, milk and dairy products and other foods such as eggs, fishery products, ...

Methods of sampling (description of sampling techniques)

Sampling of pork carcasses was done by means of swabs. The carcass samples of broiler and layer consisted of 10g of neck skin. The other samples were about 200g of meat. The detection of Salmonella has been assessed in these dilutions: 25g (cutting and minced meat of pork, chicken cuts and beef), 600 cm² (pork carcasses), and 1g (chicken and layer carcasses, chicken meat preparation).

Definition of positive finding

A sample is considered to be positive after biochemical confirmation of one Salmonella spp. in the sample.

Diagnostic/analytical methods used

Five laboratories licensed by the Federal Agency for the Safety of the Food Chain and accredited following ISO 17025 standard analyzed all the samples. The Belgian official method SP-VG-M002 was used for the detection of Salmonella in 25g, 1g or on swabs: - pre-enrichment in buffered peptone water at 37C for 16 to 20 h, - selective enrichment on the semi-solid Diassalm medium at 42C for 24 h, - isolation of positive colonies on XLD at 37C for 24 h, - confirmation of minimum 2 colonies on TSI at 37C and miniaturised biochemical tests, - serotyping and lysotyping were done at the National Reference Center for Salmonella and Shigella (NRCSS-IPH) and at the Institute Pasteur, both located in Brussels, respectively. - antibiotic resistance determination by IPH Brussels by disk diffusion method.

Preventive measures in place

Controls are made in place by the Federal Agency in case of notification.

Control program/mechanisms

The control program/strategies in place

Notification is mandatory since 1/3/2004 (Ministerial Decree on mandatory notification in the food chain of 22/1/2004). For Salmonella, absence in 25g in ready-to-eat food putted on the market is mandatory. Laboratories have to inform the Federal Agency in case of a positive sample.

Measures in case of the positive findings or single cases

Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch or single sample- Further investigation: additional sampling, possible recall, RASFF, ...

3.1.2 Salmonella in animals

3.1.2.1 Salmonella spp. in animal - Cattle (bovine animals)

Monitoring system

Sampling strategy

There was no official monitoring program for cattle in 2014. Given results were from isolates from diagnostic samples sent to the NRL Salmonella, animal health, for serotyping.

Vaccination policy

In 2014, no vaccine was authorized for the vaccination of cattle against salmonellosis.

Results of the investigation

Results from the NRL Salmonella, AH indicate that the number of Salmonella isolates from cattle (n=43) remains at the same level of 2012 (n=42). Most frequently found serotypes are Typhimurium (26) and Dublin (7). Some monophasic strains were also detected by PCR.

National evaluation of the recent situation, the trends and sources of infection

Data from the NRL Salmonella, AH show that in cattle, S. Dublin used to be the principal serotype between 2002 and 2010, but declined in 2010 and 2011 to the same low level as S. Typhimurium. In 2013 S. Typhimurium is clearly the most prevalent isolated serotype from cattle samples

3.1.2.2 Salmonella spp. in animal - Gallus gallus (fowl) - broilers

Monitoring system

Sampling strategy

Broiler flocks

The official surveillance program for broilers in accordance with Regulations (EC) Nos 2160/2003 and 200/2012 started in 2009. It is compulsory to sample all flocks on farms with a capacity of 200 or more birds as day-old chicks and in the last three weeks before slaughter.

Frequency of the sampling

Broiler flocks: Day-old chicks

Each 'batch' of day-old chicks that enters the farm must be sampled in the hatchery or when arriving on the farm.

Broiler flocks: Before slaughter at farm

Every flock is sampled in the last 3 weeks before slaughter.

Broiler flocks: At slaughter (flock based approach)

Sampling of caeca at slaughter is distributed evenly throughout the year

Type of specimen taken

Broiler flocks: Day-old chicks

Internal linings of delivery boxes or hatcher basket liners

Broiler flocks: Before slaughter at farm

Socks/ boot swabs

Broiler flocks: At slaughter (flock based approach)

Organs: caeca

Methods of sampling (description of sampling techniques)

Broiler flocks: Day-old chicks

Pieces of inner linings of the delivery boxes are sampled by the owner in the same way as for breeding flocks. The samples have to reach an accredited laboratory within 48 hours of sampling.

Broiler flocks: Before slaughter at farm

All flocks are sampled, by the owner, within 3 weeks before slaughter. The sampling is performed in accordance with Regulation (EU) n 200/2012. Samples have to reach an accredited laboratory within 48 hours.

Broiler flocks: At slaughter (flock based approach)

The intact caeca of 10 poultry from the same flock are taken at the slaughterhouse with the aim to determine the load of Salmonella spp. entering the slaughterhouse.

Case definition

Broiler flocks: Day-old chicks

A sample is considered positive if a Salmonella spp. is isolated. A flock is considered positive as soon as one sample is positive.

Broiler flocks: Before slaughter at farm

A sample is considered positive if a Salmonella spp. is isolated. A flock is considered positive as soon as one sample is positive.

Diagnostic/analytical methods used

Broiler flocks: Day-old chicks

Bacteriological method: ISO 6579:2002 annex D in accordance with regulation (EU) nr. 200/2012.

Broiler flocks: Before slaughter at farm

Bacteriological method: ISO 6579:2002 annex D in accordance with regulation (EU) nr. 200/2012.

Broiler flocks: At slaughter (flock based approach)

Vaccination policy

Broiler flocks

There is no vaccination policy for broiler flocks.

Other preventive measures than vaccination in place

Broiler flocks

Minimal requirements are laid down for holdings with at least 200 broilers on infrastructure, management, hygiene and bio-security issues in the framework of the authorization of holdings.

Control program/mechanisms

The control program/strategies in place

Broiler flocks

The minimal requirements in the framework of the authorization of farms with more than 200 birds contains preventive measures (infrastructure, management, hygiene and biosecurity) for the control of Salmonella. Following measures are taken when a flock is positive for Salmonella spp: 1 logistic slaughter of the flock at the end of production. 2 mandatory cleaning and disinfection. 3 hygienogram after disinfection and after the house has dried up. 4 swab control on the presence of Salmonella before restocking the house. If the following flock is positive for the same serotype of Salmonella, the disinfection must be performed by an external company. When the same serotype of Salmonella is found at three consecutive times, the farm must be evaluated on biosecurity and hygiene by the farm veterinarian and necessary measures must be taken. An epidemiological investigation and/or tests are performed to find the source of the infection. It is at all times prohibited to treat for Salmonella with antibiotics.

Measures in case of the positive findings or single cases

Broiler flocks: Day-old chicks

It is prohibited to treat the flock for Salmonella with antibiotics.

Broiler flocks: Before slaughter at farm

See 'the control program/strategies' in place.

Notification system in place

Zoonotic Salmonella is notifiable since the first of January 2004. Notification is done by phone, fax or by e-mail to the Federal Agency for the Safety of the Food Chain. Farmers and laboratories are obliged to notify.

Results of the investigation

5.473 batches of day-old chicks were sampled, 10 were positive for Salmonella spp. of which 5 for S. Enteritidis and 1 for S. Typhimurium. 8946 flocks of broilers were sampled in the last 3 weeks of production. 177 flocks were positive for Salmonella spp. of which 12 for S. Typhimurium and 7 for S. Enteritidis. The most common other serotypes found were S. Paratyphi B var. Java in 26 flocks, followed by S. Livingstone (22 flocks), S. Infantis (21 flocks), S. Agona (13 flocks) and S. Minnesota (11 flocks).

National evaluation of the recent situation, the trends and sources of infection

The prevalence of all serotypes in day old chicks has slightly decreased compared to 2013. The prevalence of Salmonella spp. in broiler flocks decreased slightly from 2,09% in 2013 to 1,99% in 2014.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

The total number of reported human Salmonella isolates increased in 2014 to 2.963 (2.760 in 2013) due to an increase of the number of Salmonella Typhimurium cases. When comparing the serotypes found in broilers and on poultry carcasses, meat and meat products, S. Typhimurium, S. Java, S. Livingstone and S. Infantis could be found in broilers, on carcasses and in meat and meat products. S. Typhimurium and S. Livingstone were also found in feed for poultry.

3.1.2.3 Salmonella spp. in animal - Pigs

Monitoring system

Sampling strategy

Breeding herds

For diagnostic purposes and in the framework of research projects, pigs are sampled and isolates are sent to the NRL Salmonella, Animal Health for serotyping and resistance analysis.

Multiplying herds

For diagnostic purposes and in the framework of research projects, pigs are sampled and isolates are sent to the NRL Salmonella, AH for serotyping and resistance analysis.

Fattening herds

Every year, 12 blood samples are taken for the serological surveillance of Salmonella on farms with at least 31 fattening pigs. Samples are taken for bacteriological detection on farms that are considered risk herds for Salmonella. For diagnostic purposes and in the framework of research projects, pigs are sampled and isolates are sent to the NRL Salmonella, AH for serotyping and resistance analysis.

Frequency of the sampling

Fattening herds at farm

Fattening herds with at least 31 fattening pigs are sampled every year. Samples are taken for bacteriological detection on farms that are considered risk herds for Salmonella.

Type of specimen taken

Fattening herds at farm

On farm level, blood samples are taken for serological analysis. On risk herds, overshoes are used for bacteriological detection.

Methods of sampling (description of sampling techniques)

Fattening herds at farm

Depending on the capacity of the farm, 10 to 12 blood samples are taken of the fattening pigs. The blood samples are taken of all ages. On risk herds, 4 samples are taken. Each sample consists of one pair of overshoes.

Case definition

Fattening herds at farm

Risk farms are identified as farms with a mean S/P ratio higher than 0,6 for 3 consecutive sampling rounds.

Diagnostic/analytical methods used

Fattening herds at farm

An indirect LPS--Salmonella ELISA is used for the detection of antibodies against certain Salmonella serogroups. The ISO 6576 : 2002 annex D method is used for bacteriological detection, the White-Kauffmann-LeMinor scheme for serotyping.

Vaccination policy

Breeding herds

No vaccine is authorized in Belgium for the vaccination of pigs against Salmonella.

Multiplying herds

No vaccine is authorized in Belgium for the vaccination of pigs against Salmonella.

Fattening herds

No vaccine is authorized in Belgium for the vaccination of pigs against Salmonella.

Control program/mechanisms

The control program/strategies in place

Fattening herds

Risk farms are identified as farms with a mean S/P ratio equal or higher than 0,6 for 3 consecutive sampling rounds. Following mandatory measures are applied on risk farms: 1) completion of a checklist on bio-security and other measures; 2) formulating and implementing a herd specific salmonella action plan, based on the result of the checklist; 3) bacteriological evaluation of the farm.

Measures in case of the positive findings or single cases

The measures are explained under control strategy in place.

Notification system in place

Zoonotic Salmonella is notifiable by operators and laboratories since the first of January 2004. Notification is done by phone, fax or electronic to the Federal Agency of the Safety of the Food Chain.

Results of the investigation

5.144 herds with fattening pigs were sampled in 2014. 1.058 herds had at least once a mean S/P ratio of more than 0,6. 20 herds were classified as Salmonella risk herds for the first time and 31 herds were classified as a Salmonella risk herd for a second or consecutive time. In the framework of bacteriological detection of Salmonella on risk herds, 86 samples were taken on 22 farms. Salmonella could be isolated on 82% of the farms. De main serotypes found were Salmonella Typhimurium (10 herds) and S. O4,(5),12:i:- (4 herds).

National evaluation of the recent situation, the trends and sources of infection

22 of the 51 herds at risk were bacteriologically monitored in 2014. The proportion of herds infected with *S. Typhimurium* is greater than the proportion infected with a monophasic variant of *S. Typhimurium* which was not the case in 2013 when both were seen in equal proportions. The importance of *S. Derby* keeps decreasing.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

The main serotypes found on Salmonella risk farms (fattening pigs), on carcasses and in pig meat is Salmonella Typhimurium and its monophasic variant.

3.1.2.4 Salmonella spp. in animal - Gallus gallus (fowl) - laying hens

Monitoring system

Sampling strategy

Laying hens flocks

All laying hen flocks on farms with at least 200 laying hens are under the Salmonella control programme. Flocks are sampled by the owner at the age of day old chicks, 16, 24, 39 and 54 weeks and in the last 3 weeks of production. When a flock has a second production cycle, the sampling continues every 15 weeks.

Frequency of the sampling

Laying hens: Day-old chicks

Every flock is sampled.

Laying hens: Rearing period

At the age of 16 weeks.

Laying hens: Production period

Every 15 weeks.

Laying hens: Before slaughter at farm

Every flock is sampled.

Laying hens: At slaughter

Sampling is distributed evenly throughout the year.

Type of specimen taken

Laying hens: Day-old chicks

Internal linings of delivery boxes

Laying hens: Rearing period

Socks/ boot swabs

Laying hens: Production period

Socks/ boot swabs in accordance with Regulation (EU) nr. 517/2011.

Laying hens: Before slaughter at farm

Socks/ boot swabs

Laying hens: At slaughter

Other: caeca

Methods of sampling (description of sampling techniques)

Laying hens: Day-old chicks

At the farm, 20 pieces (5 by 5 cm) of the inner linings of delivery boxes are taken of each batch. On voluntary basis, 20 living hen-chicks are brought to the laboratory for serological testing. The samples have to reach an accredited laboratory within 48 hours of sampling.

Laying hens: Rearing period

Samples are taken in accordance with Regulation (EU) No. 517/2011.

Laying hens: Production period

Samples are taken in accordance with Regulation (EU) No. 517/2011.

Laying hens: Before slaughter at farm

Samples are taken in accordance with Regulation (EU) No. 517/2011.

Case definition

Laying hens: Day-old chicks

A sample is considered positive if *S. Enteritidis* or *S. Typhimurium* is isolated. A flock is considered positive as soon as one sample is positive.

Laying hens: Rearing period

A sample is considered positive if *S. Enteritidis* or *S. Typhimurium* is isolated. A flock is considered positive as soon as one sample is positive.

Laying hens: Production period

A sample is considered positive if *S. Enteritidis* or *S. Typhimurium* is isolated. A flock is considered positive as soon as one sample is positive.

Laying hens: Before slaughter at farm

A sample is considered positive if *Salmonella* is isolated. A flock is considered positive as soon as one sample is positive.

Diagnostic/analytical methods used

Laying hens: Day-old chicks

Bacteriological method: ISO 6579:2002 annex D

Laying hens: Rearing period

Bacteriological method: ISO 6579:2002 annex D

Laying hens: Production period

Bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EU) No. 517/2011.

Laying hens: Before slaughter at farm

Bacteriological method: ISO 6579:2002 annex D

Vaccination policy

Laying hens flocks

Vaccination against Salmonella Enteritidis is compulsory and vaccination against Salmonella Typhimurium is strongly recommended.

Other preventive measures than vaccination in place

Laying hens flocks

Minimal requirements for infrastructure, management, hygiene and bio-security issues are laid down in the framework of the authorization of holdings.

Control program/mechanisms

The control program/strategies in place

Laying hens flocks

The national control program for Salmonella in laying hens is based on Regulations (EC) Nos. 2160/2003, 1177/2006 and (EU) No. 517/2011.

Recent actions taken to control the zoonoses

The farmer has the possibility to perform an extended swabcontrol after cleaning and disinfection. This way the possible source of contamination may be found.

Measures in case of the positive findings or single cases

Laying hens flocks

1) Pasteurization of eggs before human consumption.2) Cleaning and disinfection of housing after removal of the positive flock.3) Swab sampling of housing before entering a new flock. If the result is positive for Salmonella, cleaning and disinfection has to be repeated.

Notification system in place

Zoonotic Salmonella is notifiable by the farmer and the laboratory since the first of January 2004. Notification is done by phone, fax or electronic to the Federal Agency for the Safety of the Food Chain.

Results of the investigation

174 different batches with day-old chicks were tested. Salmonella was not found. During rearing, 296 flocks were sampled of which 4 were positive for Salmonella spp. (*S. Senftenberg* (3), *S. Tennessee* (1)). During production, 644 flocks were sampled of which 28 were positive for Salmonella spp. (12 for *S. Enteritidis* and 1 for *S. Typhimurium*). 7 flocks were positive for *S. Infantis*, a serotype that was not found in breeders.

National evaluation of the recent situation, the trends and sources of infection

During rearing, the prevalence increased from 0,36% in 2013 to 1,35% in 2014. This increase in prevalence was not noticed during production where a decrease from 5,94% in 2013 to 4,35% in 2014 was seen. However, the prevalence of *Salmonella Enteritidis* and *Salmonella Typhimurium* increased again to 2%.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

The total number of reported human Salmonella isolates increased in 2014 to 2.963 (2.760 in 2013) due to an increase of the number of *Salmonella Typhimurium* cases. When comparing the serotypes found in layers and on carcasses of layers, mainly *S. Enteritidis* was found in layers and on carcasses. Monophasic *S. Typhimurium* was found in egg products. However, this serotype could not be detected in layers.

3.1.2.5 Salmonella spp. in animal - Gallus gallus (fowl) - breeding flocks, unspecified

Monitoring system

Sampling strategy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Breeding flocks are sampled as day-old chicks, at the age of 4 and 16 weeks and every 2 weeks during production. An official control takes place at 16 weeks, 22 weeks, 46 weeks and 58 or 62 weeks. A specific *Salmonella* control is performed 4 times a year in the hatcheries by the owner.

Frequency of the sampling

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Every flock is sampled

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

As day old chicks and at the age of 4 and 16 weeks

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Every 2 weeks

Type of specimen taken

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Internal linings of delivery boxes

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Socks/ boot swabs

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Socks/ boot swabs

Methods of sampling (description of sampling techniques)

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

At the farm, pieces (5 by 5 cm) of the inner linings of delivery boxes are taken of each flock. 2 samples are taken, one for the hen-chicks and one for the cock-chicks. Each sample consists of 20 pieces of interlining. The two samples are analyzed separately. On voluntary basis, 20 living hen-chicks and 20 living cock-chicks are brought to the laboratory for serological testing. The samples have to be taken the day of delivery, the samples have to reach the lab within 24 hours of sampling. In the hatcheries, pooled samples from dead-in-the-shell chicks and of fluff and meconium, are taken by the owner every 3 months. These are sent to an accredited laboratory.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Samples are taken by the owner at 4 weeks and by one of the animal health organizations at 16 weeks, both in accordance with regulation (EU) Nr. 200/2010.

Breeding flocks: Production period

All samples are taken in accordance with Regulation (EC) Nr. 200/2010.

Case definition

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

A sample is considered positive if Salmonella Enteritidis, Typhimurium, Hadar, Infantis, Virchow or Paratyphi B var. Java is isolated. A flock is considered positive as soon as one sample is positive.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

A sample is considered positive if Salmonella Enteritidis, Typhimurium, Hadar, Infantis, Virchow or Paratyphi B var. Java is isolated. A flock is considered positive as soon as one sample is positive. If the farmer requests a confirmation sampling, new samples (5 feces and 2 dust samples) are taken by or under the supervision of the competent authority. The result of the confirmation sampling is binding.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

A sample is considered positive if Salmonella Enteritidis, Typhimurium, Hadar, Infantis, Virchow or Paratyphi B var. Java is isolated. A flock is considered positive as soon as one sample is positive. If the farmer requests a confirmation sampling, new samples (5 feces and 2 dust samples) are taken by or under the supervision of the competent authority. The result of the confirmation sampling is binding.

Diagnostic/analytical methods used

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EC) Nr. 200/2010.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Rearing period

Bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EC) Nr. 200/2010.

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Bacteriological method: ISO 6579:2002 annex D in accordance with Regulation (EC) Nr. 200/2010.

Vaccination policy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Vaccination against *Salmonella Enteritidis* is compulsory for parent breeding flocks and prohibited for grand parent flocks. Vaccination against *Salmonella Typhimurium* is strongly recommended for parent breeding flocks and prohibited for grandparent flocks.

Other preventive measures than vaccination in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

All holdings with breeding flocks must implement minimum requirement for infrastructure, management, hygiene and biosecurity.

Control program/mechanisms

The control program/strategies in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

The national control programme for *Salmonella* in breeding flocks is based on Regulations (EG) Nrs. 2160/2003, 200/2010 and 1177/2006.

Measures in case of the positive findings or single cases

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

1) treatment of flock with antimicrobials is forbidden; 2) Incubation of hatching eggs is prohibited; 3) Incubated hatching eggs are removed and destroyed; 4) Not yet incubated hatching eggs may be pasteurized and put on the market for human consumption; 5) Positive breeding flocks are slaughtered within the month; 6) Cleaning and disinfection of housing after removal of the breeding flock; 7) After cleaning and disinfection, a hygienogram is performed; 8) Sampling of the house (swab control) for the detection of *Salmonella*; 8) A new flock is admitted if *Salmonella* can not be found after cleaning and disinfection, otherwise the disinfection and swab control is repeated.

Notification system in place

Zoonotic *Salmonella* is notifiable since the first of January 2004. Notification is done by phone, fax or electronically to the Federal Agency for the Safety of the Food Chain. Laboratories and farmers are submitted to the notification.

Results of the investigation

Salmonella was not found in day old chicks (309 batches). During rearing (341 flocks), *S. Mbandaka* and *S. Typhimurium* were each found in 1 flock. During production, of the 503 flocks (grandparent and parent flocks), 2 flocks were positive for *S. Enteritidis*, 3 for *S. Typhimurium* and 16 flocks were positive for serotypes not included in the programme. In addition, one flock was considered negative for *Salmonella Typhimurium* and one for *S. Enteritidis* after confirmation sampling. These flocks do not count as positive flocks.

National evaluation of the recent situation, the trends and sources of infection

During rearing, the number of positive flocks (all *Salmonella* spp.) varies between 3 and 7 in the period 2008 to 2013. Only 2 positive flocks were detected in 2014. During production, the number of positive flocks for *Salmonella* serotypes for which a target is set fluctuates between 0 and 3 in recent years. In 2014, 5 positive flocks were found. The source of infection could not be traced. The number of positive flocks of other serotypes has increased again after a period of gradual decrease (16 in 2011, 12 in 2012, 10 in 2013, 16 in 2014). A positive point is the decrease in the number of suspicious flocks where the presence of *Salmonella* could not be confirmed from 11 in 2011 to 2 in 2014. Three serotypes found in breeders (*S. Anatum*, *S. Djugu* and *S. Jerusalem*) were not found in broilers. However, *S. Mbandaka* was found in 12 flocks of breeders and in 8 broiler flocks.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

The total number of reported human *Salmonella* isolates increased in 2014 to 2.963 (2.760 in 2013) due to an increase of the number of *Salmonella* Typhimurium cases. When comparing the serotypes found in broilers and on poultry carcasses, meat and meat products, *S. Typhimurium*, *S. Java*, *S. Livingstone* and *S. Infantis* could be found in broilers, on carcasses and in meat and meat products. *S. Typhimurium* and *S. Livingstone* were also found in feed destined for poultry.

3.1.2.6 *Salmonella* spp. in Ducks - breeding flocks and meat production flocks

Monitoring system

Sampling strategy

Breeding flocks

Meat production flocks

On farms with a capacity of 5000 or more birds, all flocks are sampled within 3 weeks before slaughter.

Frequency of the sampling

Breeding flocks: Day-old chicks

Breeding flocks: Production period

Meat production flocks: Day-old chicks

Meat production flocks: Before slaughter at farm

All flocks are sampled within 3 weeks before slaughter.

Type of specimen taken

Meat production flocks: Before slaughter at farm

2 pair of overshoes are taken and pooled to one sample.

Methods of sampling (description of sampling techniques)

Meat production flocks: Before slaughter at farm

On farms with more than 5000 birds, all flocks are sampled, by the owner, within 3 weeks before slaughter. 2 pair of overshoes, pooled to 1 sample, are taken. The samples have to reach an accredited laboratory within 48 hours.

Case definition

Meat production flocks: Day-old chicks

A flock is positive if *Salmonella* spp. is found.

Meat production flocks: Before slaughter at farm

A flock is positive if *Salmonella* spp. is found.

Diagnostic/analytical methods used

Meat production flocks: Before slaughter at farm

The bacteriological method used is the ISO 6579:2002 annex D method.

Vaccination policy

Breeding flocks

Meat production flocks

There is no vaccination policy.

Other preventive measures than vaccination in place

Meat production flocks

All holdings have to implement hygienic, infrastructural and management measures in the framework of the authorization of the holding.

Measures in case of the positive findings or single cases

Samples are taken for monitoring purposes only. Flocks are slaughtered at the end of the day (logistic slaughter) if samples taken before slaughter are positive.

Notification system in place

A notification system for zoonotic *Salmonella* is in place since 1 January 2004. The notification can be done by e-mail, fax or phone.

Results of the investigation

There were no flocks sampled in 2014.

National evaluation of the recent situation, the trends and sources of infection

Salmonella spp are seldom found in flocks of meat ducks.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Seen the very low number of meat production flocks of ducks in Belgium, there is very little to no impact on human cases.

Additional information

In 2014, there were no breeding flocks of ducks in Belgium.

3.1.2.7 Salmonella spp. in Geese - breeding flocks and meat production flocks

Monitoring system

Sampling strategy

Breeding flocks

Frequency of the sampling

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Meat production flocks: Day-old chicks

Meat production flocks: Before slaughter at farm

Type of specimen taken

Methods of sampling (description of sampling techniques)

Case definition

Breeding flocks: Day-old chicks

Breeding flocks: Production period

Diagnostic/analytical methods used

Vaccination policy

Breeding flocks

Meat production flocks

Other preventive measures than vaccination in place

Measures in case of the positive findings or single cases

Meat Production flocks

Notification system in place

Results of the investigation

Additional information

In 2014 there were no breeding and meat production flocks of geese in Belgium.

3.1.2.8 Salmonella spp. in Turkeys - breeding flocks and meat production flocks

Monitoring system

Sampling strategy

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

There are no professional breeding turkey flocks in Belgium.

Meat production flocks

All flocks are sampled within three weeks of slaughter.

Frequency of the sampling

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Day-old chicks

Breeding flocks (separate elite, grand parent and parent flocks when necessary): Production period

Meat production flocks: Day-old chicks

Meat production flocks: Before slaughter at farm

Every flock is sampled

Type of specimen taken

Meat production flocks: Before slaughter at farm

Socks/ boot swabs

Methods of sampling (description of sampling techniques)

Meat production flocks: Before slaughter at farm

All flocks are sampled, by the owner, within 3 weeks before slaughter conform Regulation (EU) n 1190/2012.

Case definition

A flock is positive if Salmonella is found.

Case definition

Meat production flocks: Before slaughter at farm

A flock is positive if Salmonella spp. is found.

Diagnostic/analytical methods used

Meat production flocks: Day-old chicks

Bacteriological method: ISO 6579:2002 annex D.

Meat production flocks: Before slaughter at farm

Bacteriological method: ISO 6579:2002 annex D as described in Regulation (EU) 1190/2012.

Vaccination policy

Meat production flocks

There is no vaccination policy for meat production flocks.

Other preventive measures than vaccination in place

Breeding flocks (separate elite, grand parent and parent flocks when necessary)

Meat production flocks

In the framework of the authorization of holdings, infrastructural, management, hygiene and bio-security measures must be implemented on all holdings.

Measures in case of the positive findings or single cases

Following measures are taken when a flock is positive for Salmonella spp for the first time:1 the flock is at the end of the production cycle slaughtered at the end of the day (logistic slaughter);2 there is an obligation to clean and disinfect the house;3 a hygienogram is performed after disinfection and after the house has dried up;4 a swab control on the presence of Salmonella is performed before restocking the house;If the following flock is positive for the same serotype of Salmonella, the same measures are taken and the disinfection must be performed by an external company. When the same serotype of Salmonella is found at three consecutive times, besides the above mentioned measures, the farm must be evaluated on biosecurity and hygiene by the farm veterinarian and necessary measures must be taken. An epidemiological investigation and/or tests are performed to find the source of the infection. It is at all times prohibited to treat for Salmonella with antibiotics.

Notification system in place

Zoonotic Salmonella is notifiable since 1 January 2004. Notification is done by phone, fax or e-mail.

Results of the investigation

There are no turkey breeding flocks in Belgium.⁸² meat production flocks were tested in 2014. There was one flock positive for S. Typhimurium , one for S. O4,5,12:i:- and 4 for S. Chester.

National evaluation of the recent situation, the trends and sources of infection

There is a higher incidence of Salmonella in turkey meat production flocks than seen in previous years. The cause is not known. All hatching eggs or day-old-chicks are from neighbouring countries.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

Seen the limited number of meatturkey flocks slaughtered in Belgium, there is little to no relevance of the findings in these flocks to human cases.

3.2 CAMPYLOBACTERIOSIS

3.2.1 General evaluation of the national situation

3.2.1.1 Thermophilic Campylobacter spp., unspecified - general evaluation

History of the disease and/or infection in the country

Campylobacteriosis is a leading bacterial foodborne gastrointestinal disease in humans in all parts of the world. It can also cause post-infectious complications as Guillain-Barré syndrome. In 80% of the cases, the infection route of campylobacteriosis is food, but domestic animals including pets can also be involved. The transmission of this pathogen to humans is mostly due to consumption of undercooked poultry, pork and beef, unpasteurized milk, contaminated drinking water, or contacts with the faeces of infected pets. This report will focus on *Campylobacter jejuni* and *Campylobacter coli* that are the principal strains causing enteritis in humans. The contamination with *Campylobacter* of poultry carcasses and meat is monitored since 2000 by the Federal Agency for the Safety of the Food Chain. The rate of positive poultry samples is stable, but high. Chicken and layer meat have to be well cooked and cross-contamination should be avoided during preparation.

3.2.2 Campylobacter in foodstuffs

3.2.2.1 Thermophilic Campylobacter spp., unspecified in food - Meat from broilers (*Gallus gallus*)

Monitoring system

Sampling strategy

At slaughterhouse and cutting plant

A monitoring program was organized by FASFC to evaluate the level of *Campylobacter* spp. contamination of broiler meat in Belgian slaughterhouses and cutting plants. *Campylobacter* is counted on carcasses and cuts of poultry because it is especially the quantitative load of *Campylobacter* which plays a role in the stake in danger of the consumers.

Frequency of the sampling

At slaughterhouse and cutting plant

Sampling distributed evenly throughout the year

At meat processing plant

Sampling distributed evenly throughout the year

At retail

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughterhouse and cutting plant

Neck skin samples and cuts of broilers with and without skin

At meat processing plant

Meat, minced meat, sausages and other

At retail

Meat, minced meat, sausages and other

Methods of sampling (description of sampling techniques)

At slaughterhouse and cutting plant

The matrices were carcasses, cuts and meat preparation of broilers. The *Campylobacter* spp. contamination levels were analyzed : 1g carcasses, 1g cutting meat and 1g meat preparation.

At meat processing plant

The samples were about 200 g of meat. The amount of *Campylobacter* has been assessed in 1g of sample.

At retail

The amount of *Campylobacter* has been assessed in 1g of sample.

Definition of positive finding

At slaughterhouse and cutting plant

A sample is considered positive in case of detection of more than 1.000 cfu *Campylobacter* for carcasses and meat with skin and in case of detection of more than 100 cfu *Campylobacter* for meat without skin.

At meat processing plant

A sample is considered positive in case of detection of more than 100 cfu *Campylobacter* in the sample (1.000 cfu for carcasses and meat with skin).

At retail

A sample is considered positive in case of detection of more than 100 cfu *Campylobacter* in the sample (1.000 cfu for carcasses and meat with skin).

Diagnostic/analytical methods used

At slaughterhouse and cutting plant

Bacteriological method: ISO 10272:1995

3.2.2.2 Thermophilic *Campylobacter* spp., unspecified in food

Monitoring system

Sampling strategy

A monitoring program was organized by the Federal Agency for the Safety of the Food Chain. More than 200 Belgian slaughterhouses, more than 100 meat cutting plants and more than 100 retail trades representative of the Belgian production of carcasses and meat, were selected for this study. The samples assayed were carcasses and minced meat from pork, carcasses, cuts and meat preparation from chicken, and layer carcasses. Sampling was done by a specially trained staff of the Federal Agency for the Safety of the Food Chain.

Frequency of the sampling

Samples have been taken every week from the first to the 52nd week, except during the 30th week.

Type of specimen taken

meat and dairy products

Methods of sampling (description of sampling techniques)

Sampling of pork carcasses was done by means of swabs (4 areas from the same half carcass constituting 600 cm² were putted in the same stomacher bag). The carcass samples of broiler and layer consisted of 10g of neck skin. The other samples were about 200g of meat. 10g to 25g representative of the whole sample were weighted in the laboratory, and the detection of *Campylobacter* has been assessed in these quantities or dilutions: 25g for pork minced meat, 600 cm² (pork carcasses), 0,01g for chicken carcasses and layer carcasses, 1g for chicken meat preparation, and for chicken cuts, 0,1g and 25g. No pooling has been done.

Definition of positive finding

A sample is considered to be positive after biochemical or genetic confirmation of one *Campylobacter* in the sample.

Diagnostic/analytical methods used

For detection of *Campylobacter* in meat samples or swabs the official Belgian SP-VG-M003 method was used following : - selective enrichment on Preston at 42C for 48 h,- isolation on mCCDA at 42C for 24 h - 120 h,- confirmation of minimum 1 colony with miniaturised biochemical tests or by PCR typing.

Measures in case of the positive findings or single cases

Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch or single sample- Further investigation: additional sampling, possible recall, RASFF, ...

National evaluation of the recent situation, the trends and sources of infection

The results showed that, even if the contamination by *Campylobacter* spp. of pig carcasses is not zero, the pork represents a relatively low risk for the consumer seen the evolution of this contamination during the operations of cut.

3.2.3 *Campylobacter* in animals

3.2.3.1 Thermophilic *Campylobacter* spp., unspecified in animal - *Gallus gallus* (fowl)

Monitoring system

Sampling strategy

Frequency of the sampling

At slaughter

Sampling distributed evenly throughout the year

Type of specimen taken

At slaughter

caeca

Methods of sampling (description of sampling techniques)

At slaughter

10 caeca pairs are pooled to one sample. 6 samples are taken of each examined flock. The caeca are emptied at the laboratory. The content is examined for *Campylobacter*.

Case definition

At slaughter

A sample is positive if *Campylobacter* is detected.

Measures in case of the positive findings or single cases

Samples are taken for monitoring purposes only. No measures are taken in case of positive findings.

3.3 LISTERIOSIS

3.3.1 General evaluation of the national situation

3.3.1.1 Listeria - general evaluation

National evaluation of the recent situation, the trends and sources of infection

Listeria monocytogenes has become a major concern of the food industry and public health authorities. Ingestion of food contaminated with *Listeria monocytogenes* may cause either a serious invasive illness affecting people with altered or deficient immune responses, or a non-invasive febrile gastro-enteritis. Although the incidence of listeriosis is low, the high mortality rate, which often reaches as high as 30-40%, requires early diagnosis and appropriate antimicrobial therapy. Listeriosis is transmitted to humans via contact with animals, cross-infection of foetus or newborn babies and foodborne infection. *Listeria* is ubiquitous and widely distributed in the environment (soil, vegetables, meat, milk, fish). All food associated with *Listeria monocytogenes* outbreaks were consumed without further processing or after minimal heat treatment, and many of them had a suitable environment for growth.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Recent actions taken to control the zoonoses

General food hygiene rules are essential for the prevention of human listeriosis. As some persons are at high risk (pregnant women, immunocompromised people), they are advised not to eat certain categories of food with proven elevated risk of *Listeria monocytogenes* contamination, such as unpasteurized milk and butter, soft cheeses and ice cream made from unpasteurized milk, any soft cheese crust, smoked fish, pat, cooked ham, salami, cooked meat in jelly, raw minced meat from beef, pork and poultry, steak tartar, raw fish and shellfish (oysters, mussels, shrimps), fish, meat and surimi salads, insufficiently rinsed raw vegetables, unpeeled fruit.

3.3.2 Listeria in foodstuffs

3.3.2.1 *L. monocytogenes* in food

Monitoring system

Sampling strategy

Frequency of the sampling

At the production plant

At retail

Samples are taken according to the national control program or in the frame of RASFF, complaints or suspicion.

Type of specimen taken

At retail

r

Methods of sampling (description of sampling techniques)

At retail

Definition of positive finding

At the production plant

A sample is considered to be positive after confirmation of *Listeria monocytogenes* on chromogenic medium.

At retail

A sample is considered to be positive after confirmation of *Listeria monocytogenes* on chromogenic medium.

Diagnostic/analytical methods used

At the production plant

AFNOR validated VIDAS LMO2 followed by a chromogenic medium (Rapid L. mono or ALOA)

At retail

AFNOR validated VIDAS LMO2 followed by a chromogenic medium (Rapid L. mono or ALOA)

Control program/mechanisms

The control program/strategies in place

Controls are realized by the FASFC in case of notification.

Measures in case of the positive findings

Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch or single sample- Further investigation: additional sampling, possible recall, RASFF, ...

Notification system in place

Notification is mandatory since 1/3/2004 (Ministerial Decree on mandatory notification in the food chain of 22/1/2004). For *Listeria monocytogenes*, the criterion of 100 cfu/g in ready-to-eat food putted on the market may not be exceeded. Laboratories have to inform the Federal Agency for the Safety of the Food Chain in case of a positive sample.

3.4 E. COLI INFECTIONS

3.4.1 General evaluation of the national situation

3.4.1.1 Verotoxigenic E. coli (VTEC) - general evaluation

History of the disease and/or infection in the country

National evaluation of the recent situation, the trends and sources of infection

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

3.4.2 Escherichia coli, pathogenic in foodstuffs

3.4.2.1 Verotoxigenic E. coli (VTEC) in food

Monitoring system

Sampling strategy

A monitoring program was organized by the Federal Agency for the Safety of the Food Chain. More than 200 Belgian slaughterhouses, more than 100 meat cutting plants and more than 100 retail trades representative of the Belgian production, were selected for this study. The samples assayed were carcasses, cuts and minced meat from beef and other foodstuffs. Sampling was done by a specially trained staff of the Federal Agency for the Safety of the Food Chain.

Frequency of the sampling

Samples have been taken every week from the first to the 52nd week, except during the 30th week.

Type of specimen taken

Other: Meat, sprouted seeds, cheeses and other dairy products, pre-cut fruits and vegetables and vegetables.

Methods of sampling (description of sampling techniques)

Sampling of beef carcasses was done by means of swabs (4 areas from the same half carcass constituting 1600 cm² were putted in the same stomacher bag). The samples were putted in a cool box and transported to a dispatching center of the Federal Agency for the Safety of the Food Chain and the laboratory take the samples at the dispatching center for analyses. The other samples were about 200g of meat. The detection of enterohemorrhagic E. coli has been assessed in 1600 cm² for beef carcasses and in 25g for beef minced meat and beef cuts.No pooling has been done.

Definition of positive finding

A sample is considered positive after isolation and genetic confirmation of the pathogenicity of the O157 E. coli strain in the sample. In case of isolation and genetic confirmation of the top 5 VTEC in dairy products, the sample is considered positive. In sprouted seeds, pre-cut fruits and vegetables and (non-pre-cut) vegetables a samples is also considered positive after isolation and genetic confirmation of E. coli O104:H4.

Diagnostic/analytical methods used

For detection of Escherichia coli O157, the Belgian official SP-VG-M001 method, according to the ISO 16654 (2001) was used : - pre-enrichment in m-TSB + novobiocin at 42C for 7 hours,- enrichment in CT-Mac Conkey at 37C for 16-18 hours;- immunoassay O157 (VIDAS ECO, bioMrieux),- selective immunomagnetic enrichment (Dynabeads, Dynal or VIDAS ICE, bioMrieux),- isolation on sorbitol-Mac Conkey and incubation at 42C for 18 h,- isolation and confirmation (agglutination of latex particles, Oxoid),- search for genes encoding for virulence factors in national reference laboratory.For the detection of other E.coli types, the ISO/PRF TS 13136 (2012) method is used.

Preventive measures in place

Controls are in place by the Federal Agency in case of notification.

Control program/mechanisms

The control program/strategies in place

Notification is mandatory since 1/3/2004 (Ministerial Decree on mandatory notification in the food chain of 22/1/2004). For enterohemorrhagic E. coli, absence in 25g in ready-to-eat food putted on the market is mandatory. Laboratories have to inform the Federal Agency in case of positive sample.

Measures in case of the positive findings or single cases

Meat from positive carcasses is traced back, destroyed or transformed into cooked meat products.Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch or single sample-Further investigation: additional sampling, possible recall, RASFF, ...

3.4.3 Escherichia coli, pathogenic in animals

3.4.3.1 Verotoxigenic E. coli (VTEC) in animal - Cattle (bovine animals)

Monitoring system

Sampling strategy

There was no sampling strategy for VTEC in cattle in 2014. Diagnostic veterinary laboratories sent E. coli strains to the NRL E. coli, AH for diagnostic reasons (antimicrobial susceptibility testing, pathotyping) and on a voluntary basis.

Type of specimen taken

Animals at slaughter (herd based approach)

Diagnostic/analytical methods used

Animals at farm

Results of the investigation

3.5 YERSINIOSIS

3.5.1 General evaluation of the national situation

3.5.1.1 Yersinia - general evaluation

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Only a few strains of *Y. enterocolitica* cause illness in humans. The major animal reservoir for *Y. enterocolitica* strains that cause human illness are pigs but other strains are also found in many other animals including rodents, rabbits, sheep, cattle, horses, dogs, and cats. In pigs, the bacteria are most likely to be found on the tonsils. Infection is most often acquired by eating contaminated food, especially raw or undercooked pork products. Drinking contaminated unpasteurized milk or untreated water can also transmit the infection.

3.5.2 Yersinia in animals

3.5.2.1 Yersinia in animal - Pigs

Monitoring system

Sampling strategy

Animals at slaughter (herd based approach)

A monitoring program was organized by FASFC to evaluate the level of *Yersinia enterocolitica* contamination of pigs carcasses in Belgian slaughterhouses.

Frequency of the sampling

Animals at slaughter (herd based approach)

Sampling distributed evenly throughout the year

Type of specimen taken

Animals at slaughter (herd based approach)

Surface of carcasses

Methods of sampling (description of sampling techniques)

Animals at slaughter (herd based approach)

swabs

3.6 TRICHINELLOSIS

3.6.1 General evaluation of the national situation

3.6.1.1 Trichinella - general evaluation

History of the disease and/or infection in the country

Since 1940, the Competent Authority did organize analysis for Trichinella in pigs at the slaughterhouses. The analysis is generalized since 1991. Trichinella has not been detected in carcasses of pigs and horses produced for human consumption in Belgium. One autochthonous human case, probably caused by a home raised wild boar occurred in 1979.

National evaluation of the recent situation, the trends and sources of infection

Trichinellosis is virtually absent in Belgian domestic livestock. Since systematic controls of pigs and horses are done at slaughter (EU Directive 92/45/EEC) no positive case was found. The last outbreak in humans in Belgium occurred in 1979 following the consumption of meat from a wild boar. Increased monitoring in the last decade has shown that Trichinella spp. still circulate amongst wildlife, although both the prevalence and the intensity of infections are low. EU Directive requires that also wild boars hunted in the EU for commercial purpose are examined for Trichinella. Each year about 10.000 sport-hunted wild boars were tested. Until now, one animal, in 2004, originating from Mettet (province of Namur), was found to harbor a light infection. The larvae, isolated by artificial digestion were identified by PCR to be Trichinella britovi, a species previously not demonstrated in Belgium. T. britovi has sylvatic carnivores as main hosts. Even if wild boars are not the preferred host they can acquire the infection and consequently pass it to humans. Both T. spiralis and T. britovi have been associated with human infection. One larva was recovered from a pooled sample (originating from three wild boars from a hunting party from Alle-sur-Semois) in 2007. Consecutive digestions could not reveal the causative animal, and unfortunately PCR failed to identify the Trichinella species. One larva was recovered from the digestion of an individual wild boar in 2012. The routine examination of wild boars devoted to the market has proven to be a good measure to protect the consumer against sylvatic trichinellosis. In addition, monitoring of infection through examining sentinel animals, such as the fox, is recommended to assess the prevalence of trichinellosis and to follow trends in time. In 2014 this monitoring wasn't realized. Serological examination might be an alternative for muscle digestion in screening programs, but can not be used in safeguarding consumer's health in meat inspection. An extra measure to protect the consumer is to eat meat of wild boar "well done", or to freeze the meat at -20C for 4 weeks. An important measure to avoid spreading of the infection among wildlife is not to leave offal of animal carcasses in the field after skinning.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

The last outbreak in humans in Belgium occurred in 1979 following the consumption of meat from wild boar.

Recent actions taken to control the zoonoses

Monitoring of wildlife. Routine examination of wild boars destined for human consumption. Monitoring of infection through examining sentinel animals such as the fox was not further realized in 2013 and 2014. Recommendation to consume wild boar meat after freezing at -20C for 4 weeks. Recommendation to travelers not to import raw meats of unknown origin and of susceptible animals, e.g. home made sausages, and not to consume meats of unknown quality abroad.

Suggestions to the European Union for the actions to be taken

Additional information

The status "negligible risk for Trichinella in slaughterpigs kept under industrial housing conditions" was granted by the EC to Belgium end December 2010.

3.6.2 Trichinella in animals

3.6.2.1 Trichinella in animal - Solipeds, domestic - horses

Monitoring system

Sampling strategy

Permanent surveillance at the slaughterhouses.

Frequency of the sampling

Every slaughtered animal is sampled.

Type of specimen taken

Diaphragm, tongue or masseter muscle.

Methods of sampling (description of sampling techniques)

Horse: 5 gram of diaphragm (or tongue, or masseter) for routine diagnosis, analyses on pooled samples, 10 to 25 gram for examination of individual samples.

Case definition

An animal is considered positive in case of detection and identification of Trichinella larvae in the muscle sample.

Diagnostic/analytical methods used

Artificial digestion method of collective or individual samples. The magnetic stirrer method for digestion of pooled samples as described in Commission Regulation (EC) No 2075/2005 was used on samples of 5 gram of muscles from horses.

Control program/mechanisms

The control program/strategies in place

Commission Regulation (EC) No 2075/2005 imposes systematic Trichinella examination of all slaughtered pigs, horses and wild boar and other wildlife animals by artificial digestion method of muscle before marketing.

Measures in case of the positive findings or single cases

Carcasses found positive are declared unfit for human consumption.

Notification system in place

Notification to the Federal Agency for the Safety of the Food Chain is compulsory for any positive test result.

Results of the investigation including the origin of the positive animals

No positive animals were detected this reporting year.

National evaluation of the recent situation, the trends and sources of infection

No positive horses were found in 2014.

3.6.2.2 Trichinella in animal - Pigs

Monitoring system

Sampling strategy

General

Permanent surveillance of all slaughtered pigs at the slaughterhouses in implementation of Commission Regulation (EC) No 2075/2005. Derogation for fattening pigs who do apply for the criteria set in the definition 'Region with negligible risk'

For regions with negligible Trichinella risk

Frequency of the sampling

General

Systematic Trichinella examinations of all slaughtered pigs, with the exception of some fattening pigs who do apply for the criteria set in the definition 'Region with negligible risk'.

For regions with negligible Trichinella risk

Systematic Trichinella examinations of all slaughtered pigs, with the exception of some fattening pigs who do apply for the criteria set in the definition 'Region with negligible risk'.

Type of specimen taken

General

Diaphragm muscle, 1 gram for fattening pigs, 2 grams for sows and boars.

For regions with negligible Trichinella risk

Diaphragm muscle, 1 gram for fattening pigs, 2 grams for sows and boars. No samples are examined from some fattening pigs who do apply to the criteria set in the definition of 'Region with negligible risk'.

Methods of sampling (description of sampling techniques)

General

Fattening pigs: 1 gram of diaphragm muscle to be pooled (up to 100 animals in 1 pool) Sows and boars: 2 grams of diaphragm muscle to be pooled (up to 50 animals in 1 pool)

For regions with negligible Trichinella risk

Still almost all pigs are sampled and tested, due to logistic reasons and export outside EU.

Case definition

General

An animal is considered positive in case of detection and identification of Trichinella larvae in the muscle sample.

For regions with negligible Trichinella risk

Same as general

Diagnostic/analytical methods used

General

Artificial digestion method of collected samples. (Reference method, annex I, chapter I) and Magnetic stirrer method for pooled sample digestion/on filter isolation and larva detection by a latex agglutination test (equivalent method) The analysis is done by artificial digestion: the magnetic stirrer method of pooled 100 gram sample as described in Commission Regulation (EC) No 2075/2005, reference method, 1 gram per fattening pig, 2 grams per sow and boar, and 5 grams per horse and wild boar. Serology may be done in live pigs and for epidemiological studies and monitoring on wildlife.

Officially recognised regions with negligible Trichinella risk

Belgium was granted the status of negligible Trichinella risk at the end of 2010 by the European Commission

Notification system in place

Notification to the Federal Agency for the Safety of the Food chain is compulsory for any positive test result.

Measures in case of the positive findings or single cases

Carcasses found positive are declared unfit for human consumption.

Results of the investigation including description of the positive cases and the verification of the Trichinella species

No positive cases were found in 2014

Results of the investigation including description of the positive cases and the verification of the Trichinella species

Fattening pigs raised under controlled housing conditions in integrated production system

all negative

Fattening pigs not raised under controlled housing conditions in integrated production system

all negative

Breeding sows and boars

all negative

National evaluation of the recent situation, the trends and sources of infection

Since 1992, when the European Union Council Directive requires that wild boars (*Sus scrofa*) hunted in EU for commercial purpose should be examined for *Trichinella*, the infection has only been detected three times in wild boars of Belgium. There is serological evidence of the presence of anti-*Trichinella* antibodies in wildlife. Wildlife monitoring did not reveal any larvae in winter 2010 (318 foxes examined), but yielded a larva from a pool of 20 wild animals (18 foxes and 2 badgers) in winter 2011-2012 (524 wild animals examined). Unfortunately, the larva could not be identified to the species level by PCR, nor could the individual animal be identified. During winter 2012-2013 540 wild animals were examined and three larvae were recovered from two pools of 20 foxes each.

3.7 ECHINOCOCCOSIS

3.7.1 General evaluation of the national situation

3.7.1.1 Echinococcus - general evaluation

History of the disease and/or infection in the country

At the slaughterhouses, a small number of carcasses showing lesions of *Echinococcus* (cysts) are sometimes detected and notified to the Federal Agency for the Safety of the Food Chain. In case of positive findings, carcasses are partially or totally rejected and declared unfit for human consumption.

National evaluation of the recent situation, the trends and sources of infection

Echinococcosis is caused either by *Echinococcus granulosus* or *Echinococcus multilocularis*. *Echinococcus granulosus* produces unilocular human hydatidosis. It is a small tapeworm (6 mm) that lives in the small intestine of domestic and wild canids. Sheep and cattle serve as intermediate hosts for the infection. Humans acquire infection by ingestion of typical taeniid eggs, which are excreted in the faeces of infected dogs: the oncospheres liberated from the eggs migrate via the bloodstream to the liver, lungs and other tissues to develop in hydatid cysts. Indigenous unilocular hydatidosis in man has been reported in Belgium. In 2014 no cysts were found by post-mortem inspection of the carcasses at the slaughterhouses. *Echinococcus multilocularis* causes alveolar (multilocular) echinococcosis in humans. Foxes and dogs are the definitive hosts of this parasite and small rodents the intermediate hosts. In the liver of rodents the invasive larval stage has a multi-compartmented appearance containing many protoscolices. Ingestion of the eggs by humans can result in the development of invasive cysts in the liver. In Belgium, the percentage of infected foxes varies with the region, with a decreasing rate from the South-East to the North-West: e.g. 33% in the Ardennes, 13% in the Condroz region and 1-2% in Flanders Region. The endemic region is situated under the river Meuse, on the heights of the Ardennes in the Walloon Region. In 2014 in the Flemish Region, 317 hunted foxes were analysed whereof 6 were infected (molecular confirmation). The foxes originated from the provinces of Antwerp, East-Flanders, West-Flanders, Flemish Brabant and Limburg with respectively 60, 55, 75, 54 and 73 foxes. One fox of the province of Flemish Brabant (location near to Limburg) and 5 foxes of the province of Limburg (all from the same municipality Voeren) were positive.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Post mortem visual examination is performed at the slaughterhouses in the domestic intermediate hosts: cattle, sheep, horses and pigs. Whole carcasses or parts are rejected in case *Echinococcus granulosus* cysts are found.

Recent actions taken to control the zoonoses

Consumption of berries is discouraged by warning messages, displayed to visitors of Parks and Woodlands.

3.8 RABIES

3.8.1 General evaluation of the national situation

3.8.1.1 Lyssavirus (rabies) - general evaluation

History of the disease and/or infection in the country

Since the last indigenously acquired case of rabies occurred in Belgium in a bovine coming from Bastogne (province of Luxembourg) in July 1999, Belgium obtained the official status of rabies-free country in July 2001 according to the WHO recommendations (1992) and the Office Internationale des Epizooties (OIE) guidelines (1997).

National evaluation of the recent situation, the trends and sources of infection

In October 2007, Belgium lost temporarily its official status of rabies free country due to a positive case of rabies in a dog, illegally imported from Morocco. Belgium regained again its official free status of rabies on 28 October 2008.

Recent actions taken to control the zoonoses

Surveillance system and methods used. Domestic animals with nervous symptoms that are suspected of rabies have to be notified to the Federal Agency for the Safety of the Food chain. Wildlife found dead or shot should also be declared and send for analysis to the Scientific Institute of Public Health, the National Reference laboratory of rabies. Collection of dead-found bats is recommended for rabies surveillance. Live suspected animals are killed and their brain is examined by immunofluorescence and virus cultivation in neuroblasts at the Scientific Institute of Public Health. The high percentage of examinations of cattle is in consequence of the surveillance system for TSE in cattle: all suspected BSE cases were first examined for rabies. Rabies must be considered in the differential diagnosis of BSE, although the clinical course of rabies is usually quicker than the evolution of clinical nervous symptoms in case of BSE. The oral vaccination campaign of foxes with vaccine baits started in 1989 and was stopped by the end of 2003. In the southern part of the country, below the rivers Sambre and Meuse, vaccination of dogs and cats is compulsory. In addition, all pets staying on any Belgian public camping must be vaccinated.

Suggestions to the European Union for the actions to be taken

It is highly recommended to report on the rabies virus type detected to be able to differentiate between the classical rabies type (genotype 1) and the European bat Lyssa virus types (unspecified or EBL 1 or EBL 2). Bat rabies is of public health concern. The public should be made aware of the danger of human exposure to bats, especially in case of abnormal behavior of bats. Rabies is transmitted to humans and other animals through saliva, usually by a bite. Any person exposed to bats should be previously vaccinated against rabies. Nobody should handle diseased or dead bats without protection such as gloves. Any person finding a bat behaving abnormally, in an unusual place, or under unusual circumstances, should not attempt to handle or to move the animal but should contact official authority. Education and recommendations should be given to travelers in order to reduce their risk of infection. Although dogs represent a more serious threat in many countries, yet the risk of rabies infection by bat bites also exists. Pre-exposure vaccination should be offered to persons at risk, such as laboratory workers, veterinarians, animal handlers, international travelers. Currently available vaccines are safe and effective against both the classical rabies virus and the bat Lyssa viruses.

3.8.2 Lyssavirus (rabies) in animals

3.8.2.1 Lyssavirus (rabies) in animal - Dogs

Monitoring system

Sampling strategy

The brain of dogs with nervous symptoms suspected of rabies are examined by direct immunofluorescence test and virus cultivation in neuroblasts at the Scientific Institute of Public Health, the National Reference Laboratory for rabies.

Frequency of the sampling

All suspected dogs with clinical nervous symptoms are tested.

Type of specimen taken

Brain

Methods of sampling (description of sampling techniques)

Small animals: head / carcass
Huge animals: brain (CNS)
Shipping and packaging conditions: Brains are transported as soon as possible (refrigerated if possible) in a tightly sealed packet to the National Reference Laboratory. In case of transport of a carcass, an authorization is required. The storage period of samples at the National Reference Laboratory for further analysis is one year.

Case definition

An animal is considered positive in case of a positive direct immunofluorescence test (Antigen detection) confirmed by cell cultivation of the virus or detection by RT-PCR or (rarely performed) by mice inoculation test (clinical observation of rabies symptoms).

Diagnostic/analytical methods used

Direct immunofluorescence for the detection of viral antigen, virus isolation in neuroblastoma cell culture, detection by RT-PCR, mouse inoculation test

Vaccination policy

In the Southern part of the country, below the rivers Sambre and Meuse, vaccination of dogs and cats is compulsory. In addition, all pets staying on any Belgian public camping must be vaccinated. Oral vaccination of foxes by baits started in 1989. Since there were no more cases of rabies for the last years, oral vaccination of foxes by baits was stopped by the end of 2003.

Measures in case of the positive findings or single cases

In case of positive findings national legislation has to be applied (Royal Decree of 10 February 1967, Royal Decree of 22 May 2005 and Ministerial Decree of 23 February 1967).

Notification system in place

Royal Decree of 10 February 1967, Animal Health Law of 24 March 1987 Chapter III and Royal Decree of 3 February 2014 (list of all notifiable animal diseases)
Notification of all laboratory confirmed cases to the competent Authority is mandatory.

National evaluation of the recent situation, the trends and sources of infection

3.9 STAPHYLOCOCCUS AUREUS METICILLIN RESISTANT (MRSA) INFECTION

3.9.1 Staphylococcus in foodstuffs

3.9.1.1 Staphylococcus in food

Monitoring system

Sampling strategy

Tests for Staphylococcus were performed in minced meat, dairy products, shellfish and bakery products.

Frequency of the sampling

Samples are taken according to the national control program or in the frame of RASFF, complaints or suspicion.

Type of specimen taken

minced meat, milk, shellfish and bakery products,...

Methods of sampling (description of sampling techniques)

The samples were taken according to Regulation (EC) No 2073/2005.

Definition of positive finding

To determine the conformity of a sample or a batch, the criteria laid down in the Regulation (EC) No 2073/2005 are applied.

Diagnostic/analytical methods used

The method used is according to Regulation (EC) No 2073/2005.

Measures in case of the positive findings or single cases

Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch or single sample- Further investigation: additional sampling, possible recall, RASFF, ...

3.9.2 Staphylococcus in animals

3.9.2.1 Staphylococcus in animal

Monitoring system

Sampling strategy

In 2014, monitoring of the presence of MRSA and the antimicrobial resistance was performed in laying hens and broilers. The number of holdings to be sampled was based on the results of the same monitoring in 2011.

Frequency of the sampling

The samples were taken divided over the year. Only one flock per holding is sampled. The monitoring is repeated every 3 years.

Type of specimen taken

10 nasal swabs were taken on each holding and pooled to one sample.

Case definition

A flock/holding is positive when MRSA is detected and confirmed by PCR.

Diagnostic/analytical methods used

The samples were incubated in Mueller-Hinton (MH) broth (Becton Dickinson) supplemented with NaCl (6.5%) at 37C for 18-24h. One ml of this broth was added to Tryptic Soy Broth (TSB) supplemented with cefoxitin (3.5mg/l) and aztreonam (75mg/l) and incubated at 37C for 18-24h. Per sample, one to five suspected colonies were selected from the Brilliance MRSA 2 plate. DNA was extracted as described in SOP/BAC/ANA/18. MRSA confirmation was performed using a triplex real-time PCR method. This PCR allows detecting the Staphylococcal aureus specific gene, nuc, the presence of the mecA gene responsible for methicillin resistance and the variant mecC gene. Ten microliter of this enrichment was plated on Brilliance MRSA 2 (Oxoid) and incubated 18-24h at 37C. Presence of MRSA was suspected based on colony morphology and confirmed using a triplex real-time PCR method. All MRSA isolates were spa-typed by sequencing the repetitive region of the spa gene encoding for the staphylococcal protein A. This method depicts the rapid evolution, since through recombination, the repeats may change fast. The protein A (spa) gene was amplified according to the Ridom StaphType standard protocol (www.ridom.de/staphtype) and the amplification was checked on a 2% agarose gel. Sequencing was performed with CEQ 8000 using standard protocols and sequences were compared with the international Ridom database. CC398 PCR was performed on all MRSA following protocol described by Stegger et al. 2011. This method allows the rapid detection of the S. aureus sequence type ST398.

Measures in case of the positive findings or single cases

There are no measures linked to positive findings. However, farmers are informed of the presence of MRSA on the holding and on possible measures to protect themselves. General hygiene and biosecurity measures are promoted.

Results of the investigation

159 broiler farms were sampled, 22 confirmations were performed (PCR) of which 2 were positive. SpA-typing confirmed a t1985 and a t011 strain, both belonging to CC398. 233 layer farms were sampled, 19 PCR's were performed of which 6 were positive. Spa-typing confirmed 5 t037 strains not belonging to CC398 and one t011 strain belonging to CC398. 12 rearing layer farms were sampled, MRSA could not be found on these farms.

National evaluation of the recent situation, the trends and sources of infection

Compared to the results in 2011, the prevalence of MRSA on broiler farms decreased from 6,5% to 1,25%. The prevalence in layers increased from 0,7% in 2011 to 2,4% in 2014. However, seen only the results of 2 years are available, it is too early to draw any conclusions.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

The presence of MRSA in poultry is still very low compared to the presence in pigs and bovines.

3.10 Q-FEVER

3.10.1 General evaluation of the national situation

3.10.1.1 Coxiella (Q-fever) - general evaluation

History of the disease and/or infection in the country

In 2014, the monitoring of bulk milk continued. The farms with milkgoats and milkewes were tested every 2 months. For cattle, sheep and goats, in case of abortion, samples are tested against a number of possible infectious agents including Coxiella burnetii. The circulation of Coxiella burnetii on cattle farms is known due to the presence of antibodies against Coxiella burnetii in the milk.

History of the disease and/or infection in the country

National evaluation of the recent situation, the trends and sources of infection

15 farms with milk goats had a RT-PCR positive result of their bulk milk in 2014. 8 of these farms already had at least one positive result in 2013. 3 farms with milkewes had positive RT-PCR results in 2014, 1 farm of its bulk milk and 2 farms after abortions.

National evaluation of the recent situation, the trends and sources of infection

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

A link between the presence of *Coxiella burnetii* on farms with goats, sheep or bovines and human cases has not been detected.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Recent actions taken to control the zoonoses

Milk from goats or sheep herds where *Coxiella burnetii* was found has to be pasteurized before human consumption. The location of positive herds is reported to the public health services for the purpose of warning the medical doctors.

Recent actions taken to control the zoonoses

3.11 CYSTICERCOSIS, TAENIOSIS

3.11.1 General evaluation of the national situation

3.11.1.1 Cysticerci - general evaluation

History of the disease and/or infection in the country

Cattle
Taenia saginata: 2002 total 3.336 (3.317 lightly, 18 heavily contaminated) 2003 total 3.886 (3.859 lightly, 25 heavily contaminated) 2004 total 3.002 (2.981 lightly, 21 heavily contaminated) 2005 total 2.392 (2.376 lightly, 16 heavily contaminated) 2006 total 1.824 (1.796 lightly, 28 heavily contaminated) 2007 total 1.527 (1.517 lightly, 10 heavily contaminated) 2008 total 2.374 (2.356 lightly, 18 heavily contaminated) 2009 total 1.820 (1.811 lightly, 9 heavily contaminated) 2010 total 1.766 (1.756 lightly, 10 heavily contaminated) 2011 total 1.347 (1.336 lightly, 11 heavily contaminated) 2012 total 1.214 (1.205 lightly, 9 heavily contaminated) 2013 total 994 (978 lightly, 16 heavily contaminated) 2014 total 1.172 (1.154 lightly, 18 heavily contaminated)
Pigs
The Belgian pig population is free from *Cysticercus cellulosae*. *Taenia solium* (and *Cysticercus cellulosae*) is not autochthonous in Belgium.

National evaluation of the recent situation, the trends and sources of infection

Cysticercus bovis in muscular tissue of cattle is the larval stage of the tapeworm, *Taenia saginata*, a parasitic cestode of the human gut (taeniasis). Cattle can become infected by ingestion of vegetation contaminated with *T. saginata* eggs shed in human faeces. Risk factors are access to rivers and flooding of pastures or wetland. Humans contaminate themselves by the ingestion of raw or undercooked beef containing the larval form (cysticerci). Usually pathogenicity for humans is low. The tapeworm eggs contaminate the environment directly or through surface waters. Human carriers should be treated promptly. Strict rules for the hygienic disposal or sanitation of human faeces with a method that inactivates *T. saginata* eggs should be developed. The spreading of human excrement on land should not be allowed.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Post-mortem, macroscopic examination of carcasses of adult cattle as well as calves is routinely done in all slaughterhouses. Serological examination is possible and confirmation of the lesions by PCR or DNA-test can be done. Lightly contaminated carcasses are treated by freezing at -18C for 10 days before declared fit for human consumption. Heavily contaminated carcasses are unfit for human consumption and are destroyed.

Suggestions to the European Union for the actions to be taken

The introduction of serological analyzes for the detection of cysticerci antigens in the serum of animals (cattle) should be developed. This would allow the detection of more cases than by visual inspection of carcasses at slaughterhouse.

3.12 SARCOCYSTOSIS

3.12.1 General evaluation of the national situation

3.12.1.1 Sarcocystis - general evaluation

History of the disease and/or infection in the country

At the slaughterhouses, a small number of carcasses showing myositis eosinophilica (green coloring spots of the carcass) are detected and notified to the Federal Agency for the Safety of the Food Chain. In case of positive findings, carcasses are totally rejected and declared unfit for human consumption. In 2010, 2011, 2012, 2013 and 2014 respectively 37, 44, 60, 75 and 94 cases of sarcosporidiosis in cattle were reported.

National evaluation of the recent situation, the trends and sources of infection

Sarcocystis bovihominis (bovine as intermediate host) and Sarcocystis suihominis (porcine intermediate host) occur sporadically. Domestic carnivores are hosts of the adult stage. Humans can be a definitive host for sarcosporidiosis by ingestion of infected meat or excreted oocysts and develop symptoms like diarrhea, headache, eosinophilia, abortion, congenital disorder. For human sarcosporidiosis there is no immunity development. A majority of grazing animals are inapparent carriers of tissue cysts.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Carcasses are entirely condemned when myositis eosinophilica lesions are apparent. Myositis eosinophilica is commonly associated with sarcosporidiosis but this is still not proven!

3.13 WEST NILE VIRUS INFECTIONS

3.13.1 West Nile Virus in animals

3.13.1.1 West Nile Virus in animal

Monitoring system

Sampling strategy

A surveillance of 'free range' domestic poultry and wild birds was organized based on the surveillance program of Avian Influenza since 2010. Blood samples of 1680 domestic poultry and 634 wild birds were all negative by IgG ELISA. Virological analyses of 221 pools of live wild birds (tracheal swabs) and 254 pools of intestins and brains of death wild birds were all negative by RRT-PCR. No surveillance of horses and bovines was organized in 2014.

Type of specimen taken

Blood
Oropharyngeal swabs
Tracheal swabs
Brain (CNS)
Intestin

Diagnostic/analytical methods used

IgG ELISA
Seroneutralisation test for confirmation
Reverse Real Time PCR

3.14 ESCHERICHIA COLI, NON-PATHOGENIC

3.14.1 General evaluation of the national situation

3.14.1.1 Escherichia coli, non-pathogenic - general evaluation

History of the disease and/or infection in the country

National evaluation of the recent situation, the trends and sources of infection

Recent actions taken to control the zoonoses

3.14.2 Escherichia coli, non-pathogenic in foodstuffs

3.14.2.1 E.coli, non-pathogenic, unspecified in food

Monitoring system

Sampling strategy

The hygiene of slaughtering and cutting process is watched via the evaluation of the contamination of carcasses and cutting meat by indicators of faecal contamination.

Frequency of the sampling

Every week

Type of specimen taken

Meat

Methods of sampling (description of sampling techniques)

Broilers and laying hens carcasses are taken at slaughterhouses. At cutting plants about 200g samples of meat were taken.

Definition of positive finding

Action limits were established for every matrix.

Diagnostic/analytical methods used

ISO method was used to count E. coli in food.

Measures in case of the positive findings or single cases

Monitoring/Not favorable results are sent to the FASFC.

3.15 ENTEROCOCCUS, NON-PATHOGENIC

3.15.1 Enterococcus, non-pathogenic in animals

3.15.1.1 Enterococcus spp., unspecified in animal

National evaluation of the recent situation, the trends and sources of infection

The antimicrobial resistance of non-pathogenic enterococci was monitored for the first time in 2011 in poultry, pigs and bovines. There was a high level of resistance in all species. However resistance in strains from bovine origin is lower compared to the strains from pigs and poultry.

3.16 TOXOPLASMA

3.16.1 General evaluation of the national situation

3.16.1.1 Toxoplasma - general evaluation

History of the disease and/or infection in the country

The majority of grazing animals seem to be inapparent carriers of tissue cysts.

Relevance of the findings in animals, feedingstuffs and foodstuffs to human cases (as a source of infection)

Man is infected with *Toxoplasma gondii* through ingestion of undercooked infected meat or upon accidental ingestion of sporulated oocysts from the environment. The cat is the final host, man and most warm-blooded animals are intermediate hosts. Most infections with *T. gondii* are asymptomatic, however mild (flu-like symptoms), moderate (lymphadenopathy, chronic fatigue) to severe disease (disseminated toxoplasmosis, encephalitis) may occur, the latter mainly in immunocompromised hosts. Moreover, when infection occurs in pregnant women, toxoplasmosis may cause abortion and congenital disorders. If a woman acquires primary infection during pregnancy, *Toxoplasma* can be transmitted through the placenta to the foetus and lead to congenital toxoplasmosis. A percentage of young children (1 to 14-year-old age group) may get post-natal infections with *T. gondii* and develop symptomatic toxoplasmosis (e.g. ocular disease). A number of cases of the disease in a 15 to 24-year-old age group may be referred to as acquired toxoplasmosis in immunocompetent patients, which may present a wide range of clinical signs, from lymphadenopathy to retinitis and uveitis. Immunocompetent individuals may often develop clinical toxoplasmosis. The majority of adult persons have acquired a degree of immunity to re-infection but can remain carrier.

Recent actions taken to control the zoonoses

Screening for toxoplasmosis during pregnancy is common. The seroprevalence in women tested before pregnancy is about 50%. Prevention of congenital toxoplasmosis by specific hygienic measures seems to have limited impact.

4 ANTIMICROBIAL RESISTANCE INFORMATION ON SPECIFIC ZOOSES AND ZONOTIC AGENTS

4.1 SALMONELLOSIS

4.1.1 Salmonella in foodstuffs

4.1.1.1 Antimicrobial resistance in Salmonella Meat from pig

Sampling strategy used in monitoring

Frequency of the sampling

Sampling distributed evenly throughout the year

Type of specimen taken

carcasses and cut meat

Procedures for the selection of isolates for antimicrobial testing

All strains isolated during the zoonosis monitoring program were sent to the Institute of Public Health for serotyping and determination of antimicrobial resistance. Since 2011, AMR was performed on the most prevalent ten serotypes.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre EUVSEC panel, as described in the EU-directive of 13 november 2013. The antimicrobials reported as well as the breakpoints for interpretation are listed in the table below. Quality control was performed by using an Escherichia coli ATCC 25922 strain. First panel EUVSEC Antimicrobienne ECOFF (R> mg/l) Ampicilline 8 Cefotaxime 0.5 Ceftazidime 0.5 Mropnme 0.125 Acide Nalidixique 16 Ciprofloxacin 0.064 Tetracycline 8 Colistine 2 Gentamicine 2 Trimethoprim 2 Sulfamthoxazole 256 Chloramphenicol 16 Azithromycine 16 Tigecycline 1 Second panel EUVSEC2 Antimicrobienne ECOFF* (R>mg/l) Cfoxitin 8 Cfpime 0.125 Cefotaxime+clavulanic acid >0.5 ceftazidime+clavulanic acid >2 Mropnme 0.125 Temocilline 32 Imipenem 0.5 Ertapenem 0.06 Cefotaxime 0.25 ceftazidime 0.5

Cut-off values used in testing

The cut-off values were used as described in the European Decision of 12 november 2013 on the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria.

Results of the investigation

In total 175 Salmonella strains recovery on 2014 from pork, were tested for their antimicrobial susceptibility. This includes strains from carcasses and cut meats. The resistance to cefotaxime, ceftazidime, and gentamicin remains between 0-1% since 2010. Resistance to quinolones varies between 4-6% and remains stable since 2010 without important variations, however, resistance to ampicillin reduced to 40% in 2013, has increased slightly and reached same values that in 2011, 52%. Unlikely, tetracycline resistance that had increased in 2012% from 28% in 2011 to 41% in 2012 and to 36% in 2013 is still increasing to 43% in 2014. Resistance to the trimethoprim remains stable (24%) as in 2013 (26%) after an important increase of 13% observed in 2012 vs 2011. The percentage of strains sensible to all antibiotics in 2014 was 30.86% vs 45% in 2013 which represents a decrease of 14%. However the multiresistance, the resistance of the isolates to 3 or more antibiotic's family have increased from 36% in 2013 to 41.14% in 2014.

4.1.1.2 Antimicrobial resistance in Salmonella Meat from poultry, unspecified

Sampling strategy used in monitoring

Frequency of the sampling

Sampling distributed evenly throughout the year

Type of specimen taken

carcasses from broilers and spent hens, chicken parts, and meat preparation

Procedures for the selection of isolates for antimicrobial testing

All strains isolated during the zoonosis monitoring program were sent to the Institute of Public Health for serotyping and determination of antimicrobial resistance.

Methods used for collecting data

Since 2011, the AMR was performed on the most prevalent ten serotypes.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre EUVSEC and EUVSEC2 panels, as described in the EU-directive of 13 november 2013. The antimicrobials reported as well as the breakpoints for interpretation are listed in the tables below. Quality control was performed by using an *Escherichia coli* ATCC 25922 strain. First panel EUVSEC Antimicrobienne Ampicilline Cefotaxime Ceftazidime Mropnme Acide Nalidixique Ciprofloxacin Tetracycline Colistine Gentamicine Trimthoprim Sulfamthoxazole Chloramphnicol Azithromycine Tigcycline Second panel EUVSEC2 Antimicrobien Cfoxitin Cfpime Cefotaxime+calvulanic acid ceftazidime+clavulanic acid Mropnme Temocilline Imipnm Ertapnme Cfoxitine ceftazidime

Cut-off values used in testing

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre EUVSEC panel, as described in the EU-directive of 13 november 2013. The antimicrobials reported as well as the breakpoints for interpretation are listed in the table below. Quality control was performed by using an *Escherichia coli* ATCC 25922 strain. First panel EUVSEC Antimicrobienne ECOFF (R> mg/l) Ampicilline 8 Cefotaxime 0.5 Ceftazidime 0.5 Mropnme 0.125 Acide Nalidixique 16 Ciprofloxacin 0.064 Tetracycline 8 Colistine 2 Gentamicine 2 Trimthoprim 2 Sulfamthoxazole 256 Chloramphnicol 16 Azithromycine 16 Tigcycline 1 Second panel EUVSEC2 Antimicrobien ECOFF* (R>mg/l) Cfoxitin 8 Cfpime 0.125 Cefotaxime+calvulanic acid >0.5 ceftazidime+clavulanic acid >2 Mropnme 0.125 Temocilline 32 Imipnm 0.5 Ertapnme 0.06 Cfoxitine 0.25 ceftazidime 0.5

Results of the investigation

Antimicrobial resistance in strains isolated from neck skin from broiler carcasses (EFSA-specific monitoring) During 2014, 81 *Salmonella* isolates, from poultry EFSA specification (neck skin) were tested for their antimicrobial susceptibility. Of them, 35 belonged to the serotype Paratyphi B var. Java. Of the total of salmonella tested, 3 were resistant to cefotaxime, 2 were resistant to cefotaxime and ceftazidime three were resistant to cefotaxime and meropenem and one was resistant to cefotaxime, ceftazidime and meropenem. In total 9 were β -lactamases producers and 6 of them belonged to the serotype Paratyphi B var. Java. Related to the other antimicrobials tested, the highest resistance noticed was to trimetoprim (53%) followed by quinolones and sulfamethoxazole (44%). Antimicrobial resistance in strains isolated from poultry meat. In 2014, 70 *Salmonella* isolates were tested for resistance to antimicrobials. The predominant serotype found was Enteritidis. The most important finding in 2014 was the high increase of colistin resistance detected. After a decreased in 2013 to 15%, in 2014 30% of the isolates were resistant to this antibiotic. Interestingly, all but two of the colistine resistant isolates belonged to the serotype Enteritidis and were recovered from spent hens. In addition to that, the resistance to tetracyclin was also increased 15% more than last year, reaching 22%. The majority of the isolates were recovered from poultry meat. The presence of β -lactamase producer isolates was 2.86%, 3 isolates were resistant to cefotaxime or ceftazidime, belonged to the serotype Paratyphi B and were isolated from poultry meat. No isolates resistant to meropenem were detected. The level of multidrug resistance (R to = >3 antibiotic's family) was 34.29%, and 32.86% of the isolates were sensitive to all antibiotics tested.

4.1.1.3 Antimicrobial resistance in *Salmonella* spp.

Sampling strategy used in monitoring

Frequency of the sampling

Strains of *Salmonella enterica* isolated during the zoonosis monitoring program were sent to the Scientific Institute of Public Health for serotyping and determination of antimicrobial resistance. Different food matrices were sampled, mainly poultry (carcasses from broilers and spent hens, chicken parts and meat preparations) and pork (carcasses and cut meats). Other matrices where *Salmonella* was isolated were ready-to-eat meals, meat, meat preparations, frogs legs, pudding, liquid egg product, ham and dry sausage. Since 2011, the AMR was performed on the most prevalent ten serotypes.

Methods of sampling (description of sampling techniques)

Procedures for the selection of isolates for antimicrobial testing

Laboratory methodology used for identification of the microbial isolates

Minimum Inhibitory Concentrations (MIC) were determined by the broth dilution method using Sensititre, as described in the EU-legislation Official Journal of the European Union, Commission implementing decision of 12 november 2013 on the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria. Interpretation was according to the EU-legislation. Quality control was performed by using an *Escherichia coli* ATCC 25922 strain.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

The antimicrobials tested are listed in the following table. Ampicilline Cfotaxime Ceftazidime Mropnme Acide Nalidixique Ciprofloxacin
Ttracycline Colistine Gentamicine Trimthoprime Sulfamthoxazole Chloramphnicol Azithromycine Tigcyclyne

Cut-off values used in testing

Minimum Inhibitory Concentrations (MIC) were determined by the use of broth microdilution, first panel Sensititre EUVSEC. The determination of the *Salmonella* spp. to third generation cephalosporins or meropenem has been done with the second panel, EUVSEC2. The antimicrobials of the first panel EUVSEC, tested and the breakpoints used are listed in the following table. Antimicrobienne ECOFF (R> mg/l)
Ampicilline 8 Cfotaxime 0.5 Ceftazidime 0.5 Mropnme 0.125 Acide Nalidixique 16 Ciprofloxacin 0.064 Ttracycline 8 Colistine 2 Gentamicine 2
Trimthoprime 2 Sulfamthoxazole 256 Chloramphnicol 16 Azithromycine 16 Tigcyclyne 1 The antimicrobials of the second panel EUVSEC2,
tested and the breakpoints used are listed in the following table. Antimicrobienne ECOFF* (R>mg/l) Cfoxitin 8 Cfpime 0.125
Cefotaxime+clavulanic acid 0.5 ceftazidime+clavulanic acid 2 Mropnme 0.125 Temocilline 32 Imipnm 0.5 Ertapnme 0.06 Cfotaxime 0.25
ceftazidime 0.5

Results of the investigation

Additional information

For specific results in foodstuff derived from poultry and porc, see the corresponding sections.

4.1.2 *Salmonella* in animals

4.1.2.1 Antimicrobial resistance in *Salmonella* Cattle (bovine animals)

Sampling strategy used in monitoring

Type of specimen taken

Clinical investigations, laboratory findings of the NRL *Salmonella*, animal health.

Methods of sampling (description of sampling techniques)

Control program/mechanisms

The control program/strategies in place

There was no monitoring program for Salmonella in cattle in 2014.

Results of the investigation

4.1.2.2 Antimicrobial resistance in Salmonella Pigs

Sampling strategy used in monitoring

Type of specimen taken

Methods of sampling (description of sampling techniques)

Results of the investigation

4.1.2.3 Antimicrobial resistance in Salmonella Poultry, unspecified

Description of sampling designs

Only Salmonella isolates obtained in the framework of the National Salmonella Control Programmes in broilers, meat turkeys and layers are used for the evaluation of the antimicrobial resistance in Salmonella in poultry.

Sampling strategy used in monitoring

Frequency of the sampling

The sampling strategy in poultry is explained in the different chapters on Salmonella in layers, broilers and meat turkeys.

Type of specimen taken

Methods of sampling (description of sampling techniques)

Procedures for the selection of isolates for antimicrobial testing

All Salmonella isolates obtained in the framework of the National Salmonella Control Programmes in layers and meat turkeys are selected for antimicrobial testing. In broilers, the first 170 isolates obtained are tested.

Methods used for collecting data

In the framework of the National Salmonella Control Programme, all laboratories involved in the detection of Salmonella gather the requested information concerning the sample and the sampled flock. All information is reported monthly to the Federal Agency for the Safety of the Food Chain together with the result of the analyses. All Salmonella isolates are sent to the NRL (CODA-CERVA) for serotyping. A fixed motive that indicates if AMR tests are required must accompany the isolate.

Laboratory methodology used for identification of the microbial isolates

The laboratory methods laid down in decision 2013/652/EU are used.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

All isolates from poultry are tested in the NRL (CODA/CERVA).

Cut-off values used in testing

The cut-off values described in decision 2013/652/EU are used.

Preventive measures in place

Treatment of *Salmonella* spp. with antibiotics is forbidden.

Control program/mechanisms

The control program/strategies in place

There are no mandatory control strategies in place.

Measures in case of the positive findings or single cases

There are no measures related to the finding of antimicrobial resistance in *Salmonella* in poultry.

Results of the investigation

In broilers, 167 isolates were tested on their antimicrobial resistance. All of the 21 isolates of *S. Paratyphi* B var. Java were resistant to at least one tested antimicrobial, 95,2% was resistant against more than 3 tested antibiotics (multiresistant). 46,7% of the *S. Enteritidis* isolates were resistant to at least one antimicrobial, none of the isolates were multiresistant. Multiresistance was also not seen in isolates of *S. Livingstone* and *S. Minnesota*. In layers, 45 isolates were tested. Of the 20 *S. Enteritidis* isolates, 15 were resistant against colistin. No other resistance was seen. Of the 9 *S. Infantis* isolates, 8 were resistant to trimethoprim. One isolate was also resistant to sulphonamide. All 4 *S. Typhimurium* isolates were resistant to trimethoprim, 3 of them also to ampicillin and 2 of those also to sulfonamide.

National evaluation of the recent situation, the trends and sources of infection

56% of the isolates of *S. Minnesota* showed some resistance in 2013 where in 2014, all strains were resistant to at least one antimicrobial. The same trend is seen for *S. Livingstone* where only 18% of the strains were resistant in 2013 and 90% in 2014.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

79,6% of the isolates from broilers were resistant to trimethoprim. This was also the antimicrobial with the greatest resistance (54%) in *Salmonella* isolates from samples taken in the slaughterhouse for the verification of the process hygiene criterium.

4.1.2.4 Antimicrobial resistance in *Salmonella* spp. All animals

Sampling strategy used in monitoring

Methods used for collecting data

Laboratory methodology used for identification of the microbial isolates

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Cut-off values used in testing

Results of the investigation

Additional information

4.2 CAMPYLOBACTERIOSIS

4.2.1 Campylobacter in foodstuffs

4.2.1.1 Antimicrobial resistance in Campylobacter jejuni and coli in foodstuff derived from Meat from pig

Sampling strategy used in monitoring

Procedures for the selection of isolates for antimicrobial testing

All strains isolated in the zoonosis monitoring program and originating from pork were sent to the Institute of Public Health for determination of antimicrobial resistance.

Laboratory methodology used for identification of the microbial isolates

Specification (coli/jejuni) with PCR (Debruyne et al, Res Microbiol, 2008)

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by using broth microdilution method (Sensititre EUCAMP panel). From 2014, a new European decision on the harmonization of the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria is adopted which specifies new interpretative threshold for resistance for *C. jejuni* and *C. coli*. Therefore, the antimicrobials tested and the epidemiological cut-off values (ECOFF) used are listed in the table below following the official Journal of the European Union (L303/26 14.11.2013). In order to compare results in an accurate way, recalculation of resistance to the antibiotics using the new breakpoint established in the European decision was done for the last four years, from 2010 to 2013 included. Antimicrobial Breakpoints R > (g / ml) *C. jejuni* *C. coli* Tetracycline 12 Nalidixic acid 16 Ciprofloxacin 0.50.5 Erythromycin 48 Gentamicin 22 Streptomycin 44 Campylobacter in meat and meat products: list of antimicrobials tested and breakpoints used.

Results of the investigation

In total, 17 Campylobacter isolates were analysed, in 2014, which belonged to *C. coli* (16) and *C. jejuni* (1). All the isolates tested keep a resistance to one or more antibiotics. The highest level of resistance was found for streptomycin and tetracycline, 87.5% and 81% respectively. Resistance to quinolones stops to increase and now a decrease was noticed against 2013 with 37.5% resistance. The resistance against erythromycin increased to 31% of the isolates compared to 17% in 2013. One isolate was resistant to 4 classes of antibiotics. Pork 2014 *C. coli* (n=16) Tetracycline 81 Ciprofloxacin 37.5 Nalidixic acid 37.5 Gentamicin 0 Erythromycin 31 Streptomycin 87.5 Multiresistance in *C. coli* isolated from pork. *C. coli* in pork (n=16) 2014 n% Sensible 001318.752531.253742.75416.25

4.2.1.2 Antimicrobial resistance in *Campylobacter jejuni* and *coli* in foodstuff derived from Meat from poultry, unspecified

Sampling strategy used in monitoring

Procedures for the selection of isolates for antimicrobial testing

In 2014, 362 *Campylobacter* strains isolated in the zoonoses monitoring programme and originating from poultry, (carcasses of broilers, filets, entrails, meat preparation and carcasses of spent hens)

Laboratory methodology used for identification of the microbial isolates

Specification (*coli/jejuni*) with PCR (Denis et al, 2001)

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by using broth microdilution method (Sensititre EUCAMP panel). From 2014, a new European decision on the harmonization of the monitoring and reporting of antimicrobial resistance in zoonotic and commensal bacteria is adopted which specifies new interpretative threshold for resistance for *C. jejuni* and *C. coli*. Therefore, the antimicrobials tested and the epidemiological cut-off values (ECOFF) used are listed in the table below following the official Journal of the European Union (L303/26 14.11.2013). The European decision has not included the monitoring of chloramphenicol any more. In order to compare results in an accurate way, recalculation of resistance to the antibiotics using the new breakpoint established in the European decision was done for the last four years, from 2010 to 2013 included. Antimicrobial Breakpoints R > (g / ml) *C. jejuni* *C. coli* Tetracycline 12 Nalidixic acid 16 Ciprofloxacin 0.50.5 Erythromycin 48 Gentamicin 22 Streptomycin 44 *Campylobacter* in meat and meat products: list of antimicrobials tested and breakpoints used.

Results of the investigation

In 2014, 362 *Campylobacter* strains isolated from poultry meat and carcasses were tested for antimicrobial susceptibility. In total 75 isolated were identified as *C. coli* and 286 as *C. jejuni*. One isolate was just identified to be *Campylobacter* species. Overall, resistance to quinolones was present in 49% of the strains and also tetracycline resistance was high 44%. For *C. jejuni*, 25% of all strains were sensitive to all antibiotics tested, which is a decrease compared to 2013. The resistance against tetracycline and quinolones remained high, and increased again to respectively 51.3% and 57.3%. Resistance to streptomycin, erythromycin and gentamycin remains low (< 1%).

Antibiotic	2014	2013
Tetracycline	51.33%	51.33%
Ciprofloxacin	57.33%	57.33%
Nalidixic acid	56.63%	56.63%
Gentamicin	0.30%	0.30%
Erythromycin	0.31%	0.31%
Streptomycin	0.31%	0.31%

The overall resistance of *Campylobacter* is detailed in the following tables. *C. jejuni* in poultry meat 2014 2013

Antibiotic	2014	2013
Tetracycline	51.33%	51.33%
Ciprofloxacin	57.33%	57.33%
Nalidixic acid	56.63%	56.63%
Gentamicin	0.30%	0.30%
Erythromycin	0.31%	0.31%
Streptomycin	0.31%	0.31%

The trends in antimicrobial resistance are stable since 2010, no resistance to gentamicin has been observed and very low to erythromycin following by streptomycin. The highest resistance is founded for ciprofloxacin and nalidixic acid followed by tetracycline.

4.2.2 *Campylobacter* in animals

4.2.2.1 Antimicrobial resistance in *Campylobacter jejuni* and *coli* in Pigs

Sampling strategy used in monitoring

Frequency of the sampling

Campylobacter strains isolated from pork were tested for antimicrobial susceptibility.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

Minimum Inhibitory Concentrations (MIC) were determined by using broth microdilution method (Sensititre EUCAMP panel). The antimicrobials tested and the breakpoints (following the CLSI standards) used are listed in the table below.

Antimicrobial	Breakpoints (g / ml)
jejunicola	16
Chloramphenicol	16
Tetracycline	22
Nalidixic acid	32
Ciprofloxacin	11
Erythromycin	416
Gentamicin	12
Streptomycin	24

Results of the investigation

4.2.2.2 Antimicrobial resistance in *Campylobacter jejuni* and coli in Poultry, unspecified

Description of sampling designs

The number of samples was calculated based on the prevalence in recent years of *Campylobacter* in broilers. The samples were equally divided per month over the different provinces based on the number of slaughterhouses and the capacity of the slaughterhouses in the province.

Sampling strategy used in monitoring

Frequency of the sampling

The monitoring is performed every two years spread over a year.

Laboratory methodology used for identification of the microbial isolates

An ISO 10272-1 based method is used for the direct detection of *Campylobacter* in faeces. A PCR according to the publication by Denis et al (2001) is used to confirm *Campylobacter jejuni*.

Type of specimen taken

10 caeca are taken for each batch that is sampled. The caeca are pooled to one sample.

Methods of sampling (description of sampling techniques)

Samples are taken on the main slaughter line.

Stratification procedures per animal populations and food categories

Samples were taken in those slaughterhouses with the largest capacities.

Randomisation procedures per animal populations and food categories

When and where samples were taken in the course of a month within a province was decided by the sampler.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

The monitoring of AMR is performed according to decision 2013/652/EU.

Cut-off values used in testing

The cut-off values used are those published in decision 2013/652/EU.

Results of the investigation

Of the 304 samples taken, it was possible to test 92 isolates of *Campylobacter jejuni* for antimicrobial resistance. 33% of the isolates was not resistant to any tested antimicrobial. Only one isolate was multiresistant. The most frequently found resistance was against ciprofloxacin (60%) and tetracycline (52%).

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

57% of the *Campylobacter jejuni* isolates from poultry carcasses and meat were resistant to ciprofloxacin, 52% to tetracycline. These high prevalences were also found in isolates from caeca.

4.3 ESCHERICHIA COLI, NON-PATHOGENIC

4.3.1 *Escherichia coli*, non-pathogenic in animals

4.3.1.1 Antimicrobial resistance in *E. coli*, non-pathogenic, unspecified

Description of sampling designs

Each year, the AMR of commensal *E. Coli* is monitored in poultry, pigs and bovines. The number of samples taken is calculated based on the detection percentage of commensal *E. Coli*. The samples of poultry, pigs and meat calves are taken at the slaughterhouse. The samples of young bovines at the farm.

Sampling strategy used in monitoring

Frequency of the sampling

The samples are taken all year round.

Type of specimen taken

Faecal samples are taken from the rectum at the slaughterhouse, pooled faecal samples representing 10 animals are taken at farm level.

Stratification procedures per animal populations and food categories

There is a stratified sampling by province proportionally to the number and capacity of the slaughterhouses present in each province or to the number of registered farms for the specific category.

Randomisation procedures per animal populations and food categories

The sampler at the level of the province decides which farms will be sampled. At the slaughterhouse, the official vet decides which batches will be sampled.

Procedures for the selection of isolates for antimicrobial testing

The isolates are selected at random.

Methods used for collecting data

The requested data is collected by the official veterinarian who takes the samples and centralized electronically in a database of the Federal Agency for the Safety of the Food Chain (FASFC). This central database is also fed by the laboratories that perform the detection. Data of further analyses by the NLR is reported on a frequent basis to the FASFC.

Laboratory methodology used for identification of the microbial isolates

Two methods are used, depending on the laboratory. At DGZ faecal material was inoculated on McConkey agar and incubated at 37C for 18 to 24 hours. Suspected colonies (pink, lactose positive) were inoculated on Kligler and indol medium and incubated at 37C for 18 to 24 hours. When the test outcome was positive for E. coli a colony from the Kligler medium was inoculated on Mac Conckey agar, incubated at 37C for 18-24 hours. At ARSIA, faecal material was inoculated on Gassner medium and incubated at 37C for 18 to 24 hours. Suspected colonies were purified on Columbia agar supplemented with 5% sheep blood. Identification was done by the OPNG test, Ureum test and indol test. E. coli strains were sent to CODA-CERVA and were confirmed by MALDITOF.

Laboratory used for detection for resistance

Antimicrobials included in monitoring

The monitoring is performed according to decision 2013/652/EU.

Cut-off values used in testing

The cut-offs according to decision 2013/652/EU are used.

Preventive measures in place

There are no mandatory preventive measures in place.

Control program/mechanisms

The control program/strategies in place

There is no specific control strategy for farms/batches where AMR is found. The general control strategy is based on supporting and stimulating the sector to take initiative to reduce the use of antimicrobials in general.

Recent actions taken to control the zoonoses

The sector initiated an action plan consisting of 3 strategic targets for the reduction of the use of antimicrobials in general and more specific of the use of critical important antibiotics and medicated feed. The action plan also holds 7 operational targets.

Measures in case of the positive findings or single cases

There are no mandatory measures in case of positive findings.

Notification system in place

There is no mandatory notification of AMR. The results of the official monitoring are reported on a frequent basis.

Results of the investigation

158 isolates from poultry were tested. Resistance against colistin was not detected. The most common resistances were against ampicillin (72,8%) and ciprofloxacin (69,6%). A low number of strains were resistant to gentamycin (5,7%) and ceftazidime (7,6%). 88,6 % of the isolates were resistant against at least one tested antimicrobial, 58% of the isolates were resistant against more than 3 tested antimicrobials. 184 isolates from pigs were tested. Main resistance was seen against sulphonamide (52,2%), trimethoprim (50%), tetracycline (44%), ampicillin (41,3%) and chloramphenicol (28,8%). Only minor resistance was seen against the other tested antimicrobials. 71,2% of the isolates was resistant against at least one tested antimicrobial, 33,1% against more than 3 tested antimicrobials. 164 isolates from beef cattle were tested. Main resistance was found against sulphonamide (23,8%), ampicillin (20,7%), tetracycline (18,3%), chloramphenicol (15,9%) and trimethoprim (15,2%). 28,7% of the isolates was resistant against at least one tested antimicrobial, 18,3% against more than 3 tested antimicrobials. 188 isolates from veal calves were tested. Main resistance was found against tetracycline (68,1%), sulphonamide (57,5%), ampicillin (54,8%) and trimethoprim (51,1%). 73,9% of the isolates was resistant against at least one tested antimicrobial, 54,3% against more than 3 tested antimicrobials.

National evaluation of the recent situation, the trends and sources of infection

Antimicrobial resistance in general and multiresistance is mainly found in isolates of poultry and veal calves. However, a statistically significant decreasing trend is seen in these two populations in the last two years. In poultry there was also a decreasing trend in the prevalence of ESBL-producing E. Coli over the past 2 years. AMR in isolates from beef cattle is significant lower than the other tested populations.

Relevance of the findings in animals to findings in foodstuffs and to human cases (as a source of infection)

There is also a decrease in the prevalence of ESBL-producing E. Coli on poultry carcasses at the level of the slaughterhouse.

5 INFORMATION ON SPECIFIC MICROBIOLOGICAL AGENTS

5.1 CRONOBACTER

5.1.1 Cronobacter in foodstuffs

5.1.1.1 Cronobacter in food

Monitoring system

Sampling strategy

Tests for *Cronobacter sakazakii* were performed in foodstuff intended for special nutritional uses, infant formula and milk (prepared milk in bottles for infants and young children).

Frequency of the sampling

Samples are taken according to the national control program or in the frame of RASFF, complaints or suspicion.

Type of specimen taken

Foodstuff intended for special nutritional uses (infants), infant formula and milk (infants)

Methods of sampling (description of sampling techniques)

The samples were taken according to Regulation (EC) No 2073/2005.

Definition of positive finding

To determine the conformity of a sample or a batch, the criteria laid down in the Regulation (EC) No 2073/2005 are applied.

Diagnostic/analytical methods used

The method is used according to Regulation (EC) No 2073/2005.

Measures in case of the positive findings or single cases

Measures to be taken in the case of a non-compliant result:- Notification of the producer or importer- Possibility of a counter analysis- Destruction of the non compliant batch or single sample- Further investigation: additional sampling, possible recall, RASFF, ...

5.2 HISTAMINE

5.2.1 Histamine in foodstuffs

5.2.1.1 Histamine in food

Monitoring system

Sampling strategy

Frequency of the sampling

Type of specimen taken

Methods of sampling (description of sampling techniques)

Definition of positive finding

Diagnostic/analytical methods used

Preventive measures in place

Control program/mechanisms

The control program/strategies in place

The annual control plan foresees controls of histamine in fishery products with high levels of histidine.

Measures in case of the positive findings or single cases

National evaluation of the recent situation, the trends and sources of infection

5.3 STAPHYLOCOCCAL ENTEROTOXINS

5.3.1 Staphylococcal enterotoxins in foodstuffs

5.3.1.1 Staphylococcal enterotoxins in food

Monitoring system

Sampling strategy

Tests of Staphylococcal enterotoxins were performed in samples with more than 10(5) cfu/g of Staphylococcus present.

Frequency of the sampling

Samples are taken according to the national control program or in the frame of RASFF, complaints or suspicion.

Type of specimen taken

cheeses, RTE, ...

Methods of sampling (description of sampling techniques)

The samples were taken according to Regulation (EC) No 2073/2005.

Definition of positive finding

To determine the conformity of a sample or a batch, the criteria laid down in the Regulation (EC)No 2073/2005 are applied. For products for which no legal criteria exist, a table of action limits is made available for the inspectors. (cfr. rt. 14, Reg 178/2002)

6 FOODBORNE OUTBREAKS

Foodborne outbreaks are incidences of two or more human cases of the same disease or infection where the cases are linked or are probably linked to the same food source. Situation, in which the observed human cases exceed the expected number of cases and where a same food source is suspected, is also indicative of a foodborne outbreak.

6.1 Outbreaks

6.1.1 Foodborne outbreaks

System in place for identification, epidemiological investigations and reporting of foodborne outbreaks

In Belgium different authorities are dealing with food-borne outbreaks:-The Federal Agency for the Safety of the Food chain FASFC deals with safety of foodstuffs, epidemiological investigation on foodstuffs and animal health issues in case of a food-borne outbreak. -The Communities (Flemish, French and German speaking Community) are dealing with person related matters as human health and can start an epidemiological investigation by Public health medical inspectors in case of a food-borne outbreak. -The Scientific Institute of Public Health IPH (National Reference Laboratory on Food-borne Outbreaks) analyses all suspected food samples, collects all data on food-borne outbreaks and gives scientific support to the FASFC officers and the Public Health Inspectors. A national "Platform Food-borne outbreaks", approved by the National Conference of Ministers of Public Health, brings together the different competent authorities on food safety, animal health and public health. Furthermore in 2007, for a better communication, a protected web application was made available to exchange outbreak data and laboratory results in real time between the different authorities dealing with FBO. In this web-application a common file is created for each individual outbreak, and the data and laboratory results are shared between food inspectors and human health inspectors. Data in this report came from the Federal Agency for the Safety of the Food Chain, the Public Health Inspection, the sentinel laboratories network for human microbiology, and the Federal Reference Centres for Food-borne outbreaks, for *Clostridium botulinum*, for *Salmonella* and *Shigella* and for *Listeria*.

Description of the types of outbreaks covered by the reporting:

A food-borne outbreak is defined as an incidence, observed under given circumstances, of two or more human cases of the same disease and/or infection, or a situation in which the observed number of human cases exceeds the expected number and where the cases are linked, or are probably linked, to the same food source (Directive 2003/99/EC, Article 2(d)). Data are collected from FASFC, the Flemish Community, the French community, the Brussels Common Community Committee, the sentinel laboratories network for human clinical microbiology, and the Federal Reference Centers for Food-borne outbreaks, *Salmonella* and *Shigella*, *Listeria* and *C. botulinum*. The reporting includes both general and household outbreaks. The causative agents covered are *Salmonella* spp., *Shigella* spp., *Campylobacter* spp., Verotoxigenic *E. coli*, *Listeria monocytogenes*, *Clostridium botulinum*, *Staphylococcus aureus*, *Bacillus cereus*, *Clostridium perfringens*, *Giardia*, *Norovirus*, enterotoxins of *Staphylococcus aureus* and *Bacillus cereus* and histamine

National evaluation of the reported outbreaks in the country:

Trends in numbers of outbreaks and numbers of human cases involved

During 2014, a total of 370 outbreaks of food-borne infections and intoxications and a human case of botulism were recorded in Belgium. More than 1789 people were ill, at least 63 persons were hospitalized. None of the human cases died. The number of reported outbreaks increased in 2011 as compared to former years but remains stable since then. The increase was probably due to an adapted Outbreak investigation procedure at the FASFC since 2011 and/or increased sensibility by consumers. The number of human cases involved are similar as in previous years which is also the case for the number of people hospitalized due to a collective food borne outbreak.

Relevance of the different causative agents, food categories and the agent/food category combinations

In 2014 in total 16 verified food borne outbreaks were reported and a case of human botulism. In these outbreaks the causative agent was found in the implicated food and/or it was clear by analytical epidemiology that food was at the origin of disease. All other outbreaks were classified as possible outbreaks where the agent was unknown or the agent could be only detected at human level. Bacterial toxins of *Bacillus cereus* was the most frequently detected causative agent in 11 outbreaks and was responsible for 46 human cases. Two outbreaks were caused by emetic toxin producing *Bacillus cereus* whereas enterotoxigenic *Bacillus cereus* was identified as causative agent for the remaining 9 outbreaks. Enterotoxigenic *Bacillus cereus* was also at the origin of a co-infection with coagulase positive *Staphylococcus*, where 3 persons became ill after the consumption of mixed foods. The second most reported agents were *Salmonella* and *Norovirus* with each being at the origin of 5 outbreaks. In total 80 persons became ill and 5 were hospitalized due to *Salmonella*. *Salmonella* Enteritidis was at the origin of 4 outbreaks involving eggs or egg products, including tiramisu and chocolate mousse. *Salmonella* Hadar was isolated from patients that consumed Turkey meat. *Norovirus* was at the origin of 5 outbreaks and was responsible for 275 human cases. In these outbreaks, *Norovirus* was transmitted by the food. For one of these outbreaks, the same *Norovirus* genotype was detected in both the food handlers and the human cases. This outbreak involved several clusters, including one cluster in a neighboring country. Coagulase positive *Staphylococcus* (CPS) was involved in 3 outbreaks, and 36 persons became ill, 11 others were also hospitalized. Enterotoxin A producing CPS were isolated from composed meals. Histamine was responsible for 2 outbreaks causing 4 ill people and 2 hospitalizations. Consumption of tuna fish could be linked to the outbreaks. One outbreak was caused by enterotoxinogenic *Clostridium perfringens* and caused diarrhea in 17 human cases. One human case was hospitalized. The pathogenic strain was isolated from fresh made rice balls with tomato sauce and vegetables. *E. coli* O157:H7 was at the origin of an outbreak involving 2 human cases and was probably due to the consumption of contaminated bovine meat. *Campylobacter* was linked to a single outbreak and caused diarrhea in 2 human cases after the consumption of hamburgers. *Listeria monocytogenes* 1/2a was isolated in steak tartare and was probably responsible for 2 human cases of which 1 was hospitalized. Consumption of wild boar contaminated with *Trichinella* led to a national outbreak involving 16 disseminated cases of which 14 persons were hospitalized. In 91% of the outbreaks (N=338 out of 370) no causative agent could be identified. An important reason for this is the absence of leftovers of the suspected meal in most of those outbreaks and late reporting by the consumer. Only in 38.6% (N=143 out of 370) of the outbreaks, samples (human and/or food) were sent for analysis of which 22% (N=32) resulted in the detection of a pathogen. Some of the latter outbreaks (N=17) have been categorized as a weak evidence outbreak. Most food-borne outbreaks (54.9%) were due to the consumption of meals composed of different ingredients. Meat and meat based products (bovine, pig, sheep, broiler) were responsible for 18.1 % of the outbreaks. In 10.5% of the outbreaks the implicated food was unknown.

Relevance of the different type of places of food production and preparation in outbreaks

Restaurants and take away or fast food outlets were the most important location of exposure, being the setting of 51.1 % and 19.7 %, respectively, of food-borne outbreaks in Belgium in 2014. Catering at work, institutional catering or temporary mass catering are reported in respectively 1.1 %, 1.1 and 4.3 % of the food-borne outbreaks. 18.1 % of the outbreaks happened at home.

Descriptions of single outbreaks of special interest

In 2014, 16 Belgian citizens aged between 18 and 51 years old suffered from Trichinosis. Cases suffered from high fever, muscle pains, fatigue and facial oedema. Fourteen of them were hospitalized. All human cases consumed wild boar meat in different restaurants spread within Belgium. Trace back analysis demonstrated that all wild boar meat originated from a single company in Spain. Several human cases suffering from diarrhoea and vomiting were reported over a period of 2 weeks on 8 different locations in Belgium and 1 location in a neighboring country. In total 210 persons became ill due to a *Norovirus* infection. All events had a same caterer in common. *Norovirus* was detected in the human cases but also the food handler and a personnel member resulted positive. Two different genotypes of *Norovirus* were found for 2 out of 4 food handlers (GII.P4 New Orleans 2009|ND and GIIP21|ND) of which one was confirmed in the human case, GIIP21|GII.3. Although all food samples resulted negative for *Norovirus*, epidemiological evidence links all events to the consumption of food delivered by the same company and confirms a transmission of *Norovirus* by the food. A case of human botulism type B was confirmed. The patient suffered from obstipation, dry mouth, troubled view. Consumption of lasagna in a foreign country was suspected to be at the origin of disease but could not be demonstrated.

Control measures or other actions taken to improve the situation

Logistic slaughtering is applied for poultry which means that poultry with a *Salmonella*-free certificate are slaughtered before other poultry. The vaccination of laying hens against salmonellosis, started in 2003 and is mandatory for *Salmonella* enteritidis and is strongly recommended for *Salmonella* typhimurium.

ANIMAL POPULATION TABLES

Table Susceptible animal population

Animal species	Category of animals	Population		
		holding	animal	slaughter animal (heads)
Cattle (bovine animals)	Cattle (bovine animals) - calves (under 1 year) - veal calves			336,281
	Cattle (bovine animals) (not specified)	30,138	2,615,471	501,189
Deer	Deer - farmed - fallow deer			846
	Deer - wild - fallow deer			13,492
	Deer (not specified)	2,466	7,063	
Ducks	Ducks (not specified)			46,070
Gallus gallus (fowl)	Gallus gallus (fowl) - breeding flocks, unspecified (not specified)		2,931,313	
	Gallus gallus (fowl) - broilers (not specified)		28,928,394	270,352,661
	Gallus gallus (fowl) - laying hens (not specified)		9,281,016	26,442,705
	Gallus gallus (fowl) (not specified)	1,514		297,881,136
Geese	Geese (not specified)			2,774
Goats	Goats (not specified)	10,290	45,871	6,354
Guinea fowl	Guinea fowl (not specified)			14,290
Partridges	Partridges (not specified)			14,152
Pheasants	Pheasants (not specified)			15,242
Pigeons	Pigeons (not specified)			152,112
Pigs	Pigs - breeding animals (not specified)		543,596	
	Pigs - fattening pigs (not specified)		5,339,118	
	Pigs (not specified)	8,027		11,888,367
Quails	Quails (not specified)			266
Rabbits	Rabbits (not specified)			3,114,169
Ratites (ostrich, emu, nandu)	Ratites (ostrich, emu, nandu) - farmed			342
Sheep	Sheep (not specified)	26,936	135,514	122,861
Solipeds, domestic	Solipeds, domestic (not specified)		270,149	8,337
Turkeys	Turkeys (not specified)			840,862
Wild boars	Wild boars - wild			11,264

DISEASE STATUS TABLES

Table Ovine or Caprine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Total number of herds	Number of infected herds	Number of herds with status officially free	Number of animals positive in microbiological testing under investigations of suspect cases	Number of animals tested by microbiology under investigations of suspect cases	Number of seropositive animals under investigations of suspect cases	Number of suspended herds under investigations of suspect cases	Number of animals serologically tested under investigations of suspect cases	Number of animals tested under surveillance	Total number of animals
Belgique-België	37,226	0	37,226	0	3	4	0	231	6,561	181,386

Table Bovine brucellosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Total number of herds	Number of infected herds	Number of herds with status officially free	Number of animals positive in microbiological testing under investigations of suspect cases	Number of animals tested by microbiology under investigations of suspect cases	Number of animals positive to BST under investigations of suspect cases	Number of seropositive animals under investigations of suspect cases	Number of suspended herds under investigations of suspect cases	Number of animals serologically tested under investigations of suspect cases	Number of abortions due to Brucella abortus	Number of isolations of Brucella infections	Number of notified abortions whatever cause	Number of infected herds tested under surveillance by bulk milk	Number of animals or pools tested under surveillance by bulk milk	Number of animals tested under surveillance	Number of herds tested under surveillance	Total number of animals
Belgique-België	30,138	0	30,138	0	61	0	36	18	1,227	0	0	10,577	9,090	17,869	96,030	13,403	2,615,471

DISEASE STATUS TABLES

Table Bovine tuberculosis in countries and regions that do not receive Community co-financing for eradication programme

Region	Total number of herds	Number of infected herds	Number of herds with status officially free	Number of animals detected positive in bacteriological examination	Number of animals with suspicious lesions of tuberculosis examined and submitted to histopathological and bacteriological examinations	Number of tuberculin tests carried out before the introduction into the herds	Number of animals tested with tuberculin routine testing	Interval between routine tuberculin tests	Total number of animals
Belgique-België	30,138	0	30,138	0	91	310,977	194,178	0	2,615,471

PREVALENCE TABLES

Table CAMPYLOBACTER in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Cheeses made from cows' milk - fresh - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	26	1	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Cheeses made from cows' milk - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	12	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Live bivalve molluscs - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	81	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	33	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	6	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	16	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	21	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	32	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals and pig - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	8	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals and pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	33	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals and pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	5	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from bovine animals and pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	9	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from broilers (Gallus gallus) - carcass - Slaughterhouse - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	545	119	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from other poultry species - carcass - Slaughterhouse - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	252	10	Campylobacter - Thermophilic Campylobacter spp., unspecified	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from pig - carcass - Slaughterhouse - Belgium - - Surveillance - Official sampling - Objective sampling	single	600	Colony forming unit/gram	558	33	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from pig - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	48	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from pig - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	2	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	69	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	9	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	14	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from poultry, unspecified - carcass - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	7	1	Campylobacter - C. coli	0
					6	Campylobacter - C. jejuni	0
				91	7	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from poultry, unspecified - fresh - Cutting plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	207	8	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from poultry, unspecified - fresh - Cutting plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	164	11	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from poultry, unspecified - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	13	1	Campylobacter - C. coli	0
					11	Campylobacter - C. jejuni	0
				64	13	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from poultry, unspecified - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	48	1	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from poultry, unspecified - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	110	1	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Meat from poultry, unspecified - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	16	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0
Milk, cows ¹ - raw milk - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Millilitre	41	0	Campylobacter - Thermophilic Campylobacter spp., unspecified	0

Table COXI ELLA (Q-FEVER) in animal

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	N of clinical affected herds	Zoonoses	N of units positive
Cattle (bovine animals) - Farm (not specified) - Belgium - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	1233	191	0	Coxiella (Q-fever) - C. burnetii	0
Cattle (bovine animals) - Farm (not specified) - Belgium - animal sample - blood - Monitoring - active - Industry sampling - Selective sampling	animal	4008	556	0	Coxiella (Q-fever) - C. burnetii	0
Cattle (bovine animals) - Farm (not specified) - Belgium - animal sample - foetus/stillbirth - Monitoring - active - Industry sampling - Selective sampling	animal	8902	104	3	Coxiella (Q-fever) - C. burnetii	0
Cattle (bovine animals) - Farm (not specified) - Belgium - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling	holding	29	13	0	Coxiella (Q-fever) - C. burnetii	0
Cattle (bovine animals) - Farm (not specified) - Belgium - animal sample - milk - Clinical investigations - Industry sampling - Suspect sampling	holding	59	50	0	Coxiella (Q-fever) - C. burnetii	0
Cattle (bovine animals) - Farm (not specified) - Belgium - animal sample - milk - Monitoring - active - Industry sampling - Selective sampling	holding	1	1	0	Coxiella (Q-fever) - C. burnetii	0
Cattle (bovine animals) - Farm (not specified) - Belgium - animal sample - milk - Monitoring - active - Industry sampling - Selective sampling	holding	191	161	0	Coxiella (Q-fever) - C. burnetii	0
Goats - milk goats - Farm (not specified) - Belgium - animal sample - milk - Monitoring - active - Official sampling - Census	holding	117	15	13	Coxiella (Q-fever) - C. burnetii	0
Sheep - milk ewes - Farm (not specified) - Belgium - animal sample - milk - Monitoring - active - Official sampling - Census	holding	18	1	3	Coxiella (Q-fever) - C. burnetii	0
Sheep and goats - Farm (not specified) - Belgium - animal sample - blood - Clinical investigations - Industry sampling - Suspect sampling	animal	56	6	0	Coxiella (Q-fever) - C. burnetii	0
Sheep and goats - Farm (not specified) - Belgium - animal sample - blood - Monitoring - active - Industry sampling - Selective sampling	animal	32	2	0	Coxiella (Q-fever) - C. burnetii	0
Sheep and goats - Farm (not specified) - Belgium - animal sample - foetus/stillbirth - Monitoring - active - Industry sampling - Selective sampling	animal	141	5	0	Coxiella (Q-fever) - C. burnetii	0
Sheep and goats - Farm (not specified) - Belgium - animal sample - placental swab - Monitoring - active - Industry sampling - Selective sampling	animal	12	2	1	Coxiella (Q-fever) - C. burnetii	0
Sheep and goats - Farm (not specified) - Belgium - animal sample (not specified) - Clinical investigations - Industry sampling - Suspect sampling	animal	2	0	0	Coxiella (Q-fever) - C. burnetii	0

Table CRONOBACTER in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	295	0	Cronobacter - Cronobacter sakazakii	0
Infant formula - dried - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	15	0	Cronobacter - Cronobacter sakazakii	0
Infant formula - dried - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	234	0	Cronobacter - Cronobacter sakazakii	0
Infant formula - ready-to-eat - Hospital or medical care facility - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Millilitre	129	0	Cronobacter - Cronobacter sakazakii	0

Table CYSTICERCI in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from bovine animals - carcase - Slaughterhouse - Belgium - - Surveillance - Official sampling - Suspect sampling	animal		NOT AVAILABLE	83747 0	1172	Cysticerci - Cysticerci of Taenia saginata	0

Table ECHINOCOCCUS in animal

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Cattle (bovine animals) - Slaughterhouse - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	83747 0	0	Echinococcus - E. granulosus	0
Foxes - wild - Natural habitat - Belgium - animal sample - organ/tissue - Monitoring - passive - Official sampling - Convenient sampling	animal	54	1	Echinococcus - E. multilocularis	0
		55	0	Echinococcus - E. multilocularis	0
		60	0	Echinococcus - E. multilocularis	0
		73	5	Echinococcus - E. multilocularis	0
		75	0	Echinococcus - E. multilocularis	0
		317	6	Echinococcus - E. multilocularis	0

Table ESCHERICHIA COLI, PATHOGENIC in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Cheeses made from cows' milk - fresh - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	22	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Cheeses made from cows' milk - fresh - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Cheeses made from cows' milk - soft and semi-soft - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	70	1	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O103 - eae positive	1
Cheeses made from cows' milk - soft and semi-soft - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	60	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Cheeses made from cows' milk - soft and semi-soft - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	88	1	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O26 - eae positive	1
Cheeses made from goats' milk - unspecified - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	44	1	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O103 - eae positive	1
Cheeses made from goats' milk - unspecified - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	16	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Cheeses made from goats' milk - unspecified - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	51	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Cheeses made from sheep's milk - unspecified - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Cheeses made from sheep's milk - unspecified - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Cheeses made from sheep's milk - unspecified - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	1	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC NT (Not Typeable)	1
Dairy products (excluding cheeses) - butter - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	97	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Dairy products (excluding cheeses) - cream - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	30	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Dairy products (excluding cheeses) - cream - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	23	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Fruits and vegetables - pre-cut - Border inspection activities - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Fruits and vegetables - pre-cut - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	6	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O104:H4 - EAggEC positive vtx2 positive	0
Fruits and vegetables - pre-cut - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	99	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Fruits and vegetables - pre-cut - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	98	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Meat from bovine animals - Border inspection activities - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	13	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from bovine animals - carcass - Slaughterhouse - Belgium - food sample - carcass swabs - Surveillance - Official sampling - Objective sampling	single	1600	Square centimetre	450	10	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O103 - eae positive	2
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O145 - eae positive	3
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O157:H7 - eae positive	1
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O26 - eae positive	3
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O5	1
Meat from bovine animals - Cutting plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	292	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Meat from bovine animals - minced meat - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	318	2	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC NT (Not Typeable)	1
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O26 - eae positive	1
Meat from bovine animals - minced meat - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	146	2	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC NT (Not Typeable)	1
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O103 - eae positive	1
Meat from bovine animals - minced meat - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	147	1	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC NT (Not Typeable)	1
Meat, red meat (meat from bovines, pigs, goats, sheep, horses, donkeys, bison and water buffalos) - Border inspection activities - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	230	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Milk, cows' - raw milk - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Millilitre	339	7	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC NT (Not Typeable)	3
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O111 - eae positive	1
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O26 - eae positive	1
						Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC, unspecified	2
Seeds, sprouted - Border inspection activities - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O104:H4 - EAaggEC positive vtx2 positive	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Seeds, sprouted - Border inspection activities - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Seeds, sprouted - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	15	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O104:H4 - EAggEC positive vtx2 positive	0
Seeds, sprouted - Farm (not specified) - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	15	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Seeds, sprouted - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	68	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O104:H4 - EAggEC positive vtx2 positive	0
Seeds, sprouted - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	68	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Seeds, sprouted - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	75	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O104:H4 - EAggEC positive vtx2 positive	0
Seeds, sprouted - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	75	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Seeds, sprouted - Wholesale - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	67	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC) - VTEC O104:H4 - EAggEC positive vtx2 positive	0
Seeds, sprouted - Wholesale - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	67	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Spices and herbs - fresh - Processing plant - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	71	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Spices and herbs - fresh - Wholesale - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	77	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Vegetables - Retail - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	227	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0
Vegetables - Wholesale - Belgium - food sample (not specified) - Surveillance - Official sampling - Objective sampling	single	25	Gram	229	0	Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	0

Table HISTAMINE in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Fish - Fishery products which have undergone enzyme maturation treatment in brine - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	batch	1	Gram	14	1	>200 to <= 400	Histamine	84	6

Table LISTERIA in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Bakery products - desserts - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	49	0	<100	Listeria - L. monocytogenes	49	0
Bakery products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	76	0	<100	Listeria - L. monocytogenes	61	0
Bakery products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	76	0	detection	Listeria - L. monocytogenes	15	0
Bakery products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	151	0	<100	Listeria - L. monocytogenes	151	0
Cheeses made from cows' milk - fresh - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	27	0	<100	Listeria - L. monocytogenes	5	0
Cheeses made from cows' milk - fresh - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	27	0	detection	Listeria - L. monocytogenes	22	0
Cheeses made from cows' milk - fresh - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	20	0	<100	Listeria - L. monocytogenes	11	0
Cheeses made from cows' milk - fresh - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	20	0	detection	Listeria - L. monocytogenes	9	0
Cheeses made from cows' milk - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	12	0	<100	Listeria - L. monocytogenes	12	0
Cheeses made from cows' milk - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	35	0	<100	Listeria - L. monocytogenes	35	0
Cheeses made from cows' milk - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	57	0	<100	Listeria - L. monocytogenes	57	0
Cheeses made from cows' milk - soft and semi-soft - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	94	3	<100	Listeria - L. monocytogenes	23	0
Cheeses made from cows' milk - soft and semi-soft - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	94	3	detection	Listeria - L. monocytogenes	71	3
Cheeses made from cows' milk - soft and semi-soft - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	8	0	<100	Listeria - L. monocytogenes	1	0
Cheeses made from cows' milk - soft and semi-soft - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	8	0	detection	Listeria - L. monocytogenes	7	0
Cheeses made from cows' milk - soft and semi-soft - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	27	0	detection	Listeria - L. monocytogenes	27	0
Cheeses made from cows' milk - soft and semi-soft - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	96	3	>100	Listeria - L. monocytogenes	192	3
Cheeses made from cows' milk - soft and semi-soft - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	92	0	<100	Listeria - L. monocytogenes	92	0
Cheeses made from goats' milk - unspecified - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	40	0	<100	Listeria - L. monocytogenes	3	0
Cheeses made from goats' milk - unspecified - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	40	0	detection	Listeria - L. monocytogenes	37	0
Cheeses made from goats' milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	10	0	<100	Listeria - L. monocytogenes	9	0
Cheeses made from goats' milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	0	detection	Listeria - L. monocytogenes	1	0
Cheeses made from goats' milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	48	5	<100	Listeria - L. monocytogenes	30	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Cheeses made from goats' milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	48	5	detection	Listeria - L. monocytogenes	18	5
Cheeses made from goats' milk - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	53	0	<100	Listeria - L. monocytogenes	53	0
Cheeses made from sheep's milk - unspecified - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	52	0	<100	Listeria - L. monocytogenes	8	0
Cheeses made from sheep's milk - unspecified - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	52	0	detection	Listeria - L. monocytogenes	44	0
Cheeses made from sheep's milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	11	0	<100	Listeria - L. monocytogenes	10	0
Cheeses made from sheep's milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	11	0	detection	Listeria - L. monocytogenes	1	0
Cheeses made from sheep's milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	60	2	<100	Listeria - L. monocytogenes	31	0
Cheeses made from sheep's milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	60	2	detection	Listeria - L. monocytogenes	29	2
Cheeses made from sheep's milk - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	98	0	<100	Listeria - L. monocytogenes	98	0
Cheeses made from sheep's milk - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	114	0	<100	Listeria - L. monocytogenes	114	0
Dairy products (excluding cheeses) - butter - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	127	33	detection	Listeria - L. monocytogenes	127	33
Dairy products (excluding cheeses) - cream - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	31	0	<100	Listeria - L. monocytogenes	6	0
Dairy products (excluding cheeses) - cream - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	31	0	detection	Listeria - L. monocytogenes	25	0
Dairy products (excluding cheeses) - cream - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	18	0	<100	Listeria - L. monocytogenes	9	0
Dairy products (excluding cheeses) - cream - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	18	1	detection	Listeria - L. monocytogenes	9	1
Dairy products (excluding cheeses) - dairy desserts - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	110	0	<100	Listeria - L. monocytogenes	110	0
Dairy products (excluding cheeses) - fermented dairy products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	15	0	<100	Listeria - L. monocytogenes	30	0
Dairy products (excluding cheeses) - ice-cream - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	117	0	<100	Listeria - L. monocytogenes	117	0
Dairy products (excluding cheeses) - ice-cream - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	10	0	<100	Listeria - L. monocytogenes	10	0
Dairy products (excluding cheeses) - ice-cream - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	117	0	<100	Listeria - L. monocytogenes	117	0
Dairy products (excluding cheeses) - milk powder and whey powder - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	1	0	<100	Listeria - L. monocytogenes	1	0
Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	45	0	<100	Listeria - L. monocytogenes	45	0
Dairy products (excluding cheeses) - yoghurt - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	67	0	<100	Listeria - L. monocytogenes	67	0
Dairy products (excluding cheeses) - yoghurt - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	25	0	<100	Listeria - L. monocytogenes	25	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Dairy products (excluding cheeses) - yoghurt - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	42	0	<100	Listeria - L. monocytogenes	42	0
Fishery products, unspecified - ready-to-eat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	192	0	<100	Listeria - L. monocytogenes	119	0
Fishery products, unspecified - ready-to-eat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	192	0	detection	Listeria - L. monocytogenes	73	0
Fishery products, unspecified - ready-to-eat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	405	0	<100	Listeria - L. monocytogenes	405	0
Fishery products, unspecified - ready-to-eat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	275	0	<100	Listeria - L. monocytogenes	275	0
Fishery products, unspecified - smoked - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	79	6	<100	Listeria - L. monocytogenes	30	0
Fishery products, unspecified - smoked - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	79	6	detection	Listeria - L. monocytogenes	49	0
Fishery products, unspecified - smoked - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	140	6	<100	Listeria - L. monocytogenes	140	0
Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	305	1	detection	Listeria - L. monocytogenes	305	1
Fruits and vegetables - pre-cut - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	113	0	<100	Listeria - L. monocytogenes	39	0
Fruits and vegetables - pre-cut - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	113	0	detection	Listeria - L. monocytogenes	74	0
Fruits and vegetables - pre-cut - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	112	0	<100	Listeria - L. monocytogenes	112	0
Infant formula - dried - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	30	0	detection	Listeria - L. monocytogenes	30	0
Infant formula - dried - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	514	0	detection	Listeria - L. monocytogenes	514	0
Infant formula - ready-to-eat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	148	0	detection	Listeria - L. monocytogenes	148	0
Meat from bovine animals - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	14	3	<100	Listeria - L. monocytogenes	3	0
				351	25	<100	Listeria - L. monocytogenes	218	0
				14	0	detection	Listeria - L. monocytogenes	4	0
				10	0	detection	Listeria - L. monocytogenes	1	0
				25	3	detection	Listeria - L. monocytogenes	6	3
Meat from bovine animals - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	14	0	detection	Listeria - L. monocytogenes	4	0
				10	0	detection	Listeria - L. monocytogenes	1	0
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	14	3	detection	Listeria - L. monocytogenes	6	3
				351	25	detection	Listeria - L. monocytogenes	133	25
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	149	0	<100	Listeria - L. monocytogenes	149	0
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	55	0	<100	Listeria - L. monocytogenes	55	0
				146	0	<100	Listeria - L. monocytogenes	146	0
Meat from bovine animals and pig - meat preparation - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	5	0	<100	Listeria - L. monocytogenes	5	0
Meat from bovine animals and pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	113	9	<100	Listeria - L. monocytogenes	18	0
Meat from bovine animals and pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	113	9	detection	Listeria - L. monocytogenes	95	9

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Meat from bovine animals and pig - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	84	0	<100	Listeria - L. monocytogenes	84	0
Meat from bovine animals and pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	32	7	<100	Listeria - L. monocytogenes	12	0
Meat from bovine animals and pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	32	7	detection	Listeria - L. monocytogenes	20	7
Meat from bovine animals and pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	5	0	<100	Listeria - L. monocytogenes	5	0
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	120	7	<100	Listeria - L. monocytogenes	39	0
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	120	7	detection	Listeria - L. monocytogenes	81	7
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	176	0	<100	Listeria - L. monocytogenes	91	0
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	176	0	detection	Listeria - L. monocytogenes	85	0
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	127	4	<100	Listeria - L. monocytogenes	93	0
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	127	4	detection	Listeria - L. monocytogenes	34	4
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	4	0	<100	Listeria - L. monocytogenes	1	0
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	4	0	detection	Listeria - L. monocytogenes	3	0
Meat from pig - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	113	0	<100	Listeria - L. monocytogenes	113	0
Meat from pig - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	170	1	>100	Listeria - L. monocytogenes	170	1
Meat from pig - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	117	1	>100	Listeria - L. monocytogenes	117	1
Meat from pig - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	11	0	<100	Listeria - L. monocytogenes	11	0
Meat from pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	74	15	<100	Listeria - L. monocytogenes	26	1
Meat from pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	74	15	detection	Listeria - L. monocytogenes	18	1
Meat from pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	74	15	detection	Listeria - L. monocytogenes	30	13
Meat from pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	13	0	<100	Listeria - L. monocytogenes	13	0
Meat from poultry, unspecified - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	146	1	<100	Listeria - L. monocytogenes	90	0
Meat from poultry, unspecified - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	146	1	detection	Listeria - L. monocytogenes	56	1
Meat from poultry, unspecified - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	150	0	<100	Listeria - L. monocytogenes	150	0
Meat, mixed meat - meat preparation - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	0	detection	Listeria - L. monocytogenes	5	0
Meat, mixed meat - meat preparation - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	8	5	detection	Listeria - L. monocytogenes	8	5

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Method	Zoonoses	N of units tested	N of units positive
Milk, cows' - raw milk - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	45	0	<100	Listeria - L. monocytogenes	45	0
Other processed food products and prepared dishes - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	152	1	<100	Listeria - L. monocytogenes	21	0
Other processed food products and prepared dishes - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	152	1	detection	Listeria - L. monocytogenes	131	1
Other processed food products and prepared dishes - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	25	0	<100	Listeria - L. monocytogenes	25	0
Other processed food products and prepared dishes - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	1063	1	>100	Listeria - L. monocytogenes	1,062	1
Seeds, sprouted - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	1	0	<100	Listeria - L. monocytogenes	1	0
Seeds, sprouted - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	14	0	<100	Listeria - L. monocytogenes	14	0
Seeds, sprouted - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	68	0	detection	Listeria - L. monocytogenes	68	0
Seeds, sprouted - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	74	0	<100	Listeria - L. monocytogenes	74	0
Seeds, sprouted - Wholesale - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	65	0	<100	Listeria - L. monocytogenes	65	0
Surimi - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	11	0	<100	Listeria - L. monocytogenes	11	0

Table LYSSAVIRUS (RABIES) in animal

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Bats - wild - Natural habitat - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	16	0	Lyssavirus (rabies)	0
Cats - Veterinary clinics - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	9	0	Lyssavirus (rabies)	0
Cattle (bovine animals) - Farm (not specified) - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	128	0	Lyssavirus (rabies)	0
Dogs - Veterinary clinics - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	21	0	Lyssavirus (rabies)	0
Foxes - wild - Natural habitat - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	4	0	Lyssavirus (rabies)	0
Goats - Farm (not specified) - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	56	0	Lyssavirus (rabies)	0
Raccoons - wild - Natural habitat - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	1	0	Lyssavirus (rabies)	0
Sheep - Farm (not specified) - Belgium - - Surveillance - Official sampling - Suspect sampling	animal	107	0	Lyssavirus (rabies)	0

Table SALMONELLA in animal

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	Target verification	Total units tested	Total units positive	Zoonoses	N of units positive
Gallus gallus (fowl) - broilers - Farm (not specified) - Belgium - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/flock		Y	8946	177	Salmonella - Not typeable	2
						Salmonella - Other serovars	7
						Salmonella - S. 1,4,[5],12:-:-	4
						Salmonella - S. 1,4,[5],12:i:-	2
						Salmonella - S. 21:-:-	1
						Salmonella - S. 3,10:-:-	1
						Salmonella - S. 3,19:-:-	1
						Salmonella - S. 4,12:i:-	5
						Salmonella - S. 6,7:-:-	1
						Salmonella - S. 9,46:-:-	1
						Salmonella - S. Agona	13
						Salmonella - S. Cerro	4
						Salmonella - S. Chester	3
						Salmonella - S. Derby	6
						Salmonella - S. Dublin	1
						Salmonella - S. Enteritidis	7
						Salmonella - S. Havana	2
						Salmonella - S. Idikan	2
						Salmonella - S. Infantis	21
						Salmonella - S. Java	26
						Salmonella - S. Kentucky	1
						Salmonella - S. Kottbus	3
						Salmonella - S. Lexington	1
						Salmonella - S. Livingstone	22
						Salmonella - S. Mbandaka	8
						Salmonella - S. Minnesota	11
						Salmonella - S. Montevideo	1
Salmonella - S. Ohio	1						
Salmonella - S. Rissen	1						
Salmonella - S. Senftenberg	4						
Salmonella - S. Typhimurium	12						
Salmonella - S. Virchow	1						
Salmonella - S. Wien	1						
Gallus gallus (fowl) - broilers - Farm (not specified) - Belgium - environmental sample - delivery box liner - Control and eradication programmes - Official and industry sampling - Census	herd/flock		N	5473	10	Salmonella - S. Enteritidis	5
						Salmonella - S. Idikan	1
						Salmonella - S. Mbandaka	2
						Salmonella - S. Minnesota	1
						Salmonella - S. Typhimurium	1

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	N of flocks under control programme	Target verification	Total units tested	Total units positive	Zoonoses	N of units positive
Gallus gallus (fowl) - laying hens - Farm (not specified) - Belgium - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/flock		N	296	4	Salmonella - S. Senftenberg	3
						Salmonella - S. Tennessee	1
Gallus gallus (fowl) - laying hens - Farm (not specified) - Belgium - environmental sample - boot swabs and dust - Control and eradication programmes - Official and industry sampling - Census	herd/flock		Y	644	28	Salmonella - S. Agona	2
						Salmonella - S. Bareilly	1
						Salmonella - S. Enteritidis	12
						Salmonella - S. Infantis	6
						Salmonella - S. Lexington	2
						Salmonella - S. Livingstone	1
						Salmonella - S. Senftenberg	2
						Salmonella - S. Tennessee	1
Salmonella - S. Typhimurium	1						
Gallus gallus (fowl) - laying hens - Farm (not specified) - Belgium - environmental sample - delivery box liner - Control and eradication programmes - Official and industry sampling - Census	herd/flock		N	172	0	Salmonella	0
Gallus gallus (fowl) - parent breeding flocks, unspecified - Farm (not specified) - Belgium - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/flock		N	341	2	Salmonella - S. Mbandaka	1
						Salmonella - S. Typhimurium	1
Gallus gallus (fowl) - parent breeding flocks, unspecified - Farm (not specified) - Belgium - environmental sample - boot swabs and dust - Control and eradication programmes - Official and industry sampling - Census	herd/flock		Y	503	21	Salmonella - S. 4,12:i:-	1
						Salmonella - S. 6,7:-:-	1
						Salmonella - S. Anatum	1
						Salmonella - S. Djugu	1
						Salmonella - S. Enteritidis	2
						Salmonella - S. Jerusalem	2
						Salmonella - S. Lexington	1
						Salmonella - S. Mbandaka	9
Salmonella - S. Typhimurium	3						
Gallus gallus (fowl) - parent breeding flocks, unspecified - Farm (not specified) - Belgium - environmental sample - delivery box liner - Control and eradication programmes - Official and industry sampling - Census	herd/flock		N	210	0	Salmonella	0
Pigs - fattening pigs - Farm (not specified) - Belgium - animal sample - faeces - Control and eradication programmes - Industry sampling - Selective sampling	holding		N	22	18	Salmonella - S. 4,12:i:-	3
						Salmonella - S. 4,5,12:i:-	1
						Salmonella - S. Brandenburg	1
						Salmonella - S. Derby	3
						Salmonella - S. Goldcoast	1
						Salmonella - S. Livingstone	2
						Salmonella - S. London	2
						Salmonella - S. Typhimurium	10
Turkeys - fattening flocks - Farm (not specified) - Belgium - environmental sample - boot swabs - Control and eradication programmes - Official and industry sampling - Census	herd/flock		Y	82	6	Salmonella - S. 4,[5],12:i:-	1
						Salmonella - S. Chester	4
						Salmonella - S. Typhimurium	1

Table SALMONELLA in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Bakery products - desserts - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	49	0	Salmonella	0
Bakery products - pastry - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	32	0	Salmonella	0
Bakery products - pastry - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	60	0	Salmonella	0
Cheeses made from cows' milk - fresh - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	22	0	Salmonella	0
Cheeses made from cows' milk - fresh - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	20	0	Salmonella	0
Cheeses made from cows' milk - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	12	0	Salmonella	0
Cheeses made from cows' milk - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	17	0	Salmonella	0
Cheeses made from cows' milk - soft and semi-soft - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	70	0	Salmonella	0
Cheeses made from cows' milk - soft and semi-soft - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	66	0	Salmonella	0
Cheeses made from cows' milk - soft and semi-soft - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	60	0	Salmonella	0
Cheeses made from cows' milk - soft and semi-soft - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	74	0	Salmonella	0
Cheeses made from cows' milk - soft and semi-soft - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	66	0	Salmonella	0
Cheeses made from goats' milk - unspecified - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	28	0	Salmonella	0
Cheeses made from goats' milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	16	0	Salmonella	0
Cheeses made from goats' milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	65	0	Salmonella	0
Cheeses made from goats' milk - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	42	0	Salmonella	0
Cheeses made from goats' milk - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	59	0	Salmonella	0
Cheeses made from sheep's milk - unspecified - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	0	Salmonella	0
Cheeses made from sheep's milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Salmonella	0
Cheeses made from sheep's milk - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	38	0	Salmonella	0
Cheeses made from sheep's milk - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Cheeses made from sheep's milk - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Crustaceans - unspecified - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	12	0	Salmonella	0
Crustaceans - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Crustaceans - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	30	0	Salmonella	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Crustaceans - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Crustaceans - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	30	0	Salmonella	0
Dairy products (excluding cheeses) - butter - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	92	0	Salmonella	0
Dairy products (excluding cheeses) - cream - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	30	0	Salmonella	0
Dairy products (excluding cheeses) - cream - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	18	0	Salmonella	0
Dairy products (excluding cheeses) - dairy desserts - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	44	0	Salmonella	0
Dairy products (excluding cheeses) - dairy desserts - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Dairy products (excluding cheeses) - ice-cream - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	42	0	Salmonella	0
Dairy products (excluding cheeses) - ice-cream - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	5	0	Salmonella	0
Dairy products (excluding cheeses) - ice-cream - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Dairy products (excluding cheeses) - milk powder and whey powder - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Salmonella	0
Dairy products (excluding cheeses) - milk powder and whey powder - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Egg products - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Salmonella	0
Egg products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Egg products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	20	2	Salmonella - S. Typhimurium, monophasic	2
Eggs - table eggs - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	118	0	Salmonella	0
Fishery products, unspecified - ready-to-eat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	66	0	Salmonella	0
Fishery products, unspecified - ready-to-eat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	201	0	Salmonella	0
Fishery products, unspecified - ready-to-eat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	116	0	Salmonella	0
Foodstuffs intended for special nutritional uses - dried dietary foods for special medical purposes intended for infants below 6 months - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	60	0	Salmonella	0
Frogs leg - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	4	0	Salmonella	0
Frogs leg - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	25	15	Salmonella - Not typeable	1
						Salmonella - Other serovars	2
						Salmonella - S. Bardo	1
						Salmonella - S. Hvitvingfoss	2
						Salmonella - S. Javiana	1
						Salmonella - S. Matopeni	1
						Salmonella - S. Newport	1
						Salmonella - S. Paratyphi B	1
						Salmonella - S. Sandiego	1
						Salmonella - S. Stanley	1

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Frogs leg - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	25	15	Salmonella - S. Toronto	1
						Salmonella - S. Wandsworth	1
						Salmonella - S. Weltevreden	1
Fruits - non-pre-cut - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Fruits - non-pre-cut - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Fruits - non-pre-cut - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Fruits - non-pre-cut - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Fruits and vegetables - pre-cut - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	31	0	Salmonella	0
Fruits and vegetables - pre-cut - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	60	0	Salmonella	0
Infant formula - dried - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	30	0	Salmonella	0
Infant formula - dried - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	180	0	Salmonella	0
Infant formula - ready-to-eat - Hospital or medical care facility - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	118	0	Salmonella	0
Juice - fruit juice - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	0	Salmonella	0
Juice - fruit juice - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	56	0	Salmonella	0
Live bivalve molluscs - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	81	0	Salmonella	0
Meat from bovine animals - carcass - Slaughterhouse - Belgium - food sample - carcass swabs - Surveillance - Official sampling - Objective sampling	single	1600	Square centimetre	250	0	Salmonella	0
Meat from bovine animals - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	35	0	Salmonella	0
Meat from bovine animals - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	50	0	Salmonella	0
Meat from bovine animals - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	121	1	Salmonella - S. Typhimurium	1
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	27	0	Salmonella	0
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	44	0	Salmonella	0
				51	6	Salmonella - S. Enteritidis	6
Meat from bovine animals and pig - meat preparation - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	62	2	Salmonella - S. Saintpaul	1
						Salmonella - S. Typhimurium, monophasic	1
Meat from bovine animals and pig - meat preparation - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	65	6	Salmonella - S. Typhimurium	1
						Salmonella - S. Typhimurium, monophasic	5
Meat from bovine animals and pig - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	11	1	Salmonella - S. Saintpaul	1
Meat from bovine animals and pig - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Salmonella	0
Meat from bovine animals and pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	3	Salmonella - S. 4,12:i:-	1
						Salmonella - S. Brandenburg	1
						Salmonella - S. Typhimurium	1
Meat from bovine animals and pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	11	0	Salmonella	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from broilers (Gallus gallus) - carcase - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	91	1	Salmonella - S. Paratyphi B	1
Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	293	1	Salmonella - S. Infantis	1
						Salmonella - Salmonella spp., unspecified	1
Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Belgium - animal sample - caecum - Surveillance - Official sampling - Objective sampling	single	25	Gram	175	4	Salmonella - S. Agona	1
						Salmonella - S. Chester	1
						Salmonella - S. Infantis	1
						Salmonella - S. Typhimurium	1
Meat from broilers (Gallus gallus) - carcase - Slaughterhouse - Belgium - food sample - neck skin - Surveillance - Official sampling - Objective sampling	single	25	Gram	904	50	Salmonella - S. Agona	3
						Salmonella - S. enterica subsp. enterica	11
						Salmonella - S. Enteritidis	3
						Salmonella - S. Infantis	2
						Salmonella - S. Livingstone	1
						Salmonella - S. Paratyphi B	27
						Salmonella - Salmonella spp., unspecified	3
Meat from broilers (Gallus gallus) - mechanically separated meat (MSM) - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	12	0	Salmonella	0
Meat from other animal species or not specified - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Meat from other animal species or not specified - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Meat from other poultry species - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Meat from other poultry species - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Meat from pig - carcase - Slaughterhouse - Belgium - food sample - carcase swabs - Surveillance - Official sampling - Objective sampling	single	600	Square centimetre	447	57	Salmonella - S. Brandenburg	17
						Salmonella - S. Derby	3
						Salmonella - S. enterica subsp. enterica	1
						Salmonella - S. Infantis	1
						Salmonella - S. Livingstone	2
						Salmonella - S. Rissen	1
						Salmonella - S. Typhimurium	14
						Salmonella - S. Typhimurium, monophasic	15
						Salmonella - Salmonella spp., unspecified	3
Meat from pig - fresh - Cutting plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	285	8	Salmonella - S. Brandenburg	1
						Salmonella - S. Derby	4
						Salmonella - S. Typhimurium	3
Meat from pig - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	51	0	Salmonella	0
Meat from pig - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	16	0	Salmonella	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	50	0	Salmonella	0
Meat from pig - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Meat from pig - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Meat from pig - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Meat from pig - mechanically separated meat (MSM) - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Salmonella	0
Meat from pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	10	0	Salmonella	0
Meat from pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	21	0	Salmonella	0
Meat from poultry, unspecified - fresh - Cutting plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	247	6	Salmonella - S. enterica subsp. enterica	2
						Salmonella - S. Infantis	1
						Salmonella - S. Typhimurium	2
						Salmonella - S. Typhimurium, monophasic	1
Meat from poultry, unspecified - fresh - Cutting plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	326	15	Salmonella - S. enterica subsp. enterica	1
						Salmonella - S. Indiana	1
						Salmonella - S. Infantis	2
						Salmonella - S. Livingstone	2
						Salmonella - S. Paratyphi B	4
						Salmonella - S. Typhimurium, monophasic	4
						Salmonella - Salmonella spp., unspecified	1
Meat from poultry, unspecified - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	64	4	Salmonella - S. 4,[5],12:i:-	1
						Salmonella - S. Livingstone	1
						Salmonella - S. Paratyphi B	1
						Salmonella - S. Typhimurium var. Copenhagen	1
Meat from poultry, unspecified - fresh - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	61	4	Salmonella - Not typeable	1
						Salmonella - S. Infantis	1
						Salmonella - S. Livingstone	1
						Salmonella - S. Paratyphi B	1
Meat from poultry, unspecified - meat products - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	3	0	Salmonella	0
Meat from poultry, unspecified - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Meat from poultry, unspecified - meat products - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	64	2	Salmonella - S. Paratyphi B	2
Meat from poultry, unspecified - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	46	0	Salmonella	0
Meat from poultry, unspecified - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	59	0	Salmonella	0
Meat from poultry, unspecified - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	18	1	Salmonella	1

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from rabbit - meat preparation - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	8	0	Salmonella	0
Meat from rabbit - meat preparation - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	10	Gram	65	6	Salmonella - Not typeable	2
						Salmonella - S. Infantis	1
						Salmonella - S. Paratyphi B	2
						Salmonella - S. Rissen	1
Meat from rabbit - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	115	3	Salmonella - S. Infantis	1
						Salmonella - S. Livingstone	1
						Salmonella - S. Paratyphi B	1
Meat from spent hens (Gallus gallus) - fresh - Slaughterhouse - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	585	47	Salmonella - S. Braenderup	1
						Salmonella - S. Brandenburg	1
						Salmonella - S. enterica subsp. enterica	1
						Salmonella - S. Enteritidis	36
						Salmonella - S. Putten	1
						Salmonella - S. Rissen	1
						Salmonella - S. Typhimurium	3
						Salmonella - Salmonella spp., unspecified	3
Meat from spent hens (Gallus gallus) - fresh - Slaughterhouse - Belgium - animal sample - caecum - Surveillance - Official sampling - Objective sampling	single	25	Gram	43	9	Salmonella - S. Enteritidis	4
						Salmonella - S. Gloucester	1
						Salmonella - S. Tennessee	1
						Salmonella - S. Typhimurium	3
Milk, cows' - raw milk - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Molluscan shellfish - cooked - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Molluscan shellfish - cooked - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	41	0	Salmonella	0
Other processed food products and prepared dishes - unspecified - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	45	0	Salmonella	0
Other processed food products and prepared dishes - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	25	0	Salmonella	0
Other processed food products and prepared dishes - unspecified - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	489	2	Salmonella - S. Rissen	2
Seeds, sprouted - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Salmonella	0
Seeds, sprouted - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	36	0	Salmonella	0
Seeds, sprouted - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	26	0	Salmonella	0
Seeds, sprouted - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	26	1	Salmonella - S. Toronto	1
Spices and herbs - dried - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	71	0	Salmonella	0
Spices and herbs - dried - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	59	0	Salmonella	0
Spices and herbs - fresh - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	17	0	Salmonella	0
Spices and herbs - fresh - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	50	0	Salmonella	0
Spices and herbs - fresh - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	44	0	Salmonella	0
Surimi - Border inspection activities - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	14	0	Salmonella	0
Vegetables - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	134	0	Salmonella	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Vegetables - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	138	0	Salmonella	0

Table SALMONELLA in feed

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
All feedingstuffs - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	95	4	Salmonella - S. Derby	1
						Salmonella - S. Gloucester	1
						Salmonella - S. Infantis	1
						Salmonella - S. Java	1
						Salmonella - S. Livingstone	1
						Salmonella - S. Montevideo	1
						Salmonella - S. Worthington	1
Compound feedingstuffs for cattle - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	90	0	Salmonella	0
Compound feedingstuffs for fish - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	17	0	Salmonella	0
Compound feedingstuffs for horses - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	25	0	Salmonella	0
Compound feedingstuffs for pigs - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	118	2	Salmonella - S. Agona	1
						Salmonella - S. Anatum	1
						Salmonella - S. Cerro	1
Compound feedingstuffs for poultry (non specified) - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	15	0	Salmonella	0
Compound feedingstuffs for poultry, breeders - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	86	2	Salmonella - S. Livingstone	1
						Salmonella - S. Typhimurium	1
Compound feedingstuffs for poultry, broilers - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	53	1	Salmonella - S. Yoruba	1
Compound feedingstuffs for poultry, laying hens - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	42	2	Salmonella - S. 1,3,19:-:-	1
						Salmonella - S. Livingstone	2
Compound feedingstuffs for poultry, pigeons - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	5	0	Salmonella	0
Compound feedingstuffs for rabbits - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	13	0	Salmonella	0
Compound feedingstuffs for sheep - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	15	0	Salmonella	0
Compound feedingstuffs for turkeys - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	4	0	Salmonella	0
Compound feedingstuffs, not specified - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	5	0	Salmonella	0
Feed material of cereal grain origin - barley derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	3	0	Salmonella	0
						Salmonella	0
Feed material of cereal grain origin - maize derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	1	0	Salmonella	0
				2	0	Salmonella	0
Feed material of cereal grain origin - oat derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	1	0	Salmonella	0
Feed material of cereal grain origin - other cereal grain derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	17	0	Salmonella	0

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Feed material of cereal grain origin - wheat derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	1	0	Salmonella	0
				10	0	Salmonella	0
Feed material of land animal origin - animal fat - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	2	0	Salmonella	0
Feed material of land animal origin - blood meal - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	2	0	Salmonella	0
Feed material of land animal origin - blood products - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	3	0	Salmonella	0
Feed material of land animal origin - egg powder - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	5	0	Salmonella	0
				29	1	Salmonella - S. Enteritidis	1
Feed material of land animal origin - meat and bone meal - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	7	0	Salmonella	0
				11	1	Salmonella - Salmonella spp., unspecified	1
				118	13	Salmonella - S. Agona	1
						Salmonella - S. Cerro	1
						Salmonella - S. enterica subsp. enterica	1
						Salmonella - S. Grumpensis	1
						Salmonella - S. Il 6,7:m,t-	2
						Salmonella - S. Infantis	1
						Salmonella - S. Livingstone	2
						Salmonella - S. Mbandaka	1
						Salmonella - S. Montevideo	1
						Salmonella - S. Oran	1
						Salmonella - S. Rissen	3
		Salmonella - S. Senftenberg	3				
		Salmonella - S. Sinstorf	1				
Feed material of land animal origin - poultry offal meal - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	2	0	Salmonella	0
Feed material of marine animal origin - fish meal - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	8	0	Salmonella	0
Feed material of oil seed or fruit origin - groundnut derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	1	0	Salmonella	0
Feed material of oil seed or fruit origin - linseed derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	2	0	Salmonella	0
Feed material of oil seed or fruit origin - other oil seeds derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	2	0	Salmonella	0
Feed material of oil seed or fruit origin - palm kernel derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	1	0	Salmonella	0
Feed material of oil seed or fruit origin - rape seed derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	6	0	Salmonella	0
Feed material of oil seed or fruit origin - soya (bean) derived - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	4	0	Salmonella	0
				14	0	Salmonella	0
Pet food - dog snacks (pig ears, chewing bones) - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	28	2	Salmonella - S. Agona	1
						Salmonella - S. Anatum	1

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Pet food - dog snacks (pig ears, chewing bones) - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	28	2	Salmonella - S. Livingstone	1
						Salmonella - S. Montevideo	1
						Salmonella - S. Panama	1
Pet food - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	108	2	Salmonella - S. 6,7:z29	1
						Salmonella - S. Enteritidis	1
						Salmonella - S. Havana	1
						Salmonella - S. Saintpaul	1
Premixtures - Unspecified - Belgium - feed sample - Surveillance - Official sampling - Objective sampling	batch	25	Gram	1	0	Salmonella	0

Table SARCOCYSTIS in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from bovine animals - carcase - Slaughterhouse - Belgium - - Surveillance - Official sampling - Suspect sampling	animal		NOT AVAILABLE	83747 0	94	Sarcocystis	0

Table STAPHYLOCOCCAL ENTEROTOXINS in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Bakery products - pastry - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Staphylococcal enterotoxins	0
Bakery products - pastry - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Staphylococcal enterotoxins	0
Cheeses, made from unspecified milk or other animal milk - unspecified - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	9	0	Staphylococcal enterotoxins	0
Crustaceans - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Staphylococcal enterotoxins	0
Dairy products (excluding cheeses) - butter - Farm (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	8	0	Staphylococcal enterotoxins	0
Fish (food) - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Staphylococcal enterotoxins	0
Meat from other animal species or not specified - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	4	1	Staphylococcal enterotoxins - Enterotoxin C	0
Meat from other animal species or not specified - meat preparation - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	8	1	Staphylococcal enterotoxins - Enterotoxin A	0
						Staphylococcal enterotoxins - Enterotoxin C	0
Meat from other animal species or not specified - meat products - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Staphylococcal enterotoxins	0
Other processed food products and prepared dishes - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	4	0	Staphylococcal enterotoxins - Enterotoxin C	0
				31	1	Staphylococcal enterotoxins - Enterotoxin B	0
Other processed food products and prepared dishes - pasta - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Staphylococcal enterotoxins	0
Other processed food products and prepared dishes - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	3	0	Staphylococcal enterotoxins - Enterotoxin C	0
Sauce and dressings - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Staphylococcal enterotoxins	0
Sauce and dressings - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Staphylococcal enterotoxins	0
Vegetables - Catering (not specified) - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	2	0	Staphylococcal enterotoxins	0
Vegetables - pre-cut - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	1	Staphylococcal enterotoxins - Enterotoxin C	0
Vegetables - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	25	Gram	1	0	Staphylococcal enterotoxins	0

Table STAPHYLOCOCCUS AUREUS METICILLIN RESISTANT (MRSA) in animal

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Gallus gallus (fowl) - broilers - Farm (not specified) - Belgium - animal sample - nasal swab - Monitoring - active - Official sampling - Objective sampling	holding	159	1	Staphylococcus - S. aureus, meticillin resistant (MRSA) - spa-type t011 - CC398	0
				Staphylococcus - S. aureus, meticillin resistant (MRSA) - spa-type t1985 - CC398	0
Gallus gallus (fowl) - laying hens - Farm (not specified) - Belgium - animal sample - nasal swab - Monitoring - active - Official sampling - Objective sampling	holding	233	1	Staphylococcus - S. aureus, meticillin resistant (MRSA) - spa-type t011 - CC398	0
				5	Staphylococcus - S. aureus, meticillin resistant (MRSA)
Gallus gallus (fowl) - laying hens - Farm (not specified) - Belgium - animal sample - nasal swab - Monitoring - active - Official sampling - Objective sampling	holding	12	0	Staphylococcus - S. aureus, meticillin resistant (MRSA)	0

Table TRICHINELLA in animal

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Total units tested	Total units positive	Zoonoses	N of units positive
Pigs - Slaughterhouse - Belgium - - Surveillance - Official sampling - Census	animal	11888 367	0	Trichinella	0
Solipeds, domestic - Slaughterhouse - Belgium - - Surveillance - Official sampling - Census	animal	8337	0	Trichinella	0
Wild boars - wild - Game handling establishment - Belgium - - Surveillance - Official sampling - Census	animal	11264	0	Trichinella	0

Table WEST NILE VIRUS in animal

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Vaccination status	Total units tested	Total units positive	Zoonoses	N of units positive
Birds - wild - Natural habitat - Belgium - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	No	634	0	West Nile virus	0
Birds - wild - Natural habitat - Belgium - animal sample - nasal swab - Monitoring - active - Official sampling - Selective sampling	animal	No	221	0	West Nile virus	0
Birds - wild - Natural habitat - Belgium - animal sample - organ/tissue - Monitoring - passive - Official sampling - Suspect sampling	animal	No	254	0	West Nile virus	0
Gallus gallus (fowl) - laying hens - Farm (not specified) - Belgium - animal sample - blood - Monitoring - active - Official sampling - Selective sampling	animal	No	1680	0	West Nile virus	0

Table YERSINIA in food

Matrix - Sampling stage - Sampling origin - Sample type - Sampling context - Sampler - Sampling strategy	Sampling unit	Sample weight	Sample weight unit	Total units tested	Total units positive	Zoonoses	N of units positive
Meat from bovine animals - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	16	5	Yersinia - Y. enterocolitica - biotype 1A	5
						Yersinia - Y. enterocolitica - Y. enterocolitica, unspecified	0
Meat from bovine animals - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	49	18	Yersinia - Y. enterocolitica - biotype 1A	18
						Yersinia - Y. enterocolitica - Y. enterocolitica, unspecified	0
Meat from bovine animals and pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	33	21	Yersinia - Y. enterocolitica - biotype 1A	21
						Yersinia - Y. enterocolitica - biotype 4/O:3	1
						Yersinia - Y. enterocolitica - Y. enterocolitica, unspecified	0
Meat from pig - carcase - Slaughterhouse - Belgium - - Surveillance - Official sampling - Objective sampling	single	600	Square centimetre	385	2	Yersinia - Y. enterocolitica - biotype 1A	0
						37	Yersinia - Y. enterocolitica - Y. enterocolitica, unspecified
Meat from pig - minced meat - Processing plant - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	69	20	Yersinia - Y. enterocolitica - biotype 1A	19
						Yersinia - Y. enterocolitica - biotype 4/O:3	1
						Yersinia - Y. enterocolitica - Y. enterocolitica, unspecified	0
Meat from pig - minced meat - Retail - Belgium - - Surveillance - Official sampling - Objective sampling	single	1	Gram	22	5	Yersinia - Y. enterocolitica - biotype 1A	5
						Yersinia - Y. enterocolitica - Y. enterocolitica, unspecified	0

FOODBORNE OUTBREAKS TABLES

Foodborne Outbreaks: summarized data

Causative agent	Food vehicle	Outbreak strenght							
		Strong				Weak			
		N outbreaks	N human cases	N hospitalized	N deaths	N outbreaks	N human cases	N hospitalized	N deaths
Bacillus - B. cereus	Buffet meals	1	13	0	0	2	11	0	0
	Mixed food	3	7	0	0	3	10	0	0
	Crustaceans, shellfish, molluscs and products thereof					1	2	0	0
	Unknown					1	3	0	0
Calicivirus - norovirus (Norwalk-like virus)	Mixed food	2	220	0	0	3	55	0	0
Campylobacter - Campylobacter spp., unspecified	Bovine meat and products thereof					1	2	0	0
Clostridium - C. perfringens	Mixed food	1	17	1	0				
Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	Bovine meat and products thereof					1	2	1	0
Histamine	Mixed food	1	2	0	0				
	Fish and fish products	1	2	2	0				
Listeria - L. monocytogenes - L. monocytogenes serovar 1/2a	Bovine meat and products thereof					1	2	1	0
Salmonella - S. Enteritidis	Eggs and egg products	3	68	4	0	1	2	1	0
Salmonella - S. Hadar	Turkey meat and products thereof					1	10	0	0
Staphylococcal enterotoxins - Enterotoxin A	Mixed food	1	19	11	0	1	3	0	0
Staphylococcal enterotoxins - Enterotoxin C	Mixed food	1	3	0	0				
Staphylococcal enterotoxins - Enterotoxin, unspecified	Mixed food					1	14	0	0
Trichinella	Pig meat and products thereof	1	16	14	0				
Unknown	Buffet meals					12	73	4	0
	Mixed food	1	20	5	0	180	759	15	0
	Other foods					1	2	1	0
	Bakery products					9	24	0	0
	Tap water, including well water					1	4	0	0
	Drinks, including bottled water					2	8	0	0
	Fruit, berries and juices and other products thereof					1	3	0	0
	Canned food products					2	4	1	0
	Vegetables and juices and other products thereof					5	15	0	0
	Crustaceans, shellfish, molluscs and products thereof					7	17	0	0
	Fish and fish products					13	40	1	0
	Broiler meat (Gallus gallus) and products thereof					12	51	0	0
	Other or mixed red meat and products thereof					12	34	0	0
	Pig meat and products thereof					10	29	0	0

Causative agent	Food vehicle	Outbreak strenght							
		Strong				Weak			
		N outbreaks	N human cases	N hospitalized	N deaths	N outbreaks	N human cases	N hospitalized	N deaths
Unknown	Bovine meat and products thereof					29	88	0	0
	Eggs and egg products					1	14	0	0
	Dairy products (other than cheeses)					3	8	0	0
	Unknown					37	113	2	0

Strong Foodborne Outbreaks: detailed data

Causative agent	FBO nat. code	Outbreak type	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Bacillus - B. cereus	313	Household / domestic kitchen	Mixed food	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	0	0
	343	Household / domestic kitchen	Mixed food	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Household	Take-away or fast-food outlet	Unknown	NOT AVAILABLE		1	3	0	0
	364	Household / domestic kitchen	Mixed food	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	0	0
	455	General	Buffet meals	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Household	Temporary mass catering (fairs or festivals)	Unknown	NOT AVAILABLE		1	13	0	0

Causative agent	FBO nat. code	Outbreak type	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Calicivirus - norovirus (Norwalk-like virus)	331	General	Mixed food	Descriptive epidemiological evidence	Temporary mass catering (fairs or festivals)	Temporary mass catering (fairs or festivals)	Unknown	NOT AVAILABLE		1	10	0	0
	426 447	General	Mixed food	Descriptive epidemiological evidence	Temporary mass catering (fairs or festivals)	Retail	Unknown	NOT AVAILABLE	International outbreak, descriptive epidemiological evidence and product-tracing investigation	1	210	0	0
Clostridium - C. perfringens	496	General	Mixed food	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	17	1	0
Histamine	460	Household / domestic kitchen	Mixed food	Descriptive epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	0	0
	469	General	Fish and fish products	Descriptive epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	2	0
Salmonella - S. Enteritidis	404	General	Eggs and egg products	Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans	Temporary mass catering (fairs or festivals)	Canteen or workplace catering	Unknown	NOT AVAILABLE		1	26	3	0
	405	Household / domestic kitchen	Eggs and egg products	Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	1	0

Causative agent	FBO nat. code	Outbreak type	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Salmonella - S. Enteritidis	417	General	Eggs and egg products	Detection of causative agent in food vehicle or its component - Detection of indistinguishable causative agent in humans	Temporary mass catering (fairs or festivals)	Temporary mass catering (fairs or festivals)	Unknown	NOT AVAILABLE		1	40	0	0
Staphylococcal enterotoxins - Enterotoxin A	427	General	Mixed food	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Temporary mass catering (fairs or festivals)	Temporary mass catering (fairs or festivals)	Unknown	NOT AVAILABLE		1	19	11	0
Staphylococcal enterotoxins - Enterotoxin C	310	Household / domestic kitchen	Mixed food	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Take-away or fast-food outlet	Take-away or fast-food outlet	Unknown	NOT AVAILABLE		1	3	0	0
Trichinella	603	General	Pig meat and products thereof	Descriptive epidemiological evidence	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE	Disseminated cases, descriptive epidemiological evidence and product-tracing investigation	1	16	14	0
Unknown	372	General	Mixed food	Detection of causative agent in food vehicle or its component - Symptoms and onset of illness pathognomonic to causative agent	Residential institution (nursing home or prison or boarding school) (not specified)	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	20	5	0

Weak Foodborne Outbreaks: detailed data

Causative agent	FBO nat. code	Outbreak type	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human		N deaths
											N hosp.	cases	
Bacillus - B. cereus	330	General	Buffet meals	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	8	0	0
	353	Household / domestic kitchen	Unknown	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	3	0	0
	355 484	General	Mixed food	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		2	5	0	0
	370	Household / domestic kitchen	Mixed food	Unknown	Take-away or fast-food outlet	Take-away or fast-food outlet	Unknown	NOT AVAILABLE		1	5	0	0
	502	Household / domestic kitchen	Crustaceans, shellfish, molluscs and products thereof	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	0	0
	585	Household / domestic kitchen	Buffet meals	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	3	0	0
Calicivirus - norovirus (Norwalk-like virus)	345	Household / domestic kitchen	Mixed food	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	0	0
	498	General	Mixed food	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE	descriptive environmental evidence	1	3	0	0
	537	General	Mixed food	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	50	0	0
Campylobacter - Campylobacter spp., unspecified	488	Household / domestic kitchen	Bovine meat and products thereof	Unknown	Others	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	2	0	0

Causative agent	FBO nat. code	Outbreak type	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Escherichia coli, pathogenic - Verotoxigenic E. coli (VTEC)	400	Household / domestic kitchen	Bovine meat and products thereof	Unknown	Household	Retail	Unknown	NOT AVAILABLE	descriptive environmental evidence	1	2	1	0
Listeria - L. monocytogenes - L. monocytogenes serovar 1/2a	495	Household / domestic kitchen	Bovine meat and products thereof	Unknown	Household	Retail	Unknown	NOT AVAILABLE		1	2	1	0
Salmonella - S. Enteritidis	359	Household / domestic kitchen	Eggs and egg products	Unknown	Household	Retail	Unknown	NOT AVAILABLE		1	2	1	0
Salmonella - S. Hadar	473	General	Turkey meat and products thereof	Unknown	Residential institution (nursing home or prison or boarding school) (not specified)	Canteen or workplace catering	Unknown	NOT AVAILABLE		1	10	0	0
Staphylococcal enterotoxins - Enterotoxin A	340	Household / domestic kitchen	Mixed food	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	3	0	0
Staphylococcal enterotoxins - Enterotoxin, unspecified	360	General	Mixed food	Unknown	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Restaurant or Cafe or Pub or Bar or Hotel or Catering service	Unknown	NOT AVAILABLE		1	14	0	0
Unknown		NOT AVAILABLE	Buffet meals	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		7	27	3	0
								descriptive environmental evidence		5	46	1	0
			Mixed food	Descriptive epidemiological evidence		NOT AVAILABLE	Unknown	NOT AVAILABLE		2	11	2	0
				Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		156	688	12	0
								descriptive environmental evidence		22	60	1	0
			Other foods	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		1	2	1	0
			Bakery products	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		9	24	0	0

Causative agent	FBO nat. code	Outbreak type	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths			
Unknown	NOT AVAILABLE		Tap water, including well water	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		1	4	0	0			
			Drinks, including bottled water	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		2	8	0	0			
			Fruit, berries and juices and other products thereof	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		1	3	0	0			
			Canned food products	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		2	4	1	0			
			Vegetables and juices and other products thereof	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		5	15	0	0			
			Crustaceans, shellfish, molluscs and products thereof	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		7	17	0	0			
			Fish and fish products	Unknown					NOT AVAILABLE	Unknown	NOT AVAILABLE		11	36	1	0
												descriptive environmental evidence	2	4	0	0
			Broiler meat (Gallus gallus) and products thereof	Unknown					NOT AVAILABLE	Unknown	NOT AVAILABLE		10	43	0	0
												descriptive environmental evidence	2	8	0	0
Other or mixed red meat and products thereof	Unknown					NOT AVAILABLE	Unknown	NOT AVAILABLE		11	32	0	0			
									descriptive environmental evidence	1	2	0	0			
Pig meat and products thereof	Unknown					NOT AVAILABLE	Unknown	NOT AVAILABLE		10	29	0	0			

Causative agent	FBO nat. code	Outbreak type	More food vehicle info	Nature of evidence	Setting	Place of origin of problem	Origin of food vehicle	Contributory factors	Comment	N outbreaks	N human cases	N hosp.	N deaths
Unknown		NOT AVAILABLE	Bovine meat and products thereof	Descriptive epidemiological evidence		NOT AVAILABLE	Unknown	NOT AVAILABLE		1	15	0	0
				Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		27	71	0	0
									descriptive environmental evidence	1	2	0	0
			Eggs and egg products	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		1	14	0	0
			Dairy products (other than cheeses)	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		3	8	0	0
			Unknown	Unknown		NOT AVAILABLE	Unknown	NOT AVAILABLE		31	98	2	0
									descriptive environmental evidence	6	15	0	0

ANTIMICROBIAL RESISTANCE TABLES FOR CAMPYLOBACTER

Table Antimicrobial susceptibility testing of Campylobacter - C. coli in Meat from broilers (Gallus gallus) - carcass - spent hens

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	8	16	2
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	30	30	30	30	30	30
N of resistant isolates	0	3	16	2	16	19
MIC						
0.12	5		7			
0.25	11	1	7			
0.5	14	6				10
1		11		13	1	1
2		9		12		
4			1	3	6	
8			9		7	
16		3	6			
32					1	1
64					15	18
128				2		

Table Antimicrobial susceptibility testing of Campylobacter - C. coli in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	8	16	2
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	27	27	27	27	27	27
N of resistant isolates	0	8	14	4	15	20
MIC						
0.12	1		6			
0.25	8		6			
0.5	16		1			5
1	2	11		7		2
2		8		7		
4				7	7	
8			4	2	4	
16		8	10		1	
32				1	1	
64					14	20
128				3		

Table Antimicrobial susceptibility testing of Campylobacter - C. coli in Meat from broilers (Gallus gallus) - fresh (not specified)

Sampling Stage: Unspecified

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	8	16	2
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	1	1	1	1	1	1
N of resistant isolates	0	0	1	0	1	1
MIC						
0.5	1					
1				1		
2		1				
16			1			
64					1	1

Table Antimicrobial susceptibility testing of Campylobacter - C. coli in Meat from pig - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	8	16	2
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	16	16	16	16	16	16
N of resistant isolates	0	14	6	5	6	13
MIC						
0.12			6			
0.25	1		3			
0.5	13		1			2
1	2			1		1
2		2		6		
4				2	1	
8			2	2	7	
16		14	4		2	
32						1
64					6	12
128				5		

Table Antimicrobial susceptibility testing of Campylobacter - C. coli in Meat from broilers (Gallus gallus) - meat products (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	8	16	2
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	18	18	18	18	18	18
N of resistant isolates	0	3	15	4	15	14
MIC						
0.12			2			
0.25	2	1	1			
0.5	16					3
1		13		7		
2		1		6		1
4			1	1	2	
8			3		1	
16		3	11			
64					15	14
128				4		

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Meat from broilers (Gallus gallus) - carcass - spent hens

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	76	76	76	76	76	76
N of resistant isolates	1	1	32	1	33	32
MIC						
0.12	21		37			
0.25	40	6	6			
0.5	14	28	1			44
1		32	2	51	1	
2		8		20	5	3
4		1		4	33	4
8	1		5		3	
16		1	25		1	
32					3	
64					30	25
128				1		

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Gallus gallus (fowl) - broilers (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	92	92	92	92	92	92
N of resistant isolates	0	0	56	1	56	48
MIC						
0.12	18		31			
0.25	50	5	3			
0.5	24	30	2			44
1		53		66	1	
2		3		22	4	2
4		1	2	3	19	
8			20		9	
16			34		3	3
32					1	2
64					55	41
128				1		

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Meat from broilers (Gallus gallus) - carcass (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	132	132	132	132	132	132
N of resistant isolates	0	0	82	1	79	72
MIC						
0.12	28		45			
0.25	68	6	3			
0.5	36	44	2			58
1		67		94		2
2		15		32	6	1
4			1	5	39	
8			19	1	7	
16			62		1	
32					1	3
64					78	68

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Meat from broilers (Gallus gallus) - fresh (not specified)

Sampling Stage: Unspecified

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	2	2	2	2	2	2
N of resistant isolates	0	0	1	0	1	1
MIC						
0.12			1			
0.25	2					
0.5		1				1
1		1		2		
2					1	
16			1			
64					1	1

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Meat from broilers (Gallus gallus) - fresh (not specified)

Sampling Stage: Unspecified

Sampling Type: unknown

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	2	2	2	2	2	2
N of resistant isolates	0	0	2	0	2	2
MIC						
0.12	1					
0.25	1					
0.5		2				
1				2		
16			2			
64					2	2

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Meat from pig - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0
MIC						
0.12			1			
0.25	1					
0.5						1
1		1				
2				1		
8					1	

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Meat from broilers (Gallus gallus) - meat products (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	67	67	67	67	67	67
N of resistant isolates	0	0	42	0	42	37
MIC						
0.12	18		21			
0.25	34	6	4			
0.5	15	23				28
1		28		43		2
2		10		20	6	1
4				4	14	
8			10		5	
16			32			
32						1
64					42	35

Table Antimicrobial susceptibility testing of Campylobacter - C. jejuni in Meat from broilers (Gallus gallus) - meat products (not specified)

Sampling Stage: Retail

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	9	9	9	9	9	9
N of resistant isolates	0	0	7	0	7	6
MIC						
0.12	1		2			
0.25	2					
0.5	6	2				3
1		6		6		
2		1		2	1	
4				1	1	
8			2			
16			5			
64					7	6

Table Antimicrobial susceptibility testing of Campylobacter - Thermophilic Campylobacter spp., unspecified in Meat from broilers (Gallus gallus) - carcase - spent hens

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Aminoglycosides - Streptomycin	Fluoroquinolones - Ciprofloxacin	Macrolides - Erythromycin	Quinolones - Nalidixic acid	Tetracyclines - Tetracycline
ECOFF	2	4	0.5	4	16	1
Lowest limit	0.12	0.25	0.12	1	1	0.5
Highest limit	16	16	16	128	64	64
N of tested isolates	1	1	1	1	1	1
N of resistant isolates	0	0	1	1	1	1
MIC						
0.5	1					
1		1				
16			1			
64					1	1
128				1		

ANTIMICROBIAL RESISTANCE TABLES FOR SALMONELLA

Table Antimicrobial susceptibility testing of Salmonella - Not typeable in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified) Sampling Type: environmental sample - boot swabs Sampling Context: Control and eradication programmes
 Sampler: Industry sampling Sampling Strategy: Objective sampling Programme Code: AMR MON pn12
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Cephalosporins - Cefoxitin	Cephalosporins + β lactamase inhibitores - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitores - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
Ceftazidime synergy test	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	0	0	0	0	0
MIC										
<=0.015	1									
0.06			1							
0.25		1						1		
0.5									1	
2				1			1			
4					1					
8						1				1

Table Antimicrobial susceptibility testing of Salmonella - Not typeable in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified) Sampling Type: environmental sample - boot swabs Sampling Context: Control and eradication programmes
 Sampler: Industry sampling Sampling Strategy: Objective sampling Programme Code: AMR MON
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	1	0	4	0	0	4	0	4	1	1	4
MIC														
<=0.015						1								

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	1	0	4	0	0	4	0	4	1	1	4
MIC														
<=0.03			5											
<=0.25				4			1							1
0.25						2								
<=0.5	5				4									
0.5						2	3							
<=1										4				
1							1							
<=2													2	
2				1	1				1	1				
<=4											1			
4								4					2	
<=8		3												
8								1						
16		2										1		
32									1			1		
>32														4
64												1		
>64									3				1	
128												1		
>128											4			
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - Not typeable in Gallus gallus (fowl) - broilers - during rearing period

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	1	0	0	0	0	1	0	0	0	0	1	1	0	1
<=0.03			1											
<=0.25				1			1							
<=0.5					1									
0.5						1								
<=1								1						
<=2													1	
2										1				
<=8		1												
8								1						
32	1													
>32														1
>128											1			
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - Not typeable in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	1	0	1
0.03						1								
0.06			1											
<=0.25				1			1							
<=0.5					1									
<=1								1		1				
<=2													1	
2	1													
<=4											1			
<=8		1												
8								1						
>32														1
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - Not typeable in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015						1								
<=0.03			1											
<=0.25				1			1							1
<=0.5	1				1									
<=1									1	1				
<=2								1					1	
<=4											1			
<=8		1												
64												1		

Table Antimicrobial susceptibility testing of Salmonella - Not typeable in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			1											
0.03						1								
<=0.25				1										
<=0.5	1				1									
0.5							1							1
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
256												1		

Table Antimicrobial susceptibility testing of Salmonella - Other serovars in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON pnl2

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitors - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitors - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
Ceftazidime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	0	1	0	0	0
MIC										
<=0.015	1									
<=0.03			1							
0.25		1						1		
0.5									1	
2				1	1					
4							1			
8						1				
16										1

Table Antimicrobial susceptibility testing of Salmonella - Other serovars in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	1	0	0	1	0	1	0	0	1
MIC														
<=0.03			1											
<=0.5	1													
0.5						1								

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	1	0	0	1	0	1	0	0	1
MIC														
1							1							
2				1						1				
4					1			1					1	
16		1												
>32														1
64												1		
>64									1					
>128											1			

Table Antimicrobial susceptibility testing of Salmonella - Other serovars in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	0	0	0	0	6	0	3	0	5	4	3	1	5
MIC														
<=0.03			5											
0.06			1											
0.12						1								
<=0.25				6									2	1
0.25						1								
<=0.5	6				5									
0.5						2							1	
<=1								1	5					
1					1	2								2
<=2												3		
2								1	1				1	
4							1	1						
<=8		4												
8							3			1				
16		2					2							
32											1			5
64								3				3		
128										5				
256											1			
1024												4		

Table Antimicrobial susceptibility testing of Salmonella - S. 1,4,12:i:- in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	0	0	1	0	1	1	0	1
MIC														
<=0.03			1											
<=0.5	1			1										
0.5				1										
1						1	1							
2										1				
4													1	
8								1						
16		1												
>32														1
>64									1					
>128											1			
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 3,10:-: - in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5					1									
<=1									1	1				
1	1													
<=2														
<=4											1		1	
<=8		1												
8								1						
>32														1
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 3,19:-: - in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	1	0	0	1	0	0	1	0	2
MIC														
<=0.015						1								
<=0.03			2											
<=0.25				2			2							
<=0.5	2				2									
0.5						1								
<=1									1	2				
<=2													2	
<=4											1			
4								1						
<=8		2												
8								1			1			
>32														2
64												1		
>64									1					
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 3,19:-:- in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
16												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. 4,12:-: - in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	0	0	1	0	1	0	0	1
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
0.5						1								
<=1										1				
<=2													1	
<=8		1												
8								1						
32												1		
>32														1
>64									1					
>128											1			

Table Antimicrobial susceptibility testing of Salmonella - S. 4,12:-: - in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	3	0	0	2	0	3	2	0	3
<=0.03			3											
<=0.25				2										
<=0.5	3				2									
0.5				1		1	2							
<=1										3				
1					1	2	1							
<=2													1	
4									1				2	
8								2						
16		3						1				1		
>32														3
>64									2					
>128											3			
>1024												2		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,12:i:- in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	3	0	0	3	3	3
MIC														
<=0.015						1								
<=0.03			2											
0.03						2								
0.06			1											
<=0.25				3			1							
<=0.5	2				3									
0.5							2							
<=1										1				
1	1													
2										2				
<=4											3			
4								3						
<=8		3												
>32														3
>64									3				3	
>1024												3		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,12:i:- in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	2	0	0	2	2	2
MIC														
<=0.03			1											
0.03						2								
0.06			1											
<=0.25				2			2							
<=0.5	2				2									
<=1										2				
<=4											2			
<=8		2												
8								2						
>32														2
>64									2				2	
>1024												2		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,12:i:- in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	24	24	24	24	24	24	24	24	24	24	24	24	24	24
N of resistant isolates	0	3	0	0	0	0	0	0	22	2	0	22	19	24
<=0.015						5								
<=0.03			22											
0.03						15								
0.06			2			4								
<=0.25				24			14							
<=0.5	22				23									
0.5							8							
<=1									1	18				
1	2				1		2							
<=2													3	
2									1	4				
<=4											18			
4								10		2			1	
<=8		18												
8								10			6		1	
16		3						4						
32		1										1		
>32														24
64		1												
>64									22				19	
128		1										1		
512												1		
>1024												21		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,5,12:i:- in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	0	0	1	0	0	2	0	1	1	2	5
<=0.015						1								
<=0.03			5											
0.03						3								
<=0.25				5			1							
<=0.5	5				4									
0.5						1	4							
<=1									3	1				
1					1									
<=2													3	
2										4				
<=4											4			
4								3						
<=8		5												
8								2						
16												2		
32												1		
>32														5
>64									2				2	
128												1		
>128											1			
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,5,12:i:- in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	1	0	0
<=0.015						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
0.5							1							
<=1										1				
<=2													1	
<=4											1			
4								1						
<=8		1												
>64									1					
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,5,12:i:- in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	3	0	0	3	3	3
MIC														
<=0.03			2											
0.03						3								
0.06			1											
<=0.25				3			3							
<=0.5	3				3									
2										3				
<=4											3			
4								1						
<=8		3												
8								2						
>32														3
>64									3				3	
>1024												3		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,5:b in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON pni2

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitors - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitors - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent
Ceftazidime synergy test	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	0	1	0	0	0
MIC	0.03	1								
	0.06		1							
	0.25							1		
	0.5	1								
	1								1	
	2			1	1					
	4						1			
	8					1				
	16									1

Table Antimicrobial susceptibility testing of Salmonella - S. 4,5:b in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	1	0	2	0	0	2	0	2	1	1	2
MIC	<=0.03		2											
	<=0.25			1										

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	1	0	2	0	0	2	0	2	1	1	2
MIC														
<=0.5	2				1									
<=1										2				
1						2	2							
2				1	1									
4													1	
8								1						
16		2						1					1	
>32														
64												1		
>64									2					
>128											2			
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 4,5:i:- in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	1	2	1
MIC														
<=0.03			2											
0.03						1								
0.06						1								
<=0.25				2			2							1
<=0.5	2				2									
<=1									1	1				
2										1				
<=4											2			
4								2						
<=8		2												
32												1		
>32														1
>64									1				2	
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 4:i:- in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	1	1	0	1	1	1
<=0.015						1								
0.06			1											
<=0.25				1			1							
<=0.5	1				1									
<=4											1			
4								1						
<=8		1												
>16										1				
>32														1
>64									1				1	
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 6,7:-: in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 6,7:-: in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
0.5							1							
<=1										1				
<=2													1	
2									1					
<=4											1			
4								1						
<=8		1												
256												1		

Table Antimicrobial susceptibility testing of Salmonella - S. 6,7:- in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1										
<=0.5	1				1									
0.5							1							
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
16								1						
>32														1
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Abony in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
0.5														1
<=1								1						
<=2													1	
2										1				
<=4											1			
4								1						
<=8		1												
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Agona in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5					1									
1	1													
<=2													1	
2									1	1				
<=4											1			
<=8		1												
8								1						
32												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. Agona in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13
N of resistant isolates	1	0	0	0	0	1	0	0	1	0	0	2	0	8
MIC														
<=0.015						9								
<=0.03			11											
0.03						3								
0.06			2											
<=0.25				13			6							4
0.25						1								
<=0.5	11				12									
0.5							7							1
<=1									6	9				
1	1				1									
<=2													13	
2									6	4				
<=4											12			
4								11						
<=8		13												
8								2						
16	1										1			
32												3		
>32														8
64												2		
>64									1					
128												5		
256												1		
>1024												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Agona in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5							1							1
<=1										1				
<=2													1	
2									1					
<=4											1			
4								1						
<=8		1												
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Agona in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.015						3								
<=0.03			5											
0.03						2								
<=0.25				5									4	1
<=0.5	5				5									
0.5													1	4
<=1								5	5					
<=2												5		
<=4										5				
4							2							
<=8		5												
8							3							
32											2			
64											3			

Table Antimicrobial susceptibility testing of Salmonella - S. Agona in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.015						1								
<=0.03			2											
0.03						1								
<=0.25				2			2							
<=0.5	2				2									
<=1									2	2				
<=2														
<=4											2		2	
<=8		2												
8								2						
>32														2
64												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Anatum in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.015						1								
<=0.03			2											
0.03						1								
<=0.25				2			2							
<=0.5	2				2									
<=1									2	2				
<=2														
<=4											2		2	
<=8		2												
8								2						
>32														2
64												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Bareilly in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.015						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5							1							1
<=1										1				
<=2													1	
2									1					
<=4											1			
4								1						
<=8		1												
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Braenderup in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			1											
0.03						1								
<=0.25				1										1
<=0.5	1				1									
0.5							1							
<=1										1				
<=2													1	
2									1					
<=4											1			
4								1						
<=8		1												
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Brandenburg in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
<=1									1					
1													1	
<=2												1		
2								1						
<=4										1				
4							1							
<=8		1												
32											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Brandenburg in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcass swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						2								
<=0.03			6											
0.03						3								
0.06						1								
<=0.25				6									1	5
<=0.5	4				6									
0.5													5	1
<=1								2	5					
1	2													
<=2												4		
2								4	1					
<=4										5				
4							5					2		
<=8		6												
8										1				
16							1							
32											4			
64											2			

Table Antimicrobial susceptibility testing of Salmonella - S. Brandenburg in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
0.5													1	
<=1									1					
<=2												1		
2								1						
<=4										1				
4							1							
<=8		1												
32											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Brandenburg in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			1											
0.03						1								
<=0.25				1										
<=0.5	1				1									
0.5													1	1
<=2												1		
2								1	1					
<=4										1				
8							1							
16		1												
64											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Brandenburg in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	2	0	0	0	1
MIC														
<=0.015						2								
<=0.03			2											
<=0.25				2			2							1
<=0.5	1				2									
1	1													
<=2													2	
2									2					
<=4											2			
4										2				
<=8		2												
8								2						
>32														1
64												1		
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Cerro in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	1	0	0	1	0	0	2	0	3
MIC														
<=0.03			3											
0.03						2								
<=0.25				3			2							
0.25						1								
<=0.5	2				3									
0.5							1							
<=1									2	3				
1	1													
<=2													3	
<=4											2			
4								1						
<=8		3												
8								2						
16											1			
>32														3
>64									1					
256												1		
512													1	
>1024													1	

Table Antimicrobial susceptibility testing of Salmonella - S. Cerro in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.015						1								
<=0.03			2											
0.03						1								
<=0.25				2										2
<=0.5	2				2									
0.5							2							
<=1									1	2				
<=2													2	
2									1					
<=4											2			
4								2						
<=8		2												
256												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Chester in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	1	0	0	0	1
MIC														
<=0.03			2											
0.03						2								
<=0.25				2			1							1
<=0.5	2				2									
0.5							1							
<=1									1	1				
<=2													2	
2									1					
<=4											2			
4														
<=8		2												
8								1						
>32														1
64												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Chester in Turkeys - fattening flocks (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	0	0	0	0	0	1	2	0	1	1	4
MIC														
<=0.015						2								
<=0.03			5											
0.03						3								
<=0.25				5			2							
<=0.5	5				5									
0.5							1							1
1							2							
<=2													3	
2									3	3				
<=4											4			
4								4	1	2			1	
<=8		4												
8								1			1			
16		1												
32												2		
>32														4
64												2		
>64									1				1	
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Chester in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	0	0	0
MIC														
<=0.015						1								
<=0.03			2											
0.03						1								
<=0.25				2										2
<=0.5	2				2									
0.5													2	
<=1								2						
<=2												1		
2									1					
<=4										2				
4									1			1		
<=8		2												
8							2							
64											2			

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
>32														1
256												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	2	0	1
MIC														
<=0.015						2								
<=0.03			2											
<=0.25				2			2							
<=0.5	2				2									
0.5														1
<=1									1	2				
<=2													2	
<=4											2			
4								1						
<=8		2												
8								1						
>32														1
>64									1					
>1024												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	4	4	4	4	4	4	4	3	4	4	4	4	4	4
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						4								
<=0.03			4											
<=0.25				4									1	1
<=0.5	3				4									
0.5													2	3
<=1								2	3					
1	1												1	
<=2												4		
2								1	1					
<=4										4				
4							3							
<=8		2												
8							1							
16		2												
32											4			

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcass swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	31	31	31	31	31	31	31	31	31	31	31	31	31	31
N of resistant isolates	1	0	0	0	0	3	1	0	1	3	2	1	4	1
MIC														
0.016						13								
<=0.03			30											
0.03						14								
0.06			1			1								
<=0.25				31									9	12
0.25						1								
<=0.5	27				31									
0.5						1							14	18
<=1								14	16					
1	2												4	
<=2												21		
2	1					1		15	14				3	
<=4										26				
4							8	2				9		
<=8		21												
8							20			2			1	
16	1	10					2		1					
32											13	1		1
64							1				14			
128										3	2			
512											1			
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5													1	1
<=1								1	1					
<=2												1		
<=4										1				
4							1							
16		1												
64											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	2	0	0
MIC														
0.016						2								
<=0.03			2											
<=0.25				2										
<=0.5	2				2									
0.5													1	2
<=1								2	1					
1													1	
2									1					
<=4										2				
4							1							
<=8		1												
8							1							
16		1												
64											2	2		

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	11	11	11	11	11	11	11	11	11	11	11	11	11	11
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	5	1	10
MIC														
<=0.015						6								
<=0.03			11											
0.03						5								
<=0.25				11			7							
<=0.5	11				11									
0.5							4							1
<=1									8	8				
<=2													10	
2									3	3				
<=4											11			
4								3						
<=8		10												
8								8						
16		1										1		
>32														10
64												1	1	
128												1		
256												3		
512												2		
>1024												3		

Table Antimicrobial susceptibility testing of Salmonella - S. Derby in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1										1				
<=2													1	
2									1					
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Dublin in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	4	0	0	0	1	0	0	0	6	1	5	0	5
<=0.015						4								
<=0.03			5											
0.03						1								
0.06			1											
<=0.25				6			5							
0.25						1								
<=0.5	6				6									
0.5							1							1
<=1									6					
<=2													6	
<=4											5			
4								6						
<=8		2												
8										6				
32												1		
>32														5
>128		4									1			
>1024												5		

Table Antimicrobial susceptibility testing of Salmonella - S. Dublin in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	1	0	0	0	0	0	0	0	1	0	1	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1					
<=2													1	
<=4											1			
4										1				
8								1						
>32														1
>128		1												
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
32												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	0	0	0	0	0	0	3	0	0	0	5
MIC														
<=0.015						3								
<=0.03			4											
0.03						2								
0.06			1											
<=0.25				5			3							
<=0.5	5				5									
0.5							2							
<=1									2					
<=2								1					5	
2									3	2				
<=4											5			
4								3		2				
<=8		5												
8								1		1				
32												1		
>32														5
64												2		
128												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	0	3	0	0	0	2
MIC														
<=0.015						2								
<=0.03			3											
0.03						1								
<=0.25				3			2							1
<=0.5	3				3									
0.5							1							
<=1									1					
<=2													3	
2									2					
<=4											3			
4								2		1				
<=8		3												
8								1		2				
>32														2
64												2		
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - laying hens - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	0	0	0	0	0	0	0	0	6	0	0	0	5
MIC														
<=0.015						1								
<=0.03			6											
0.03						5								
<=0.25				6			5							
<=0.5	6				6									
0.5							1							1
<=1									3					
<=2													6	
2									3					
<=4											6			
4								1		5				
<=8		6												
8								5		1				
>32														5
64												5		
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	12	12	12	12	12	12	12	12	12	12	12	12	12	12
N of resistant isolates	0	0	0	0	0	0	0	0	0	1	0	0	0	6
<=0.015						3								
<=0.03			12											
0.03						9								
<=0.25				12			8							2
<=0.5	12				12									
0.5							4							4
<=1									8	6				
<=2								1					12	
2									4	5				
<=4											12			
4								5						
<=8		12												
8								6		1				
16												1		
32												2		
>32														6
64												2		
128												7		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - broilers - during rearing period

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			2											
0.03						2								
<=0.25				2			2							
<=0.5	2				2									
0.5														2
<=1									2	2				
<=2													2	
<=4											2			
4								2						
<=8		2												
128												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	36	36	36	36	36	36	36	36	36	36	36	36	36	36
N of resistant isolates	0	0	0	0	0	4	0	0	17	4	0	0	0	0
MIC														
0.016						18								
<=0.03			36											
0.03						14								
<=0.25				36									8	14
0.25						4								
<=0.5	34				36									
0.5													24	20
<=1								11	7					
1	2												4	2
<=2							1					19		
2								24	12					
<=4										26				
4							28	1	12			17		
<=8		36												
8							7		5	6				
32											23			
64											13			
128										4				

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			1											
0.03						1								
<=0.25				1									1	
<=0.5	1				1									
0.5														1
<=2												1		
2								1	1					
4							1							
8										1				
16		1												
32											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	0	0	0
MIC														
0.016						1								
<=0.03			2											
0.03						1								
<=0.25				2										1
<=0.5	2				2									
0.5													2	1
<=1								2						
<=2												2		
2									1					
<=4										2				
4							1		1					
<=8		2												
8							1							
32											2			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	0	0	0	0	0	0	0	6	0	0	0	0	0
0.016						2								
<=0.03			6											
0.03						4								
<=0.25				6									1	4
<=0.5	6				6									
0.5													5	2
<=1								1						
<=2												6		
2								5						
<=4										6				
4							6		5					
<=8		6												
8									1					
32											6			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: unknown

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	0	0	0
<=0.03			1											
0.03						1								
<=0.25				1										
<=0.5	1				1									
0.5													1	1
2								1						
<=4										1				
4							1		1			1		
<=8		1												
32											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1										1				
<=2													1	
2									1					
<=4											1			
4								1						
<=8		1												
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse Sampling Type: food sample - neck skin Sampling Context: Monitoring - EFSA specifications
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON pnl2
 Analytical Method: Micromethod dilution (in microtiter plate) (not specified)
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitores - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitores - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	Negative/Absent	NOT AVAILABLE	NOT AVAILABLE
Ceftazidime synergy test	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	Negative/Absent	NOT AVAILABLE	NOT AVAILABLE
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	64
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0
MIC										
<=0.015	1									
<=0.03			1							
<=0.06								1		
0.12				1						
<=0.25					1		1			
0.25		1							1	
2						1				
8										1

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse Sampling Type: food sample - neck skin Sampling Context: Monitoring - EFSA specifications
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON
 Analytical Method: Micromethod dilution (in microtiter plate) (not specified)
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracycline - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	11	11	11	11	11	11	11	11	11	11	11	11	11	11
N of resistant isolates	0	0	0	1	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.015						9								
<=0.03			10											
0.03						2								

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	11	11	11	11	11	11	11	11	11	11	11	11	11	11
N of resistant isolates	0	0	0	1	0	0	0	0	0	0	0	0	0	0
MIC														
0.06			1											
<=0.25				10									5	5
<=0.5	11				9									
0.5													6	4
1				1	1									2
<=2							1					9		
2					1			11	11					
<=4										8				
4							10					2		
<=8		9												
8										3				
16		2												
32											9			
64											2			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
0.5														1
<=2													1	
2									1	1				
<=4											1			
4								1						
<=8		1												
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - unspecified - before slaughter

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.03			1											
0.03						3								
0.06			2											
<=0.25				3			2							1
<=0.5	3				3									
0.5							1							
<=1									2	1				
<=2													3	
2									1	2				
<=4											3			
<=8		3												
8								3						
>32														2
64												2		
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis in Gallus gallus (fowl) - broilers - day-old chicks

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - delivery box liner

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	1	0	0	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=2													1	
2									1					
<=4											1			
4										1				
<=8		1												
8								1						
32												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis - Not typeable in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
N of resistant isolates	1	0	0	0	0	0	0	2	0	0	3	0	0	3
0.016						3								
<=0.03			3											
0.03						1								
0.06			1											
<=0.25				4									3	1
<=0.5	3				4									
0.5													1	
<=2												2		
2								2	4					
<=4										4				
4							3					2		
<=8		4												
8	1						1							
32											1			3
64								2						
1024											3			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis - Not typeable in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	2	0	0	0	0	0	3	0	0	3	2	0	1
<=0.03			5											
0.03						4								
0.06						1								
<=0.25				4									3	4
<=0.5	5				5									
0.5				1									1	
<=1									1					
1													1	
<=2												2		
2								2	4					
<=4										3				
4							1					1		
<=8		2												
8							3			1				
16		1					1			1	1			
32		1												1
64		1						3			1	2		
1024											3			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis - Not typeable in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	1	0	0	1	1	0	1
MIC														
<=0.03			1											
0.06						1								
<=0.25				1										
<=0.5	1				1									
1													1	
2									1					
8							1			1				
16		1												
32														1
64								1				1		
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis - Not typeable in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	1	1	1	0	1	0	0	0	1
MIC														
<=0.03			2											
0.03						1								
0.12						1								
<=0.25				2									2	1
<=0.5	1				2									
<=1								1						
<=2												1		
2	1								2					
<=4										1				
4							1						1	
<=8		2												
32							1				1			1
64								1						
128										1	1			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis - Not typeable in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						1								
<=0.03			1											
<=0.25				1									1	1
<=0.5	1				1									
<=1								1						
<=2												1		
2														
<=4										1				
4							1							
<=8		1												
16											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Enteritidis - Not typeable in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	15	15	15	15	15	15	15	15	15	15	15	15	15	15
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	1	0	0	1
<=0.015						8								
<=0.03			15											
0.03						7								
<=0.25				15									5	6
<=0.5	13				15									
0.5													8	7
<=1									13					
1	2												2	1
<=2												7		
2								14	1					
<=4										15				
4							15	1				8		
<=8		12												
8													1	
16		3												
32											11			1
64											2			
256											1			
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Gaminara in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.015						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
0.5							1							
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Gloucester in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	1	1	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1										1				
4								1						
<=8		1												
8											1			
>32														1
>64									1				1	
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Goldcoast in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	1	1	2
MIC														
<=0.015						1								
<=0.03			2											
0.03						1								
<=0.25				2										
<=0.5	2				1									
0.5							2							
<=1										1				
1					1									
<=2													1	
2									1	1				
<=4											2			
4								1						
<=8		2												
8								1						
>32														2
64												1		
>64									1				1	
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Hadar in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	0	0	0	0	1	0	1	1
MIC														
<=0.03			1											
<=0.25				1			1							
<=0.5					1									
<=1									1	1				
1	1													
2						1								
4								1						
<=8		1												
32												1		
>32														1
64													1	
>128											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Havana in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
<=0.015						2								
<=0.03			2											
<=0.25				2			2							
<=0.5	2				2									
<=1									2	2				
<=2													2	
<=4											2			
4								1						
<=8		2												1
8								1						
32												1		
>32														2

Table Antimicrobial susceptibility testing of Salmonella - S. Idikan in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Idikan in Gallus gallus (fowl) - broilers - day-old chicks

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - delivery box liner

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
>32														1
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Indiana in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
0.016						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
0.5													1	
<=1								1	1					
<=2							1					1		
<=4										1				
<=8		1												
32											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	1	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5							1							
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
>32														1
512												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.015						2								
<=0.03			2											
<=0.25				2			1							
<=0.5	2				2									
0.5							1							
<=1									1	1				
<=2													2	
2									1	1				
<=4											2			
4								1						
<=8		2												
8								1						
>32														2
64												1		
256												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
<=0.015						3								
<=0.03			3											
<=0.25				3			1							
<=0.5	3				3									
0.5							2							1
<=1									2	3				
<=2													3	
2									1					
<=4											3			
4								2						
<=8		3												
8								1						
>32														2
128												2		
256												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Gallus gallus (fowl) - laying hens - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.03			2											
0.03						2								
<=0.25				2			2							
<=0.5	2				2									
<=1									1	1				
<=2													2	
2									1	1				
<=4											2			
<=8		2												
8								2						
>32														2
64												1		
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	11	11	11	11	11	11	11	11	11	11	11	11	11	11
N of resistant isolates	0	0	0	0	0	7	3	0	0	0	7	7	5	7
MIC														
<=0.015						3								
<=0.03			10											
0.03						1								
0.06			1											
0.12						1								
<=0.25				11			2							
<=0.5	11				10									
0.5						4	2							4
<=1									1	10				
1					1	2	4							
<=2													4	
2							3		4	1				
<=4											4			
4								5	6				2	
<=8		4												
8								4						
16		7						2						
>32														7
64												2		
>64													5	
128											1	2		
>128											6			
>1024												7		

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	0	0	0	0	1	1	1	1
MIC														
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
<=1										1				
1						1	1							
2									1					
16		1												
>32														1
>64													1	
>128											1			
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
N of resistant isolates	0	0	0	0	0	3	0	0	0	3	3	3	3	2
MIC														
<=0.03			4											
0.03						1								
0.12						1								
<=0.25				4										1
0.25						1								
<=0.5	4				4									
0.5						1							1	1
<=1									3					
<=2												1		
2								2	1				2	
<=4										1				
4							2	2					1	
<=8		2												
8							2							
16		2												
32											1			2
64												3		
128										3				
1024											3			

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5														1
<=1									1					
1													1	
<=2												1		
2								1						
<=4										1				
4							1							
<=8		1												
64											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	1	0	0	0	1	1	1	1	0
MIC														
0.016						1								
<=0.03			2											
<=0.25				2										2
<=0.5	2				2									
0.5						1								1
<=1									1					
<=2												1		
2								1	1					1
<=4										1				
4							1	1						
<=8		1												
8							1							
16		1												
64												1		
128										1	1			
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	1	0	0	0	5	0	1	0	5	6	5	1	6
MIC														
<=0.03			5											
0.03						1								
0.06			1											
0.12						2								
<=0.25				6										
0.25						3								
<=0.5	6				6									
<=1									1					
1													5	
2								3	5				1	
<=4										1				
4							1	2				1		
<=8		5												
8							5							
32		1												6
64								1				5		
128										5				
1024											6			

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
N of resistant isolates	0	0	0	0	0	3	0	0	0	3	3	3	1	3
<=0.015						1								
<=0.03			4											
0.12						2								
<=0.25				4									1	
0.25						1								
<=0.5	4				4									
0.5														1
<=1									4					
1													2	
<=2												1		
2								3					1	
<=4										1				
4							3	1						
<=8		2												
8							1							
16		2												
32														3
64														
128										3	1			
1024											3			

Table Antimicrobial susceptibility testing of Salmonella - S. Infantis in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1										1				
<=2													1	
2									1					
<=4											1			
4								1						
<=8		1												
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Jerusalem in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=2													1	
2									1	1				
<=4											1			
<=8		1												
8								1						
>32														1
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Kedougou in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
0.5														1
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
256												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Kentucky in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1					
<=2													1	
2										1				
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Kottbus in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.015						3								
<=0.03			3											
<=0.25				3			1							1
<=0.5	3				3									
0.5							2							
<=1								2	2					
<=2													3	
2									1	1				
<=4											3			
4								2						
<=8		3												
8								1						
>32														2
64												3		

Table Antimicrobial susceptibility testing of Salmonella - S. Lexington in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1					
<=2													1	
2										1				
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Lexington in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1					
<=2													1	
2										1				
<=4											1			
4								1						
<=8		1												
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Lexington in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	18	18	18	18	18	18	18	18	18	18	18	18	18	18
N of resistant isolates	0	0	0	0	0	0	0	1	0	0	0	0	0	16
<=0.015						5								
<=0.03			18											
0.03						13								
<=0.25				18			13							
<=0.5	15				18									
0.5							5							2
<=1									11	13				
1	2													
<=2													18	
2	1								7	5				
<=4											17			
4								5						
<=8		18												
8								12			1			
32								1				5		
>32														16
64												13		

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Gallus gallus (fowl) - broilers - during rearing period

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.015						2								
<=0.03			2											
<=0.25				2			2							
<=0.5	2				2									
<=1									1	2				
<=2													2	
2									1					
<=4											2			
4								1						
<=8		2												
8								1						
>32														2
64												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
0.016						1								
<=0.03			2											
0.03						1								
<=0.25				2										
<=0.5	2				2									
0.5													1	
<=1									1					
1													1	2
<=2												2		
2								2	1					
<=4										2				
4							2							
<=8		2												
32											1			
128											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4
N of resistant isolates	0	0	0	0	0	1	0	1	0	0	1	0	1	1
MIC														
0.016						1								
<=0.03			4											
0.03						2								
<=0.25				4										
<=0.5	4				4									
0.5						1							2	3
<=1								2	3					
1													1	
<=2												2		
2								1	1				1	
<=4										3				
4							4					2		
<=8		3												
16		1								1				
32											1			1
64								1			2			
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	1	0
MIC														
0.016						2								
<=0.03			2											
<=0.25				2										1
<=0.5	2				2									
0.5													1	
<=1								1	1					
1														1
<=2												2		
2								1	1				1	
<=4										2				
4						2								
<=8		2												
32											2			

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	3
MIC														
<=0.015						2								
<=0.03			3											
0.03						1								
<=0.25				3			3							
<=0.5	3				3									
<=1								2		1				
<=2													3	
2									1	2				
<=4											3			
<=8		3												
8								3						
32												2		
>32														3
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	1	0	1	0	0	1	1	0
MIC														
<=0.03			2											
0.03						2								
<=0.25				2										
<=0.5	2				2									
0.5													1	1
<=1								1						
1														1
<=2												1		
2								1	1					
<=4										2				
4							1						1	
<=8		1												
16		1												
32											1	1		
64							1				1			

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	7	7	7	7	7	7	7	7	7	7	7	7	7	7
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	7
<=0.015						2								
<=0.03			7											
0.03						5								
<=0.25				7			6							
<=0.5	7				7									
0.5							1							
<=1									6	7				
<=2													7	
2									1					
<=4											7			
4								2						
<=8		7												
8								4						
16								1						
32												2		
>32														7
64												5		

Table Antimicrobial susceptibility testing of Salmonella - S. Livingstone in Gallus gallus (fowl) - broilers - day-old chicks

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - delivery box liner

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5							1							
<=1										1				
<=2													1	
2									1					
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. London in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	1	2	4
MIC														
<=0.015						3								
<=0.03			5											
0.03						2								
<=0.25				5			3							
<=0.5	5				5									
0.5							1							1
<=1									4					
1							1							
<=2													3	
2										5				
<=4											5			
4								4						
<=8		5												
8								1						
>32														4
64												3		
>64									1				2	
128												1		
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Mbandaka in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	9	9	9	9	9	9	9	9	9	9	9	9	9	9
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	5
<=0.015						8								
<=0.03			9											
0.03						1								
<=0.25				9			6							2
<=0.5	9				9									
0.5							3							2
<=1									7	8				
<=2													9	
2									2	1				
<=4											9			
4								4						
<=8		9												
8								4						
16								1						
>32														5
64												4		
128												5		

Table Antimicrobial susceptibility testing of Salmonella - S. Mbandaka in Gallus gallus (fowl) - broilers - during rearing period

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.015						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
0.5							1							
<=1								1		1				
<=2													1	
<=4											1			
<=8		1												
8								1						
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Mbandaka in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015						6								
<=0.03			5											
0.06			1											
<=0.25				6			5							2
<=0.5	6				6									
0.5							1							4
<=1									2	5				
<=2													6	
2									4	1				
<=4											6			
4								5						
<=8		6												
8								1						
64												1		
128												4		
256												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Mbandaka in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
32												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. Mbandaka in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
>32														1
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Minnesota in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	12	12	12	12	12	12	12	12	12	12	12	12	12	12
N of resistant isolates	0	0	0	0	0	0	0	0	5	0	0	2	0	12
MIC														
<=0.03			11											
0.03						12								
0.06			1											
<=0.25				12			11							
<=0.5	12				12									
0.5							1							
<=1									7	6				
<=2													12	
2										6				
<=4											12			
<=8		10												
8								8						
16		2						4						
>32														12
64												1		
>64									5					
128												7		
256												2		
512												1		
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Montevideo in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
<=0.015						2								
<=0.03			1											
0.06			1											
<=0.25				2			2							
<=0.5	2				2									
<=2													2	
2									2	2				
<=4											2			
4								2						
<=8		2												
16												1		
32												1		
>32														2

Table Antimicrobial susceptibility testing of Salmonella - S. Montevideo in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<=0.015						1								
<=0.03			1											
<=0.25				1										1
<=0.5	1				1									
0.5							1							
<=1									1	1				
<=2													1	
4								1						
<=8		1												
8											1			
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Ohio in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Ohio in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5							1							
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Panama in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	1	0	0	1	1	1	0	0	1
0.06			1											
0.25						1								
<=0.5	1													
0.5							1							
<=2													1	
>4				1										
<=8		1												
8								1		1				
>8					1									
>32														1
64												1		
>64									1					
>128											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	7	7	7	7	7	7	7	8	7	7	7	7	7	7
N of resistant isolates	0	0	0	0	0	4	0	5	0	4	3	0	0	7
0.016						3								
<=0.03			6											
0.06			1											
<=0.25				7									3	
0.25						1								
<=0.5	7				7									
0.5						3							4	
<=1								2	4					
<=2							2					7		
2								1	3					
<=4										3				
4							4							
<=8		7												
8							1							
32											2			7
64								5						
128										4	2			
1024											3			

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	1	0	1	0	1	0	0	0	1
0.06			1											
<=0.25													1	
<=0.5	1													
0.5						1								
<=2												1		
2									1					
4				1			1							
<=8		1												
8					1									
32											1			1
64								1						
128										1				

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	0	1	0	1	1	0	0	1
MIC														
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
1						1							1	
2									1					
4												1		
8							1							
16		1												
32														1
64								1						
128										1				
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	10	10	10	10	10	10	10	10	10	10	10	10	10	10
N of resistant isolates	1	0	0	0	0	8	0	3	0	8	1	0	0	8
0.016						2								
<=0.03			10											
<=0.25				10									3	1
0.25						1								
<=0.5	9				10									
0.5						7							7	1
<=1								6	5					
<=2												10		
2								1	5					
<=4										1				
4							4							
<=8		10												
8							6			1				
16	1										1			
32								1			2			8
64								2			4			
128										8	1			
256											1			
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	1	0	1	0	1	0	0	0	1
MIC														
<=0.03			1											
0.25						1								
<=0.5	1													
0.5													1	
<=1									1					
<=2												1		
4				1			1							
<=8		1												
8					1									
32											1			1
64								1						
128										1				

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON pni2

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitores - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitores - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	CTX-M-2	CTX-M-2	CTX-M-2	CTX-M-2	CTX-M-2	CTX-M-2	CTX-M-2	CTX-M-2	CTX-M-2	CTX-M-2
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	Positive/Present	NOT AVAILABLE
Ceftazidime synergy test	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	Positive/Present	NOT AVAILABLE
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	64
N of tested isolates	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	2	2	2	2	0	0	0
MIC										
<=0.03			1							
0.03	2									
0.06			1							
0.25		1								
0.5		1						2		
1									2	
16						2	1			2
32				2			1			
64					2					

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	34	34	34	34	34	34	34	34	34	34	34	34	34	34
N of resistant isolates	0	4	0	2	2	25	4	15	0	25	23	5	0	33
MIC														
<=0.015						8								
<=0.03			33											

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	34	34	34	34	34	34	34	34	34	34	34	34	34	34
N of resistant isolates	0	4	0	2	2	25	4	15	0	25	23	5	0	33
0.03						1								
0.06			1											
<=0.25				29									16	1
0.25						1								
<=0.5	33				30									
0.5				3		14							11	
<=1								19	29					
1	1				2	7							7	
<=2												19		
2						3			5					
<=4										9				
4				2			10						10	
<=8		20												
8					2		9							
16		10					11				5			
32							2	1			3			33
64							2	14			3	5		
128		4								25				
1024											23			

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi B in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON pnl2

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitors - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitors - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
Ceftazidime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	2	2	0	2	0	0	0
MIC	0.03	2								
	0.06		2							
	0.12							2		
	0.25								1	
	0.5	2							1	
	4			2		2				
	8				1					
	16				1					2
	32						2			

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi B in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	19	19	19	19	19	19	19	19	19	19	19	19	19	19
N of resistant isolates	1	0	0	2	2	19	0	0	18	0	19	10	0	19
MIC	<=0.03		17											
	0.06		2											

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	19	19	19	19	19	19	19	19	19	19	19	19	19	19
N of resistant isolates	1	0	0	2	2	19	0	0	18	0	19	10	0	19
MIC														
<=0.25				17			5							
0.25						6								
<=0.5	18				16									
0.5						4	10							
<=1									1	15				
1					1	8	4							
<=2													12	
2						1				4				
4								8					7	
>4				2										
<=8		12												
8								6						
>8					2									
16	1	7						5						
32									1			3		
>32														19
64												3		
>64									17					
128												3		
>128											19			
>1024												10		

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi B in Gallus gallus (fowl) - broilers - during rearing period

Sampling Stage: Farm (not specified) Sampling Type: animal sample - faeces Sampling Context: Control and eradication programmes
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON pnl2
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitores - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitores - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent
Ceftazidime synergy test	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent	Negative/Absent
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	1	1	1	1	0
MIC										
<=0.03			1							
0.03	1									
0.5		1		1						
8										1
16								1		
32					1		1		1	
>64						1				

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi B in Gallus gallus (fowl) - broilers - during rearing period

Sampling Stage: Farm (not specified) Sampling Type: animal sample - faeces Sampling Context: Control and eradication programmes
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	1	1	1	0	0	1	0	1	2	0	2
MIC														
<=0.03			2											
0.03						1								
<=0.25				1			1							
0.25							1							

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	1	1	1	0	0	1	0	1	2	0	2
<=0.5	2				1									
0.5							1							
<=1									1					
<=2													2	
2										2				
<=4											1			
4								1						
>4				1										
<=8		2												
8								1						
>8					1									
>32														2
>64									1					
>128											1			
>1024												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Paratyphi B in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	1	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1					
<=2													1	
2										1				
<=4											1			
<=8		1												
8								1						
>32														1
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Poona in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=2													1	
2									1	1				
<=4											1			
4								1						
<=8		1												
>32														1
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Rissen in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Rissen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	1	1	0
MIC														
0.016						1								
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5														1
<=1									1					
2								1					1	
<=4										1				
4							1							
16		1												
64											1	1		

Table Antimicrobial susceptibility testing of Salmonella - S. Rissen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	10	10	10	10	10	10	10	10	10	10	10	10	10	10
N of resistant isolates	0	1	0	0	0	0	0	1	0	0	1	7	1	1
0.016						8								
<=0.03			9											
0.03						2								
0.12			1											
<=0.25				10									1	
<=0.5	8				10									
0.5													2	9
<=1									3					
1	2												6	
<=2												2		
2								9	7				1	
<=4										10				
4							3					1		
<=8		5												
8							6							
16		4					1							
32											1			1
64								1			7	7		
128		1									1			
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Rissen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	3	0	0
MIC														
0.016						2								
<=0.03			3											
0.03						1								
<=0.25				3										
<=0.5	3				3									
0.5														2
<=1									1					
1													3	1
2								3	2					
<=4										3				
4							2							
8							1							
16		3												
64											3	3		

Table Antimicrobial susceptibility testing of Salmonella - S. Senftenberg in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	4
MIC														
<=0.015						1								
<=0.03			5											
0.03						4								
<=0.25				5			4							
<=0.5	5				5									
0.5							1							1
<=1									4	5				
<=2								1					5	
2									1					
<=4											5			
4								2						
<=8		5												
8								2						
32												1		
>32														4
64												3		
128												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Tennessee in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.015						1								
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=2													1	
2									1	1				
<=4											1			
<=8		1												
8								1						
>32														1
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Cattle (bovine animals) - young cattle (1-2 years)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Clinical investigations

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	16	16	16	16	16	16	16	16	16	16	16	16	16	16
N of resistant isolates	0	2	0	0	0	4	0	0	12	0	4	5	7	15
<=0.015						8								
<=0.03			16											
0.03						4								
<=0.25				16			9							1
0.25						1								
<=0.5	15				16									
0.5						3	5							
<=1										9				
1	1						2							
<=2													9	
2									4	7				
<=4											11			
4								10						
<=8		14												
8								6			1			
32												9	1	
>32														15
64												2	1	
>64									12				5	
>128		2									4			
>1024												5		

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	2	0	0	1	0	2
MIC														
<=0.015						1								
<=0.03			2											
0.03						1								
<=0.25				2			1							
<=0.5	2				2									
0.5							1							
<=1										1				
<=2													2	
2										1				
<=4											2			
<=8		2												
8								2						
32												1		
>32														2
>64									2					
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Gallus gallus (fowl) - laying hens - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	1	0	0	1	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1										1				
<=2													1	
<=4											1			
<=8		1												
8								1						
>32														1
>64									1					
>1024												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified) Sampling Type: environmental sample - boot swabs Sampling Context: Control and eradication programmes
 Sampler: Industry sampling Sampling Strategy: Objective sampling Programme Code: AMR MON pnl2
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitores - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitores - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
Ceftazidime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
ECOFF	0.06	1	0.125	0.125	0.5	8	2	0.5	2	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	0	1	0	0	0
MIC	0.03	1								
	0.06		1							
	0.12						1			
	0.5	1						1		
	8			1		1				
	16				1					1
	64						1			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified) Sampling Type: environmental sample - boot swabs Sampling Context: Control and eradication programmes
 Sampler: Industry sampling Sampling Strategy: Objective sampling Programme Code: AMR MON
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13
N of resistant isolates	0	1	0	1	1	1	0	0	6	0	1	2	2	11
MIC	<=0.015					2								
	<=0.03		12											
	0.03					10								
	0.06		1											

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13
N of resistant isolates	0	1	0	1	1	1	0	0	6	0	1	2	2	11
<=0.25				12			11							2
<=0.5	11				12									
0.5						1	2							
<=1									5	7				
1	2													
<=2													11	
2									2	6				
<=4											11			
4								9						
>4				1										
<=8		12											1	
8								4			1			
>8					1									
16												3		
32												4		
>32														11
64												3	1	
>64									6				1	
>128		1									1			
>1024												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring - active

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			1											
0.06						1								
<=0.25				1										
<=0.5	1				1									
0.5														1
<=1										1				
1							1							
4									1				1	
8								1			1			
16		1												
64												1		

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Turkeys - fattening flocks (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	1	0	0	0	1	0	0	1	0	1	0	1	1
MIC														
<=0.03			1											
<=0.25				1										
<=0.5	1				1									
0.5						1								
<=1										1				
1							1							
8								1						
32											1			
>32														1
64												1		
>64									1				1	
>128		1												

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	3	3	3	3	3	3	3	3	3	3	3	3	3	3
N of resistant isolates	0	0	0	0	0	0	0	1	0	0	0	1	1	0
MIC														
0.016						1								
<=0.03			3											
0.03						2								
<=0.25				3									1	2
<=0.5	3				3									
0.5													1	1
<=1								2						
<=2												1		
2									3				1	
<=4										2				
4							2						1	
<=8		3												
8							1			1				
16											1			
32											2	1		
64								1						

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	39	39	39	39	39	39	39	39	39	39	39	39	39	39
N of resistant isolates	0	3	0	0	0	0	0	25	2	0	13	10	2	3
MIC														
0.016						8								
<=0.03			38											
0.03						28								
0.06			1			3								
<=0.25				38									16	26
<=0.5	32				39									
0.5				1									19	10
<=1								9	13					
1	7												2	
<=2							1					23		
2								5	24				2	
<=4										33				
4							22		2			6		
<=8		29												
8							14			5				
16		7					2	1		1	3			
32											13	2		3
64								24			9	8		
128		3												
256											1			
1024											13			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MIC														
<=0.03			1											
0.03						1								
<=0.25				1									1	
<=0.5	1				1									
0.5														1
<=1								1	1					
<=2												1		
<=4										1				
<=8		1												
8							1							
64											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Gallus gallus (fowl) - breeding flocks, unspecified (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	2	0	0	2	0	2
<=0.03			2											
0.03						2								
<=0.25				2			2							
<=0.5	2				2									
<=1										2				
<=2													2	
<=4											2			
<=8		2												
8								2						
>32														2
>64									2					
>1024												2		

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Pigs - breeding animals - raised under controlled housing conditions (not specified)

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - faeces

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	41	41	41	41	41	41	41	41	41	41	41	41	41	41
N of resistant isolates	1	7	0	0	0	2	0	1	28	6	1	21	25	37
<=0.015						15								
<=0.03			38											
0.03						20								
0.06			3			4								
<=0.25				41			25							3
0.25						1								
<=0.5	36				40									
0.5						1	11							1
<=1									6	25				
1	4				1		5							
<=2								1					13	
2									6	10				
<=4											36			
4								22	1	3			2	
<=8		31												
8								16		3	2		1	
16	1	3						1			2	2		
32												11	1	
>32														37
64		1										6	3	
>64								1	28				21	
128		4										1		
>128		2									1			
>1024													21	

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Meat from broilers (Gallus gallus) - carcase (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - neck skin

Sampling Context: Monitoring - EFSA specifications

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	0	1	0	1	1	0	0	1
<=0.03			1											
<=0.25				1										
<=0.5	1													
<=1									1					
1					1	1							1	
4												1		
16		1					1							
32														1
64								1						
128										1				
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
MIC														
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
4								1						
<=8		1												
16												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium var. Copenhagen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	5	6	6	6	5	5	5	6	6	5	6	5	6	5
N of resistant isolates	0	1	0	0	0	0	0	5	0	0	5	4	1	4
MIC														
<=0.03			6											
0.03						5								
<=0.25				6										
<=0.5	4				5									
0.5													3	1
<=1									4					
1	1												2	
2									2				1	
<=4										4				
4							4	1						
<=8		3												
8							1			1		1		
16		2												
32														4
64								5			1	4		
128		1												
1024											5			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium var. Copenhagen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	23	23	23	23	23	23	23	23	23	23	23	23	23	23
N of resistant isolates	0	2	0	0	0	5	0	21	1	3	22	18	1	18
MIC														
0.016						10								
<=0.03			21											
0.03						8								
0.06			2											
0.12						1								
<=0.25				22									2	3
0.25						2								
<=0.5	20				21									
0.5				1		2							9	2
<=1										12				
1	3												11	
<=2												2		
2					2			2	10					1
<=4										19				
4							10						2	
<=8		14												
8							11		1				1	
16		7					2			1				
32											1			18
64								21				18		
128		2								3				
1024											22			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium var. Copenhagen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	1	0	0	0	0	1	1	0	1	1	1	1	1	1
0.016						1								
<=0.03			1											
0.06			1											
<=0.25				2										1
<=0.5	1				2									
<=1								1	1					
1													1	
<=2							1					1		
2						1		1						
<=4											1			
<=8		2												
8													1	
16	1													
32											1			1
64							1					1		
128										1				
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium var. Copenhagen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: animal sample - caecum

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	1	0	0	1	1	0	1
MIC														
<=0.03			1											
0.06						1								
<=0.25				1										
<=0.5	1				1									
1													1	
2									1					
8							1			1				
16		1												
32														1
64								1				1		
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium var. Copenhagen in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	1	1	0	1
MIC														
0.016						1								
<=0.03			2											
0.03						1								
<=0.25				2									1	
<=0.5	1				2									
0.5													1	1
<=1									1					
1	1													
<=2												1		
2								2	1					
<=4										2				
4							2							
<=8		1												
16		1												
32											1			1
64												1		
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium, monophasic in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Processing plant

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	7	6	6	6	7	7	7	6	6	7	6	7	6	7
N of resistant isolates	0	0	0	0	0	0	0	6	1	0	6	5	0	1
MIC														
0.016						1								
<=0.03			6											
0.03						6								
<=0.25				5									4	5
<=0.5	5				6									
0.5				1									2	1
1	2				1									
<=2												1		
2									5					
<=4										6				
4							5		1			1		
<=8		6												
8							2			1				
32														1
64								6				5		
1024											6			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium, monophasic in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Slaughterhouse

Sampling Type: food sample - carcase swabs

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	40	40	40	40	40	40	40	40	40	40	40	40	40	40
N of resistant isolates	0	8	0	0	0	1	0	30	3	1	31	28	3	14
0.016						10								
<=0.03			40											
0.03						24								
0.06						5								
<=0.25				40									12	22
<=0.5	36				40									
0.5						1							18	4
<=1								6	15					
1	2												7	
<=2							4					9		
2	2							4	22				3	
<=4										24				
4							20		3			3		
<=8		23												
8							14			10				
16		9					2			5				
32										1	5			14
64		2						30			3	28		
128		6									1			
1024											31			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium, monophasic in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample (not specified)

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	1	0	0	1	2	0	0
MIC														
0.016						2								
<=0.03			1											
0.06			1											
<=0.25				2										2
<=0.5	2				2									
0.5													2	
<=1								1						
2									2					
<=4										2				
4							2							
<=8		2												
32											1			
64								1				2		
1024											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Typhimurium, monophasic in Meat, mixed meat - minced meat (not specified)

Sampling Stage: Retail

Sampling Type: food sample - meat

Sampling Context: Monitoring

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: AMR MON

Analytical Method: Micromethod dilution (in microtiter plate) (not specified)

Country of Origin: European Union

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.064	16	8	2	16	256	8	1	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	2	1	1	4	8	2	0.25	0.25
Highest limit	32	128	16	4	8	8	64	64	16	128	1024	64	8	32
N of tested isolates	6	6	6	6	6	6	6	6	6	6	6	6	6	6
N of resistant isolates	0	1	0	0	0	0	0	6	0	0	6	5	0	1
<=0.03			6											
0.03						6								
<=0.25				6									5	5
<=0.5	5				6									
<=1									5					
1													1	
2	1								1					
<=4										5				
4							1						1	
<=8		5												
8							4			1				
16							1							
32														1
64								6					5	
128		1												
1024											6			

Table Antimicrobial susceptibility testing of Salmonella - S. Virchow in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	0	0	0	0	1	0	0	1
<=0.03			1											
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
1						1								
<=2													1	
<=8		1												
16								1						
32												1		
>32														1
128											1			

Table Antimicrobial susceptibility testing of Salmonella - S. Wien in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Farm (not specified)

Sampling Type: environmental sample - boot swabs

Sampling Context: Control and eradication programmes
Programme Code: AMR MON

Sampler: Industry sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=2													1	
2									1	1				
<=4											1			
4								1						
<=8		1												
32												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. Worthington in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<=0.03			1											
0.03						1								
<=0.25				1			1							
<=0.5	1				1									
<=1									1	1				
<=2													1	
<=4											1			
<=8		1												
8								1						
16												1		
>32														1

Table Antimicrobial susceptibility testing of Salmonella - S. Yoruba in Compound feedingstuffs, not specified (not specified)

Sampling Stage: Feed mill

Sampling Type: feed sample

Sampling Context: Monitoring

Sampler: Official and industry sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.5	2	0.06	1	16	4	2	16	256	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	0	0	0	0	0	0	0	0	2
MIC														
<=0.03			2											
0.03						2								
<=0.25				2			2							
<=0.5	2				2									
<=1									2	2				
<=2													2	
<=4											2			
<=8		2												
8								2						
>32														2
64												1		
256												1		

ANTIMICROBIAL RESISTANCE TABLES FOR INDICATOR ESCHERICHIA COLI

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON pn12
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem		Carbapenems - Imipenem		Carbapenems - Meropenem		Cephalosporins - Cefepime		Cephalosporins - Cefotaxime		Cephalosporins - Cefoxitin		Cephalosporins - Ceftazidime		Cephalosporins + β lactamase inhibitors - Cefotaxime + Clavulanic acid		Cephalosporins + β lactamase inhibitors - Ceftazidime + Clavulanic acid		Penicillins - Temocillin						
ESBL genotype	NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE						
AMPC genotype	NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE						
CARBAPENEM genotype	NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE						
Cefotaxime synergy test	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent					
Ceftazidime synergy test	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent					
ECOFF	0.06	0.06	0.5	0.5	0.125	0.125	0.125	0.125	0.25	0.25	8	8	0.5	0.5	0.25	0.25	0.5	0.5	32	32					
Lowest limit	0.015	0.015	0.12	0.12	0.03	0.03	0.06	0.06	0.25	0.25	0.5	0.5	0.25	0.25	0.06	0.06	0.12	0.12	0.5	0.5					
Highest limit	2	2	16	16	32	32	64	64	64	64	128	128	128	128	64	64	128	128	128	128					
N of tested isolates	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13	13					
N of resistant isolates	0	0	0	0	0	0	11	11	13	13	5	5	12	12	5	5	5	5	0	0					
MIC																									
<=0.015	5	2																							
<=0.03					7	6																			
0.03	2	2																							
<=0.06																	5	1							
0.06																									
<=0.12																			5	1					
0.12																									
0.25							1	4																	
0.5							2	4	1	1					1										
1								1						2			1								
2											2	1	2												
4									3			2	2	1	2	1	3			4	2	3			
8											1	2	3					3	1						
16													3					2	1	1			1	1	2
32															2	1									
64																			3						

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Gallus gallus (fowl) - broilers - before slaughter

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: AMR MON
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.25	0.5	0.03	1	16	8	2	16	64	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	158	158	158	158	158	158	158	158	158	158	158	158	158	158
N of resistant isolates	9	33	0	13	11	111	0	7	115	0	100	92	72	78
MIC														
<=0.015						36								
<=0.03			155											
0.03						11								
0.06			3			1								
0.12						8								
<=0.25				145			142							45
0.25						35								
<=0.5	103				147									
0.5				2		29	16							27
<=1								3		155				
1	44				2	16								8
<=2								9					81	
2	2			3		3			19	3				
<=4											47			
4	1			2	2			53	20				4	
>4				6										
<=8		120										14		
8	2				3	10		65	1		7		1	
>8					4	9								
16	3	5						24			4	19		
32	1	8						2	1		1	31	2	
>32	2													78
64		6						2	1		13	2	21	
>64								3	113				49	
128		6									35			
>128		13									51			
>1024												92		

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year) - veal calves

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pni2
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitors - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitors - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
Ceftazidime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
ECOFF	0.06	0.5	0.125	0.125	0.25	8	0.5	0.25	0.5	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	0	1	0	0	0
MIC										
<=0.03			1							
0.03	1									
<=0.06								1		
<=0.12									1	
0.25		1								
2							1			
4				1		1				1
64					1					

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Cattle (bovine animals) - calves (under 1 year) - veal calves

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.25	0.5	0.03	1	16	8	2	16	64	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	187	187	187	187	187	187	187	187	187	187	187	187	187	187
N of resistant isolates	11	48	0	1	1	48	0	6	103	5	39	108	128	96
MIC														
<=0.015						100								
<=0.03			185											
0.03						39								
0.06			2			6								
0.12						5								
<=0.25				186			167							55
0.25						14								
<=0.5	115				186									
0.5						3	17							30
<=1								5		181				
1	59					4	3							6
<=2									12					53
2	2				1				28	1				
<=4											144			
4	1					1			61	4			5	1
>4				1										
<=8		118										26		
8	1					1			83	2	4		1	
>8						14								
16	3	21							25			34	1	
32		11							3			12	3	
>32	6													95
64		8									4	7	24	
>64									3				100	
128		3							103			1		
>128		26									25			
512												1		
>1024														106

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Cattle (bovine animals) - meat production animals - calves (under 1 year)

Sampling Stage: Farm (not specified) Sampling Type: animal sample - faeces Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pnl2
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem		Carbapenems - Imipenem		Carbapenems - Meropenem		Cephalosporins - Cefepime		Cephalosporins - Cefotaxime		Cephalosporins - Cefoxitin		Cephalosporins - Ceftazidime		Cephalosporins + β lactamase inhibitors - Cefotaxime + Clavulanic acid		Cephalosporins + β lactamase inhibitors - Ceftazidime + Clavulanic acid		Penicillins - Temocillin	
ESBL genotype	NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE	
AMPC genotype	NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE	
CARBAPENEM genotype	NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE		NOT AVAILABLE	
Cefotaxime synergy test	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent
Ceftazidime synergy test	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent	Positive/Present	Negative/Absent
ECOFF	0.06	0.06	0.5	0.5	0.125	0.125	0.125	0.125	0.25	0.25	8	8	0.5	0.5	0.25	0.25	0.5	0.5	32	32
Lowest limit	0.015	0.015	0.12	0.12	0.03	0.03	0.06	0.06	0.25	0.25	0.5	0.5	0.25	0.25	0.06	0.06	0.12	0.12	0.5	0.5
Highest limit	2	2	16	16	16	16	32	32	64	64	64	64	128	128	64	64	128	128	128	128
N of tested isolates	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4
N of resistant isolates	0	0	1	1	0	0	3	3	4	4	1	1	4	4	1	1	1	1	0	0
MIC																				
<=0.015	1				2		1													
<=0.03																				
0.03	1																			
<=0.06															1					
0.06	2				1															
<=0.12			1																2	
0.12							1										1			
0.25			1		1										1					
0.5																			1	
1			1						1				1						1	
2							1				1		1						1	
4													1						1	
8							1				2								2	
16							1		1				1						1	
32									1											
64											1									
>64									1											

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Cattle (bovine animals) - meat production animals - calves (under 1 year)

Sampling Stage: Farm (not specified) Sampling Type: animal sample - faeces Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.25	0.5	0.03	1	16	8	2	16	64	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	163	163	163	163	163	163	163	163	163	163	163	163	163	163
N of resistant isolates	8	26	0	4	4	14	0	1	33	1	12	38	29	25
MIC														
<=0.015						129								
<=0.03			160											
0.03						20								
0.06			3											
<=0.25				159			158							65
0.25						7								
<=0.5	101				159									
0.5						2	4							66
<=1									5	160				
1	50			1			1							7
<=2								16					126	
2	4				1				57	2				
<=4											149			
4					1			81	68				8	
>4				3										
<=8		136										35		
8	5				2	3		63			2			
>8						2								
16	3	1						2		1		46		
32		1										34	1	
>32														25
64		6									1	10	11	
>64								1	33				17	
128		5										6		
>128		14										5		
>1024													38	

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Pigs - fattening pigs (not specified)

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON pni2
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Carbapenems - Ertapenem	Carbapenems - Imipenem	Carbapenems - Meropenem	Cephalosporins - Cefepime	Cephalosporins - Cefotaxime	Cephalosporins - Cefoxitin	Cephalosporins - Ceftazidime	Cephalosporins + β lactamase inhibitors - Cefotaxime + Clavulanic acid	Cephalosporins + β lactamase inhibitors - Ceftazidime + Clavulanic acid	Penicillins - Temocillin
ESBL genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
AMPC genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
CARBAPENEM genotype	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE	NOT AVAILABLE
Cefotaxime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
Ceftazidime synergy test	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present	Positive/Present
ECOFF	0.06	0.5	0.125	0.125	0.25	8	0.5	0.25	0.5	32
Lowest limit	0.015	0.12	0.03	0.06	0.25	0.5	0.25	0.06	0.12	0.5
Highest limit	2	16	16	32	64	64	128	64	128	128
N of tested isolates	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	1	1	0	1	0	0	0
MIC										
<=0.015	1									
<=0.03			1							
<=0.06								1		
<=0.12		1							1	
4				1		1	1			
8										1
32					1					

Table Antimicrobial susceptibility testing of Escherichia coli, non-pathogenic - E.coli, non-pathogenic, unspecified in Pigs - fattening pigs (not specified)

Sampling Stage: Slaughterhouse Sampling Type: animal sample - caecum Sampling Context: Monitoring - active
 Sampler: Official sampling Sampling Strategy: Objective sampling Programme Code: OTHER AMR MON
 Analytical Method: Dilution - sensititre
 Country of Origin: Belgium

AM substance	Aminoglycosides - Gentamicin	Amphenicols - Chloramphenicol	Carbapenems - Meropenem	Cephalosporins - Cefotaxime	Cephalosporins - Ceftazidime	Fluoroquinolones - Ciprofloxacin	Glycylcyclines - Tigecycline	Macrolides - Azithromycin	Penicillins - Ampicillin	Polymyxins - Colistin	Quinolones - Nalidixic acid	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
ECOFF	2	16	0.125	0.25	0.5	0.03	1	16	8	2	16	64	8	2
Lowest limit	0.5	8	0.03	0.25	0.5	0.015	0.25	2	1	1	4	8	2	0.25
Highest limit	32	128	16	4	8	8	8	64	64	16	128	1024	64	32
N of tested isolates	184	184	184	184	184	184	184	184	184	184	184	184	184	184
N of resistant isolates	4	53	0	1	1	5	0	2	76	1	2	96	81	92
MIC														
<=0.015						158								
<=0.03			181											
0.03						21								
0.06			3			1								
0.12						1								
<=0.25				183			178							52
0.25						1								
<=0.5	113				183									
0.5						2	6							34
<=1								8		180				
1	63													6
<=2								20					100	
2	4				1				47	3				
<=4											178			
4	3							106	52				2	
>4				1										
<=8		127										32		
8								48	1	1	4		1	1
16	1	4						8	2			35	1	
32		25						1	1			19		
>32														91
64		16							1	5		2	25	
>64									68				55	
128		9									2			
>128		3												
256												3		
>1024												93		

OTHER ANTIMICROBIAL RESISTANCE TABLES

Table Antimicrobial susceptibility testing of Staphylococcus - S. aureus, meticillin resistant (MRSA) - MRSA, unspecified in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - nasal swab

Sampling Context: Control and eradication programmes

Sampler: Official sampling

Sampling Strategy: Objective sampling

Programme Code: OTHER AMR MON

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM Substance	Aminoglycosides - Gentamicin	Aminoglycosides - Kanamycin	Aminoglycosides - Streptomycin	Amphenicols - Chloramphenicol	Antimycobacterial drugs - Rifampicin	Cephalosporins - Cefoxitin	Fluoroquinolones - Ciprofloxacin	Fusidanes - Fusidic acid	Glycopeptides (Cyclic peptides, Polypeptides) - Vancomycin	Lincosamides - Clindamycin	Macrolides - Erythromycin	Monocarboxylic acid - Mupirocin	Oxazolidines - Linezolid	Penicillins - Penicillin
Performed CC MRSA characterisation	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Performed MLST MRSA characterisation	No	No	No	No	No	No	No	No	No	No	No	No	No	No
ECOFF	2	8	16	16	0.03	4	1	0.5	2	0.25	1	1	4	0.12
Lowest limit	1	4	4	4	0.016	0.5	0.25	0.5	1	0.12	0.25	0.5	1	0.12
Highest limit	16	64	32	64	0.5	16	8	4	16	4	8	256	8	2
N of tested isolates	5	5	5	5	5	5	5	5	5	5	5	5	5	5
N of resistant isolates	1	5	5	5	5	5	0	1	0	1	5	1	0	5
<=0.12										3				
<=0.25							4							
0.25										1				
<=0.5								4				4		
0.5							1							
>0.5					5									
<=1	4								5				2	
2								1					3	
>2														5
>4										1				
8	1													
>8											5			
>16						5								
>32			5											
64				4										
>64		5		1										
256												1		

Table Antimicrobial susceptibility testing of Staphylococcus - S. aureus, meticillin resistant (MRSA) - MRSA, unspecified in Gallus gallus (fowl) - laying hens - adult - CONTINUED

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - nasal swab

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Pleuromutilins - Tiamulin	Streptogramins - Quinupristin/Dalfo- pristin	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
Performed CC MRSA characterisation	Yes	Yes	Yes	Yes	Yes
Performed MLST MRSA characterisation	No	No	No	No	No
ECOFF	2	1	128	1	2
Lowest limit	0.5	0.5	64	0.5	2
Highest limit	4	4	512	16	32
N of tested isolates	5	5	5	5	5
N of resistant isolates	1	1	5	5	0
MIC					
<=0.5	3	4			
1	1				
<=2					5
>4	1	1			
>16				5	
>512			5		

Table Antimicrobial susceptibility testing of Staphylococcus - S. aureus, meticillin resistant (MRSA) - spa-type t011 - CC398 in Gallus gallus (fowl) - laying hens - adult

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - nasal swab

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM Substance	Aminoglycosides - Gentamicin	Aminoglycosides - Kanamycin	Aminoglycosides - Streptomycin	Amphenicols - Chloramphenicol	Antimycobacterial drugs - Rifampicin	Cephalosporins - Cefoxitin	Fluoroquinolones - Ciprofloxacin	Fusidanes - Fusidic acid	Glycopeptides (Cyclic peptides, Polypeptides) - Vancomycin	Lincosamides - Clindamycin	Macrolides - Erythromycin	Monocarboxylic acid - Mupirocin	Oxazolidines - Linezolid	Penicillins - Penicillin
Performed CC MRSA characterisation	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Performed MLST MRSA characterisation	No	No	No	No	No	No	No	No	No	No	No	No	No	No
ECOFF	2	8	16	16	0.03	4	1	0.5	2	0.25	1	1	4	0.12
Lowest limit	1	4	4	4	0.016	0.5	0.25	0.5	1	0.12	0.25	0.5	1	0.12
Highest limit	16	64	32	64	0.5	16	8	4	16	4	8	256	8	2
N of tested isolates	2	2	2	2	2	2	2	2	2	2	2	2	2	2
N of resistant isolates	0	0	0	0	0	2	1	1	0	2	1	0	0	2
MIC														
<=0.016					2									
<=0.25										1				
<=0.5								1				2		
0.5							1			1				
<=1	2								2				1	
2													1	
>2														2
<=4		2	2	1										
>4								1		1				
8				1		1	1							
>8											1			
16						1								

Table Antimicrobial susceptibility testing of Staphylococcus - S. aureus, meticillin resistant (MRSA) - spa-type t011 - CC398 in Gallus gallus (fowl) - laying hens - adult - CONTINUED

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - nasal swab

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Pleuromutilins - Tiamulin	Streptogramins - Quinupristin/Dalfo pristin	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
Performed CC MRSA characterisation	Yes	Yes	Yes	Yes	Yes
Performed MLST MRSA characterisation	No	No	No	No	No
ECOFF	2	1	128	1	2
Lowest limit	0.5	0.5	64	0.5	2
Highest limit	4	4	512	16	32
N of tested isolates	2	2	2	2	2
N of resistant isolates	1	0	0	1	2
MIC					
<=0.5	1			1	
1		2			
4					1
>4	1				
>16				1	
>32					1
<=64			1		
128			1		

Table Antimicrobial susceptibility testing of Staphylococcus - S. aureus, meticillin resistant (MRSA) - spa-type t1985 - CC398 in Gallus gallus (fowl) - breeding flocks for broiler production line - during rearing period

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - nasal swab

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM Substance	Aminoglycosides - Gentamicin	Aminoglycosides - Kanamycin	Aminoglycosides - Streptomycin	Amphenicols - Chloramphenicol	Antimycobacterial drugs - Rifampicin	Cephalosporins - Cefoxitin	Fluoroquinolones - Ciprofloxacin	Fusidanes - Fusidic acid	Glycopeptides (Cyclic peptides, Polypeptides) - Vancomycin	Lincosamides - Clindamycin	Macrolides - Erythromycin	Monocarboxylic acid - Mupirocin	Oxazolidines - Linezolid	Penicillins - Penicillin
Performed CC MRSA characterisation	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Performed MLST MRSA characterisation	No	No	No	No	No	No	No	No	No	No	No	No	No	No
ECOFF	2	8	16	16	0.03	4	1	0.5	2	0.25	1	1	4	0.12
Lowest limit	1	4	4	4	0.016	0.5	0.25	0.5	1	0.12	0.25	0.5	1	0.12
Highest limit	16	64	32	64	0.5	16	8	4	16	4	8	256	8	2
N of tested isolates	1	1	1	1	1	1	1	1	1	1	1	1	1	1
N of resistant isolates	0	0	0	0	0	1	1	0	0	1	1	0	0	1
MIC														
<=0.016					1									
<=0.5								1				1		
<=1	1								1				1	
2							1							
>2														1
<=4		1	1											
>4										1				
8				1		1								
>8											1			

Table Antimicrobial susceptibility testing of Staphylococcus - S. aureus, meticillin resistant (MRSA) - spa-type t1985 - CC398 in Gallus gallus (fowl) - breeding flocks for broiler production line - during rearing period - CONTINUED

Sampling Stage: Farm (not specified)

Sampling Type: animal sample - nasal swab

Sampling Context: Control and eradication programmes
Programme Code: OTHER AMR MON

Sampler: Official sampling

Sampling Strategy: Objective sampling

Analytical Method: Dilution - sensititre

Country of Origin: Belgium

AM substance	Pleuromutilins - Tiamulin	Streptogramins - Quinupristin/Dalfo pristin	Sulfonamides - Sulfamethoxazole	Tetracyclines - Tetracycline	Trimethoprim
Performed CC MRSA characterisation	Yes	Yes	Yes	Yes	Yes
Performed MLST MRSA characterisation	No	No	No	No	No
ECOFF	2	1	128	1	2
Lowest limit	0.5	0.5	64	0.5	2
Highest limit	4	4	512	16	32
N of tested isolates	1	1	1	1	1
N of resistant isolates	0	0	0	1	1
MIC					
<=0.5	1				
1		1			
>16				1	
>32					1
<=64			1		